

African Journal of Microbiology Research

Volume 7 Number 39 27 September 2013

ISSN 1996-0808



*Academic
Journals*

ABOUT AJMR

The **African Journal of Microbiology Research (AJMR)** (ISSN 1996-0808) is published Weekly (one volume per year) by Academic Journals.

African Journal of Microbiology Research (AJMR) provides rapid publication (weekly) of articles in all areas of Microbiology such as: Environmental Microbiology, Clinical Microbiology, Immunology, Virology, Bacteriology, Phycology, Mycology and Parasitology, Protozoology, Microbial Ecology, Probiotics and Prebiotics, Molecular Microbiology, Biotechnology, Food Microbiology, Industrial Microbiology, Cell Physiology, Environmental Biotechnology, Genetics, Enzymology, Molecular and Cellular Biology, Plant Pathology, Entomology, Biomedical Sciences, Botany and Plant Sciences, Soil and Environmental Sciences, Zoology, Endocrinology, Toxicology. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles are peer-reviewed.

Submission of Manuscript

Please read the **Instructions for Authors** before submitting your manuscript. The manuscript files should be given the last name of the first author

[Click here to Submit manuscripts online](#)

If you have any difficulty using the online submission system, kindly submit via this email ajmr@academicjournals.org.

With questions or concerns, please contact the Editorial Office at ajmr@academicjournals.org.

Editors

Prof. Dr. Stefan Schmidt,

*Applied and Environmental Microbiology
School of Biochemistry, Genetics and Microbiology
University of KwaZulu-Natal
Private Bag X01
Scottsville, Pietermaritzburg 3209
South Africa.*

Prof. Fukai Bao

*Department of Microbiology and Immunology
Kunming Medical University
Kunming 650031,
China*

Dr. Jianfeng Wu

*Dept. of Environmental Health Sciences,
School of Public Health,
University of Michigan
USA*

Dr. Ahmet Yilmaz Coban

*OMU Medical School,
Department of Medical Microbiology,
Samsun,
Turkey*

Dr. Seyed Davar Siadat

*Pasteur Institute of Iran,
Pasteur Square, Pasteur Avenue,
Tehran,
Iran.*

Dr. J. Stefan Rokem

*The Hebrew University of Jerusalem
Department of Microbiology and Molecular Genetics,
P.O.B. 12272, IL-91120 Jerusalem,
Israel*

Prof. Long-Liu Lin

*National Chiayi University
300 Syuefu Road,
Chiayi,
Taiwan*

N. John Tonukari, Ph.D

*Department of Biochemistry
Delta State University
PMB 1
Abraka, Nigeria*

Dr. Thaddeus Ezeji

*Assistant Professor
Fermentation and Biotechnology Unit
Department of Animal Sciences
The Ohio State University
1680 Madison Avenue
USA.*

Associate Editors

Dr. Mamadou Gueye

*MIRCEN/ Laboratoire commun de microbiologie
IRD-ISRA-UCAD, BP 1386,
DAKAR, Senegal.*

Dr. Caroline Mary Knox

*Department of Biochemistry, Microbiology and
Biotechnology
Rhodes University
Grahamstown 6140
South Africa.*

Dr. Hesham Elsayed Mostafa

*Genetic Engineering and Biotechnology Research
Institute (GEBRI)
Mubarak City For Scientific Research,
Research Area, New Borg El-Arab City,
Post Code 21934, Alexandria, Egypt.*

Dr. Wael Abbas El-Naggar

*Head of Microbiology Department,
Faculty of Pharmacy,
Mansoura University,
Mansoura 35516, Egypt.*

Dr. Abdel Nasser A. El-Moghazy

*Microbiology, Molecular Biology, Genetics Engineering
and Biotechnology
Dept of Microbiology and Immunology
Faculty of Pharmacy
Al-Azhar University
Nasr city,
Cairo, Egypt*

Editorial Board

Dr. Barakat S.M. Mahmoud

*Food Safety/Microbiology
Experimental Seafood Processing Laboratory
Costal Research and Extension Center
Mississippi State University
3411 Frederic Street
Pascagoula, MS 39567
USA*

Prof. Mohamed Mahrous Amer

*Poultry Disease (Viral Diseases of poultry)
Faculty of Veterinary Medicine,
Department of Poultry Diseases
Cairo university
Giza, Egypt*

Dr. Xiaohui Zhou

*Molecular Microbiology, Industrial Microbiology,
Environmental Microbiology, Pathogenesis, Antibiotic
resistance, Microbial Ecology
Washington State University
Bustad Hall 402 Department of Veterinary
Microbiology and Pathology, Pullman,
USA*

Dr. R. Balaji Raja

*Department of Biotechnology,
School of Bioengineering,
SRM University,
Chennai
India*

Dr. Aly E Abo-Amer

*Division of Microbiology, Botany Department, Faculty
of Science, Sohag University.
Egypt.*

Dr. Haoyu Mao

*Department of Molecular Genetics and Microbiology
College of Medicine
University of Florida
Florida, Gainesville
USA.*

Dr. Rachna Chandra

*Environmental Impact Assessment Division
Environmental Sciences
Sálim Ali Center for Ornithology and Natural History
(SACON),
Anaikatty (PO), Coimbatore-641108, India*

Dr. Yongxu Sun

*Department of Medicinal Chemistry and
Biomacromolecules
Qiqihar Medical University, Qiqihar 161006
Heilongjiang Province
P.R. China*

Dr. Ramesh Chand Kasana

*Institute of Himalayan Bioresource Technology
Palampur, Distt. Kangra (HP),
India*

Dr. S. Meena Kumari

*Department of Biosciences
Faculty of Science
University of Mauritius
Reduit*

Dr. T. Ramesh

*Assistant Professor
Marine Microbiology
CAS in Marine Biology
Faculty of Marine Sciences
Annamalai University
Parangipettai - 608 502
Cuddalore Dist. Tamilnadu,
India*

Dr. Pagano Marcela Claudia

*Post doctoral fellowship at Department of Biology,
Federal University of Ceará - UFC,
Brazil.*

Dr. EL-Sayed E. Habib

*Associate Professor,
Dept. of Microbiology,
Faculty of Pharmacy,
Mansoura University,
Egypt.*

Dr. Pongsak Rattanachaikunsopon

*Department of Biological Science,
Faculty of Science,
Ubon Ratchathani University,
Warin Chamrap, Ubon Ratchathani 34190,
Thailand*

Dr. Gokul Shankar Sabesan

*Microbiology Unit, Faculty of Medicine,
AIMST University
Jalan Bedong, Semeling 08100,
Kedah,
Malaysia*

Dr. Kwang Young Song

*Department of Biological Engineering,
School of Biological and Chemical Engineering,
Yanbian University of Science and Technology,
Yanji,
China.*

Dr. Kamel Belhamel

*Faculty of Technology,
University of Bejaia
Algeria*

Dr. Sladjana Jevremovic

*Institute for Biological Research
Sinisa Stankovic,
Belgrade,
Serbia*

Dr. Tamer Edirne

*Dept. of Family Medicine, Univ. of Pamukkale
Turkey*

Dr. R. Balaji Raja M.Tech (Ph.D)

*Assistant Professor,
Department of Biotechnology,
School of Bioengineering,
SRM University,
Chennai.
India*

Dr. Minglei Wang

University of Illinois at Urbana-Champaign, USA

Dr. Mohd Fuat ABD Razak

*Institute for Medical Research
Malaysia*

Dr. Davide Pacifico

*Istituto di Virologia Vegetale – CNR
Italy*

Prof. Dr. Akrum Hamdy

*Faculty of Agriculture, Minia University, Egypt
Egypt*

Dr. Ntobeko A. B. Ntusi

*Cardiac Clinic, Department of Medicine,
University of Cape Town and
Department of Cardiovascular Medicine,
University of Oxford
South Africa and
United Kingdom*

Prof. N. S. Alzoreky

*Food Science & Nutrition Department,
College of Agricultural Sciences & Food,
King Faisal University,
Saudi Arabia*

Dr. Chen Ding

*College of Material Science and Engineering,
Hunan University,
China*

Dr Svetlana Nikolić

*Faculty of Technology and Metallurgy,
University of Belgrade,
Serbia*

Dr. Sivakumar Swaminathan

*Department of Agronomy,
College of Agriculture and Life Sciences,
Iowa State University,
Ames, Iowa 50011
USA*

Dr. Alfredo J. Anceno

*School of Environment, Resources and Development
(SERD),
Asian Institute of Technology,
Thailand*

Dr. Iqbal Ahmad

*Aligarh Muslim University,
Aligarh
India*

Dr. Josephine Nketsia-Tabiri

*Ghana Atomic Energy Commission
Ghana*

Dr. Juliane Elisa Welke

*UFRGS – Universidade Federal do Rio
Grande do Sul
Brazil*

Dr. Mohammad Nazrul Islam

*NIMR; IPH-Bangalore & NIUM
Bangladesh*

Dr. Okonko, Iheanyi Omezuruike

*Department of Virology,
Faculty of Basic Medical Sciences,
College of Medicine,
University of Ibadan,
University College Hospital,
Ibadan,
Nigeria*

Dr. Giuliana Noratto

*Texas A&M University
USA*

Dr. Phanikanth Venkata Turlapati

*Washington State University
USA*

Dr. Khaleel I. Z. Jawasreh

*National Centre for Agricultural Research and
Extension, NCARE
Jordan*

Dr. Babak Mostafazadeh, MD

*Shaheed Beheshti University of Medical Sciences
Iran*

Dr. S. Meena Kumari

*Department of Biosciences
Faculty of Science
University of Mauritius
Reduit
Mauritius*

Dr. S. Anju

*Department of Biotechnology,
SRM University, Chennai-603203
India*

Dr. Mustafa Maroufpor

Iran

Prof. Dong Zhichun

*Professor, Department of Animal Sciences and
Veterinary Medicine,
Yunnan Agriculture University,
China*

Dr. Mehdi Azami

*Parasitology & Mycology Dept,
Baghaeei Lab.,
Shams Abadi St.
Isfahan
Iran*

Dr. Anderson de Souza Sant'Ana

*University of São Paulo.
Brazil.*

Dr. Juliane Elisa Welke

*UFRGS – Universidade Federal do Rio Grande do Sul
Brazil*

Dr. Paul Shapshak

*USF Health,
Depts. Medicine (Div. Infect. Disease & Internat Med)
and Psychiatry & Beh Med.
USA*

Dr. Jorge Reinheimer

*Universidad Nacional del Litoral (Santa Fe)
Argentina*

Dr. Qin Liu

*East China University of Science
and Technology
China*

Dr. Xiao-Qing Hu

*State Key Lab of Food Science and Technology
Jiangnan University
P. R. China*

Prof. Branislava Kocic

*Specialist of Microbiology and Parasitology
University of Nis, School of Medicine Institute
for Public Health Nis, Bul. Z. Djindjica 50, 18000 Nis
Serbia*

Dr. Rafel Socias

*CITA de Aragón,
Spain*

Prof. Kamal I. Mohamed

*State University of New York at Oswego
USA*

Dr. Adriano Cruz

*Faculty of Food Engineering-FEA
University of Campinas (UNICAMP)
Brazil*

Dr. Mike Agenbag (Michael Hermanus Albertus)

*Manager Municipal Health Services,
Joe Gqabi District Municipality
South Africa*

Dr. D. V. L. Sarada

*Department of Biotechnology,
SRM University, Chennai-603203
India.*

Dr. Samuel K Ameyaw

*Civista Medical Center
United States of America*

Prof. Huaizhi Wang

*Institute of Hepatopancreatobiliary
Surgery of PLA Southwest Hospital,
Third Military Medical University
Chongqing400038
P. R. China*

Prof. Bakhiet AO

*College of Veterinary Medicine, Sudan
University of Science and Technology
Sudan*

Dr. Saba F. Hussain

*Community, Orthodontics and Paediatric Dentistry
Department
Faculty of Dentistry
Universiti Teknologi MARA
40450 Shah Alam, Selangor
Malaysia*

Prof. Dr. Zohair I.F.Rahemo

*State Key Lab of Food Science and Technology
Jiangnan University
P. R. China*

Dr. Afework Kassu

*University of Gondar
Ethiopia*

Prof. Isidro A. T. Savillo

*ISCOF
Philippines*

Dr. How-Yee Lai

*Taylor's University College
Malaysia*

Dr. Nidheesh Dadheech

*MS. University of Baroda, Vadodara, Gujarat, India.
India*

Dr. Omitoyin Siyanbola

*Bowen University,
Iwo
Nigeria*

Dr. Franco Mutinelli

*Istituto Zooprofilattico Sperimentale delle Venezie
Italy*

Dr. Chanpen Chanchao

*Department of Biology,
Faculty of Science,
Chulalongkorn University
Thailand*

Dr. Tsuyoshi Kasama

*Division of Rheumatology,
Showa University
Japan*

Dr. Kuender D. Yang, MD.

*Chang Gung Memorial Hospital
Taiwan*

Dr. Liane Raluca Stan

*University Politehnica of Bucharest,
Department of Organic Chemistry "C.Nenitzescu"
Romania*

Dr. Muhamed Osman

*Senior Lecturer of Pathology & Consultant
Immunopathologist
Department of Pathology,
Faculty of Medicine,
Universiti Teknologi MARA,
40450 Shah Alam, Selangor
Malaysia*

Dr. Mohammad Feizabadi

*Tehran University of medical Sciences
Iran*

Prof. Ahmed H Mitwalli

*State Key Lab of Food Science and Technology
Jiangnan University
P. R. China*

Dr. Mazyar Yazdani

*Department of Biology,
University of Oslo,
Blindern,
Oslo,
Norway*

Dr. Ms. Jemimah Gesare Onsare

*Ministry of Higher, Education
Science and Technology
Kenya*

Dr. Babak Khalili Hadad

*Department of Biological Sciences,
Roudehen Branch,
Islamic Azad University,
Roudehen
Iran*

Dr. Ehsan Sari

*Department of Plant Pathology,
Iranian Research Institute of Plant Protection,
Tehran,
Iran.*

Dr. Snjezana Zidovec Lepej

*University Hospital for Infectious Diseases
Zagreb,
Croatia*

Dr. Dilshad Ahmad

*King Saud University
Saudi Arabia*

Dr. Adriano Gomes da Cruz

*University of Campinas (UNICAMP)
Brazil*

Dr. Hsin-Mei Ku

*Agronomy Dept. NCHU 250 Kuo
Kuang Rd, Taichung,
Taiwan*

Dr. Fereshteh Naderi

*Physical chemist,
Islamic Azad University,
Shahre Ghods Branch
Iran*

Dr. Adibe Maxwell Ogochukwu

*Department of Clinical Pharmacy and Pharmacy
Management,
University of Nigeria,
Nsukka.
Nigeria*

Dr. William M. Shafer

*Emory University School of Medicine
USA*

Dr. Michelle Bull

*CSIRO Food and Nutritional Sciences
Australia*

Prof. Dr. Márcio Garcia Ribeiro (DVM, PhD)

*School of Veterinary Medicine and Animal Science-
UNESP,
Dept. Veterinary Hygiene and Public Health,
State of Sao Paulo
Brazil*

Prof. Dr. Sheila Nathan

*National University of Malaysia (UKM)
Malaysia*

Prof. Ebiamadon Andi Brisibe

*University of Calabar,
Calabar,
Nigeria*

Dr. Julie Wang

*Burnet Institute
Australia*

Dr. Jean-Marc Chobert

*INRA- BIA, FIPL
France*

Dr. Zhilong Yang, PhD

*Laboratory of Viral Diseases
National Institute of Allergy and Infectious Diseases,
National Institutes of Health*

Dr. Dele Raheem

*University of Helsinki
Finland*

Dr. Li Sun

*PLA Centre for the treatment of infectious diseases,
Tangdu Hospital,
Fourth Military Medical University
China*

Dr. Biljana Miljkovic-Selimovic

*School of Medicine,
University in Nis,
Serbia; Referent laboratory for Campylobacter and
Helicobacter,
Center for Microbiology,
Institute for Public Health, Nis
Serbia*

Dr. Xinan Jiao

*Yangzhou University
China*

Dr. Endang Sri Lestari, MD.

*Department of Clinical Microbiology,
Medical Faculty,
Diponegoro University/Dr. Kariadi Teaching Hospital,
Semarang
Indonesia*

Dr. Hojin Shin

*Pusan National University Hospital
South Korea*

Dr. Yi Wang

*Center for Vector Biology, 180 Jones Avenue
Rutgers University, New Brunswick, NJ 08901-8536
USA*

Dr. Heping Zhang

*The Key Laboratory of Dairy Biotechnology and
Engineering,
Ministry of Education,
Inner Mongolia Agricultural University.
China*

Prof. Natasha Potgieter

*University of Venda
South Africa*

Dr. Alemzadeh

*Sharif University
Iran*

Dr. Sonia Arriaga

*Instituto Potosino de Investigación Científicay
Tecnológica/División de Ciencias Ambientales
Mexico*

Dr. Armando Gonzalez-Sanchez

*Universidad Autonoma Metropolitana Cuajimalpa
Mexico*

Dr. Pradeep Parihar

*Lovely Professional University, Phagwara, Punjab.
India*

Dr. William H Roldán

*Department of Medical Microbiology,
Faculty of Medicine,
Peru*

Dr. Kanzaki, L I B

*Laboratory of Bioprospection. University of Brasilia
Brazil*

Prof. Philippe Dorchies

*Laboratory of Bioprospection. University of Brasilia
Brazil*

Dr. C. Ganesh Kumar

*Indian Institute of Chemical Technology,
Hyderabad
India*

Dr. Farid Che Ghazali

*Universiti Sains Malaysia (USM)
Malaysia*

Dr. Samira Bouhdid

*Abdelmalek Essaadi University,
Tetouan,
Morocco*

Dr. Zainab Z. Ismail

*Department of Environmental Engineering, University
of Baghdad.
Iraq*

Dr. Ary Fernandes Junior

*Universidade Estadual Paulista (UNESP)
Brasil*

Dr. Papaevangelou Vassiliki

*Athens University Medical School
Greece*

Dr. Fangyou Yu

*The first Affiliated Hospital of Wenzhou Medical
College
China*

Dr. Galba Maria de Campos Takaki

*Catholic University of Pernambuco
Brazil*

Dr. Kwabena Ofori-Kwakye

*Department of Pharmaceutics,
Kwame Nkrumah University of Science & Technology,
KUMASI
Ghana*

Prof. Dr. Liesel Brenda Gende

*Arthropods Laboratory, School of Natural and Exact
Sciences, National University of Mar del Plata
Buenos Aires,
Argentina.*

Dr. Adeshina Gbonjubola

*Ahmadu Bello University,
Zaria.
Nigeria*

Prof. Dr. Stylianos Chatzipanagiotou

*University of Athens – Medical School
Greece*

Dr. Dongqing BAI

*Department of Fishery Science,
Tianjin Agricultural College,
Tianjin 300384
P. R. China*

Dr. Dingqiang Lu

*Nanjing University of Technology
P.R. China*

Dr. L. B. Sukla

*Scientist –G & Head, Biominerals Department,
IMMT, Bhubaneswar
India*

Dr. Hakan Parlakpınar

*MD. Inonu University, Medical Faculty, Department
of Pharmacology, Malatya
Turkey*

Dr Pak-Lam Yu

*Massey University
New Zealand*

Dr Percy Chimwamurombe

*University of Namibia
Namibia*

Dr. Euclésio Simionatto

*State University of Mato Grosso do Sul-UEMS
Brazil*

Dr. Hans-Jürg Monstein

*Clinical Microbiology, Molecular Biology Laboratory,
University Hospital, Faculty of Health Sciences, S-581
85 Linköping
Sweden*

Dr. Ajith, T. A

*Associate Professor Biochemistry, Amala Institute of
Medical Sciences, Amala Nagar, Thrissur, Kerala-680
555
India*

Dr. Feng-Chia Hsieh

*Biopesticides Division, Taiwan Agricultural Chemicals
and Toxic Substances Research Institute, Council of
Agriculture
Taiwan*

Prof. Dra. Suzan Pantaroto de Vasconcellos

*Universidade Federal de São Paulo
Rua Prof. Artur Riedel, 275 Jd. Eldorado, Diadema, SP
CEP 09972-270
Brasil*

Dr. Maria Leonor Ribeiro Casimiro Lopes Assad

*Universidade Federal de São Carlos - Centro de
Ciências Agrárias - CCA/UFSCar
Departamento de Recursos Naturais e Proteção
Ambiental
Rodovia Anhanguera, km 174 - SP-330
Araras - São Paulo
Brasil*

Dr. Pierangeli G. Vital

*Institute of Biology, College of Science, University of
the Philippines
Philippines*

Prof. Roland Ndip

*University of Fort Hare, Alice
South Africa*

Dr. Shawn Carraher

*University of Fort Hare, Alice
South Africa*

Dr. José Eduardo Marques Pessanha

*Observatório de Saúde Urbana de Belo
Horizonte/Faculdade de Medicina da Universidade
Federal de Minas Gerais
Brasil*

Dr. Yuanshu Qian

*Department of Pharmacology, Shantou University
Medical College
China*

Dr. Helen Treichel

*URI-Campus de Erechim
Brazil*

Dr. Xiao-Qing Hu

*State Key Lab of Food Science and Technology
Jiangnan University
P. R. China*

Dr. Olli H. Tuovinen

*Ohio State University, Columbus, Ohio
USA*

Prof. Stoyan Groudev

*University of Mining and Geology "Saint Ivan Rilski"
Sofia
Bulgaria*

Dr. G. Thirumurugan

*Research lab, GIET School of Pharmacy, NH-5,
Chaitanya nagar, Rajahmundry-533294.
India*

Dr. Charu Gomber

*Thapar University
India*

Dr. Jan Kuever

*Bremen Institute for Materials Testing,
Department of Microbiology,
Paul-Feller-Str. 1, 28199 Bremen
Germany*

Dr. Nicola S. Flanagan

*Universidad Javeriana, Cali
Colombia*

Dr. André Luiz C. M. de A. Santiago

*Universidade Federal Rural de Pernambuco
Brazil*

Dr. Dhruva Kumar Jha

*Microbial Ecology Laboratory,
Department of Botany,
Gauhati University,
Guwahati 781 014, Assam
India*

Dr. N Saleem Basha

*M. Pharm (Pharmaceutical Biotechnology)
Eritrea (North East Africa)*

Prof. Dr. João Lúcio de Azevedo

*Dept. Genetics-University of São Paulo-Faculty of
Agriculture- Piracicaba, 13400-970
Brasil*

Dr. Julia Inés Fariña

*PROIMI-CONICET
Argentina*

Dr. Yutaka Ito

*Kyoto University
Japan*

Dr. Cheruiyot K. Ronald

*Biomedical Laboratory Technologist
Kenya*

Prof. Dr. Ata Akcil

*S. D. University
Turkey*

Dr. Adhar Manna

*The University of South Dakota
USA*

Dr. Cícero Flávio Soares Aragão

*Federal University of Rio Grande do Norte
Brazil*

Dr. Gunnar Dahlen

*Institute of odontology, Sahlgrenska Academy at
University of Gothenburg
Sweden*

Dr. Pankaj Kumar Mishra

*Vivekananda Institute of Hill Agriculture, (I.C.A.R.),
ALMORA-263601, Uttarakhand
India*

Dr. Benjamas W. Thanomsub

*Srinakharinwirot University
Thailand*

Dr. Maria José Borrego

*National Institute of Health – Department of Infectious
Diseases
Portugal*

Dr. Catherine Carrillo

*Health Canada, Bureau of Microbial Hazards
Canada*

Dr. Marcotty Tanguy

*Institute of Tropical Medicine
Belgium*

Dr. Han-Bo Zhang

*Laboratory of Conservation and Utilization for Bio-
resources
Key Laboratory for Microbial Resources of the
Ministry of Education,
Yunnan University, Kunming 650091.
School of Life Science,
Yunnan University, Kunming,
Yunnan Province 650091.
China*

Dr. Ali Mohammed Somily

*King Saud University
Saudi Arabia*

Dr. Nicole Wolter

*National Institute for Communicable Diseases and
University of the Witwatersrand,
Johannesburg
South Africa*

Dr. Marco Antonio Nogueira

*Universidade Estadual de Londrina
CCB/Depto. De microbiologia
Laboratório de Microbiologia Ambiental
Caixa Postal 6001
86051-980 Londrina.
Brazil*

Dr. Bruno Pavoni

*Department of Environmental Sciences University of
Venice
Italy*

Dr. Shih-Chieh Lee

*Da-Yeh University
Taiwan*

Dr. Satoru Shimizu

*Horonobe Research Institute for the Subsurface
Environment,
Northern Advancement Center for Science &
Technology
Japan*

Dr. Tang Ming

*College of Forestry, Northwest A&F University,
Yangling
China*

Dr. Olga Gortzi

*Department of Food Technology, T.E.I. of Larissa
Greece*

Dr. Mark Tarnopolsky

*Mcmaster University
Canada*

Dr. Sami A. Zabin

*Al Baha University
Saudi Arabia*

Dr. Julia W. Pridgeon

*Aquatic Animal Health Research Unit, USDA, ARS
USA*

Dr. Lim Yau Yan

*Monash University Sunway Campus
Malaysia*

Prof. Rosemeire C. L. R. Pietro

*Faculdade de Ciências Farmacêuticas de Araraquara,
Univ Estadual Paulista, UNESP
Brazil*

Dr. Nazime Mercan Dogan

*PAU Faculty of Arts and Science, Denizli
Turkey*

Dr Ian Edwin Cock

*Biomolecular and Physical Sciences
Griffith University
Australia*

Prof. N K Dubey

*Banaras Hindu University
India*

Dr. S. Hemalatha

*Department of Pharmaceutics, Institute of
Technology,
Banaras Hindu University, Varanasi. 221005
India*

Dr. J. Santos Garcia A.

*Universidad A. de Nuevo Leon
Mexico India*

Dr. Somboon Tanasupawat

*Department of Biochemistry and Microbiology,
Faculty of Pharmaceutical Sciences,
Chulalongkorn University,
Bangkok 10330
Thailand*

Dr. Vivekananda Mandal

*Post Graduate Department of Botany,
Darjeeling Government College,
Darjeeling – 734101.
India*

Dr. Shihua Wang

*College of Life Sciences,
Fujian Agriculture and Forestry University
China*

Dr. Victor Manuel Fernandes Galhano

*CITAB-Centre for Research and Technology of Agro-
Environment and Biological Sciences, Integrative
Biology and Quality Research Group,
University of Trás-os-Montes and Alto Douro,
Apartado 1013, 5001-801 Vila Real
Portugal*

Dr. Maria Cristina Maldonado

*Instituto de Biotecnología. Universidad Nacional de
Tucuman
Argentina*

Dr. Alex Soltermann

*Institute for Surgical Pathology,
University Hospital Zürich
Switzerland*

Dr. Dagmara Sirova

*Department of Ecosystem Biology, Faculty Of Science,
University of South Bohemia,
Branisovska 37, Ceske Budejovice, 37001
Czech Republic*

Dr. E. O Igbinosa

*Department of Microbiology,
Ambrose Alli University,
Ekpoma, Edo State,
Nigeria.*

Dr. Hodaka Suzuki

*National Institute of Health Sciences
Japan*

Dr. Mick Bosilevac

*US Meat Animal Research Center
USA*

Dr. Nora Lía Padola

*Imunoquímica y Biotecnología- Fac Cs Vet-UNCPBA
Argentina*

Dr. Maria Madalena Vieira-Pinto

*Universidade de Trás-os-Montes e Alto Douro
Portugal*

Dr. Stefano Morandi

*CNR-Istituto di Scienze delle Produzioni Alimentari
(ISPA), Sez. Milano
Italy*

Dr Line Thorsen

*Copenhagen University, Faculty of Life Sciences
Denmark*

Dr. Ana Lucia Falavigna-Guilherme

*Universidade Estadual de Maringá
Brazil*

Dr. Baoqiang Liao

*Dept. of Chem. Eng., Lakehead University, 955 Oliver
Road, Thunder Bay, Ontario
Canada*

Dr. Ouyang Jinping

*Patho-Physiology department,
Faculty of Medicine of Wuhan University
China*

Dr. John Sorensen

*University of Manitoba
Canada*

Dr. Andrew Williams

*University of Oxford
United Kingdom*

Dr. Chi-Chiang Yang

*Chung Shan Medical University
Taiwan, R.O.C.*

Dr. Quanming Zou

*Department of Clinical Microbiology and Immunology,
College of Medical Laboratory,
Third Military Medical University
China*

Prof. Ashok Kumar

*School of Biotechnology,
Banaras Hindu University, Varanasi
India*

Dr. Chung-Ming Chen

*Department of Pediatrics, Taipei Medical University
Hospital, Taipei
Taiwan*

Dr. Jennifer Furin

*Harvard Medical School
USA*

Dr. Julia W. Pridgeon

*Aquatic Animal Health Research Unit, USDA, ARS
USA*

Dr. Alireza Seidavi

*Islamic Azad University, Rasht Branch
Iran*

Dr. Thore Rohwerder

*Helmholtz Centre for Environmental Research UFZ
Germany*

Dr. Daniela Billi

*University of Rome Tor Vergata
Italy*

Dr. Ivana Karabegovic

*Faculty of Technology, Leskovac, University of Nis
Serbia*

Dr. Flaviana Andrade Faria

*IBILCE/UNESP
Brazil*

Prof. Margareth Linde Athayde

*Federal University of Santa Maria
Brazil*

Dr. Guadalupe Virginia Nevarez Moorillon

*Universidad Autonoma de Chihuahua
Mexico*

Dr. Tatiana de Sousa Fiuza

*Federal University of Goias
Brazil*

Dr. Indrani B. Das Sarma

*Jhulelal Institute of Technology, Nagpur
India*

Dr. Guanghua Wang

*Northeast Institute of Geography and Agroecology,
Chinese Academy of Sciences
China*

Dr. Renata Vadkertiova

*Institute of Chemistry, Slovak Academy of Science
Slovakia*

Dr. Charles Hocart

*The Australian National University
Australia*

Dr. Guoqiang Zhu

*University of Yangzhou College of Veterinary Medicine
China*

Dr. Guilherme Augusto Marietto Gonçalves

*São Paulo State University
Brazil*

Dr. Mohammad Ali Faramarzi

*Tehran University of Medical Sciences
Iran*

Dr. Suppasil Maneerat

*Department of Industrial Biotechnology, Faculty of
Agro-Industry, Prince of Songkla University, Hat Yai
90112
Thailand*

Dr. Francisco Javier Las heras Vazquez

*Almeria University
Spain*

Dr. Cheng-Hsun Chiu

*Chang Gung memorial Hospital, Chang Gung
University
Taiwan*

Dr. Ajay Singh

*DDU Gorakhpur University, Gorakhpur-273009 (U.P.)
India*

Dr. Karabo Shale

*Central University of Technology, Free State
South Africa*

Dr. Lourdes Zélia Zanoni

*Department of Pediatrics, School of Medicine, Federal
University of Mato Grosso do Sul, Campo Grande,
Mato Grosso do Sul
Brazil*

Dr. Tulin Askun

*Balikesir University
Turkey*

Dr. Marija Stankovic

*Institute of Molecular Genetics and Genetic
Engineering
Republic of Serbia*

Dr. Scott Weese

*University of Guelph
Dept of Pathobiology, Ontario Veterinary College,
University of Guelph,
Guelph, Ontario, N1G2W1,
Canada*

Dr. Sabiha Essack

*School of Health Sciences
South African Committee of Health Sciences
University of KwaZulu-Natal
Private Bag X54001
Durban 4000
South Africa*

Dr. Hare Krishna

*Central Institute for Arid Horticulture,
Beechwal, Bikaner-334 006, Rajasthan,
India*

Dr. Anna Mensuali

*Dept. of Life Science,
Scuola Superiore
Sant'Anna*

Dr. Ghada Sameh Hafez Hassan

*Pharmaceutical Chemistry Department,
Faculty of Pharmacy, Mansoura University,
Egypt*

Dr. Kátia Flávia Fernandes

*Biochemistry and Molecular Biology
Universidade Federal de Goiás
Brasil*

Dr. Abdel-Hady El-Gilany

*Public Health & Community Medicine
Faculty of Medicine,
Mansoura University
Egypt*

Dr. Hongxiong Guo

*STD and HIV/AIDS Control and Prevention,
Jiangsu provincial CDC,
China*

Dr. Konstantina Tsaousi

*Life and Health Sciences,
School of Biomedical Sciences,
University of Ulster*

Dr. Bhavnaben Gowan Gordhan

*DST/NRF Centre of Excellence for Biomedical TB
Research
University of the Witwatersrand and National Health
Laboratory Service
P.O. Box 1038, Johannesburg 2000,
South Africa*

Dr. Ernest Kuchar

*Pediatric Infectious Diseases,
Wroclaw Medical University,
Wroclaw Teaching Hospital,
Poland*

Dr. Hongxiong Guo

*STD and HIV/AIDS Control and Prevention,
Jiangsu provincial CDC,
China*

Dr. Mar Rodriguez Jovita

*Food Hygiene and Safety, Faculty of Veterinary
Science.
University of Extremadura,
Spain*

Dr. Jes Gitz Holler

*Hospital Pharmacy,
Aalesund. Central Norway Pharmaceutical Trust
Professor Brochs gt. 6. 7030 Trondheim,
Norway*

Prof. Chengxiang FANG

*College of Life Sciences,
Wuhan University
Wuhan 430072, P.R.China*

Dr. Anchalee Tungtrongchitr

*Siriraj Dust Mite Center for Services and Research
Department of Parasitology,
Faculty of Medicine Siriraj Hospital,
Mahidol University
2 Prannok Road, Bangkok Noi,
Bangkok, 10700, Thailand*

Instructions for Author

Electronic submission of manuscripts is strongly encouraged, provided that the text, tables, and figures are included in a single Microsoft Word file (preferably in Arial font).

The **cover letter** should include the corresponding author's full address and telephone/fax numbers and should be in an e-mail message sent to the Editor, with the file, whose name should begin with the first author's surname, as an attachment.

Article Types

Three types of manuscripts may be submitted:

Regular articles: These should describe new and carefully confirmed findings, and experimental procedures should be given in sufficient detail for others to verify the work. The length of a full paper should be the minimum required to describe and interpret the work clearly.

Short Communications: A Short Communication is suitable for recording the results of complete small investigations or giving details of new models or hypotheses, innovative methods, techniques or apparatus. The style of main sections need not conform to that of full-length papers. Short communications are 2 to 4 printed pages (about 6 to 12 manuscript pages) in length.

Reviews: Submissions of reviews and perspectives covering topics of current interest are welcome and encouraged. Reviews should be concise and no longer than 4-6 printed pages (about 12 to 18 manuscript pages). Reviews are also peer-reviewed.

Review Process

All manuscripts are reviewed by an editor and members of the Editorial Board or qualified outside reviewers. Authors cannot nominate reviewers. Only reviewers randomly selected from our database with specialization in the subject area will be contacted to evaluate the manuscripts. The process will be blind review.

Decisions will be made as rapidly as possible, and the Journal strives to return reviewers' comments to authors as fast as possible. The editorial board will re-review manuscripts that are accepted pending revision. It is the goal of the AJMR to publish manuscripts within weeks after submission.

Regular articles

All portions of the manuscript must be typed double-spaced and all pages numbered starting from the title page.

The Title should be a brief phrase describing the contents of the paper. The Title Page should include the authors' full names and affiliations, the name of the corresponding author along with phone, fax and E-mail information. Present addresses of authors should appear as a footnote.

The Abstract should be informative and completely self-explanatory, briefly present the topic, state the scope of the experiments, indicate significant data, and point out major findings and conclusions. The Abstract should be 100 to 200 words in length. Complete sentences, active verbs, and the third person should be used, and the abstract should be written in the past tense. Standard nomenclature should be used and abbreviations should be avoided. No literature should be cited.

Following the abstract, about 3 to 10 key words that will provide indexing references should be listed.

A list of non-standard **Abbreviations** should be added. In general, non-standard abbreviations should be used only when the full term is very long and used often. Each abbreviation should be spelled out and introduced in parentheses the first time it is used in the text. Only recommended SI units should be used. Authors should use the solidus presentation (mg/ml). Standard abbreviations (such as ATP and DNA) need not be defined.

The Introduction should provide a clear statement of the problem, the relevant literature on the subject, and the proposed approach or solution. It should be understandable to colleagues from a broad range of scientific disciplines.

Materials and methods should be complete enough to allow experiments to be reproduced. However, only truly new procedures should be described in detail; previously published procedures should be cited, and important modifications of published procedures should be mentioned briefly. Capitalize trade names and include the manufacturer's name and address. Subheadings should be used. Methods in general use need not be described in detail.

Results should be presented with clarity and precision. The results should be written in the past tense when describing findings in the authors' experiments. Previously published findings should be written in the present tense. Results should be explained, but largely without referring to the literature. Discussion, speculation and detailed interpretation of data should not be included in the Results but should be put into the Discussion section.

The Discussion should interpret the findings in view of the results obtained in this and in past studies on this topic. State the conclusions in a few sentences at the end of the paper. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined.

The Acknowledgments of people, grants, funds, etc should be brief.

Tables should be kept to a minimum and be designed to be as simple as possible. Tables are to be typed double-spaced throughout, including headings and footnotes. Each table should be on a separate page, numbered consecutively in Arabic numerals and supplied with a heading and a legend. Tables should be self-explanatory without reference to the text. The details of the methods used in the experiments should preferably be described in the legend instead of in the text. The same data should not be presented in both table and graph form or repeated in the text.

Figure legends should be typed in numerical order on a separate sheet. Graphics should be prepared using applications capable of generating high resolution GIF, TIFF, JPEG or Powerpoint before pasting in the Microsoft Word manuscript file. Tables should be prepared in Microsoft Word. Use Arabic numerals to designate figures and upper case letters for their parts (Figure 1). Begin each legend with a title and include sufficient description so that the figure is understandable without reading the text of the manuscript. Information given in legends should not be repeated in the text.

References: In the text, a reference identified by means of an author's name should be followed by the date of the reference in parentheses. When there are more than two authors, only the first author's name should be mentioned, followed by 'et al'. In the event that an author cited has had two or more works published during the same year, the reference, both in the text and in the reference list, should be identified by a lower case letter like 'a' and 'b' after the date to distinguish the works.

Examples:

Abayomi (2000), Agindotan et al. (2003), (Kelebeni, 1983), (Usman and Smith, 1992), (Chege, 1998;

1987a,b; Tijani, 1993,1995), (Kumasi et al., 2001) References should be listed at the end of the paper in alphabetical order. Articles in preparation or articles submitted for publication, unpublished observations, personal communications, etc. should not be included in the reference list but should only be mentioned in the article text (e.g., A. Kingori, University of Nairobi, Kenya, personal communication). Journal names are abbreviated according to Chemical Abstracts. Authors are fully responsible for the accuracy of the references.

Examples:

Chikere CB, Omoni VT and Chikere BO (2008). Distribution of potential nosocomial pathogens in a hospital environment. *Afr. J. Biotechnol.* 7: 3535-3539.

Moran GJ, Amii RN, Abrahamian FM, Talan DA (2005). Methicillinresistant *Staphylococcus aureus* in community-acquired skin infections. *Emerg. Infect. Dis.* 11: 928-930.

Pitout JDD, Church DL, Gregson DB, Chow BL, McCracken M, Mulvey M, Laupland KB (2007). Molecular epidemiology of CTXM-producing *Escherichia coli* in the Calgary Health Region: emergence of CTX-M-15-producing isolates. *Antimicrob. Agents Chemother.* 51: 1281-1286.

Pelczar JR, Harley JP, Klein DA (1993). *Microbiology: Concepts and Applications*. McGraw-Hill Inc., New York, pp. 591-603.

Short Communications

Short Communications are limited to a maximum of two figures and one table. They should present a complete study that is more limited in scope than is found in full-length papers. The items of manuscript preparation listed above apply to Short Communications with the following differences: (1) Abstracts are limited to 100 words; (2) instead of a separate Materials and Methods section, experimental procedures may be incorporated into Figure Legends and Table footnotes; (3) Results and Discussion should be combined into a single section.

Proofs and Reprints: Electronic proofs will be sent (e-mail attachment) to the corresponding author as a PDF file. Page proofs are considered to be the final version of the manuscript. With the exception of typographical or minor clerical errors, no changes will be made in the manuscript at the proof stage.

Fees and Charges: Authors are required to pay a \$550 handling fee. Publication of an article in the African Journal of Microbiology Research is not contingent upon the author's ability to pay the charges. Neither is acceptance to pay the handling fee a guarantee that the paper will be accepted for publication. Authors may still request (in advance) that the editorial office waive some of the handling fee under special circumstances

Copyright: © 2013, Academic Journals.

All rights Reserved. In accessing this journal, you agree that you will access the contents for your own personal use but not for any commercial use. Any use and or copies of this Journal in whole or in part must include the customary bibliographic citation, including author attribution, date and article title.

Submission of a manuscript implies: that the work described has not been published before (except in the form of an abstract or as part of a published lecture, or thesis) that it is not under consideration for publication elsewhere; that if and when the manuscript is accepted for publication, the authors agree to automatic transfer of the copyright to the publisher.

Disclaimer of Warranties

In no event shall Academic Journals be liable for any special, incidental, indirect, or consequential damages of any kind arising out of or in connection with the use of the articles or other material derived from the AJMR, whether or not advised of the possibility of damage, and on any theory of liability.

This publication is provided "as is" without warranty of any kind, either expressed or implied, including, but not limited to, the implied warranties of merchantability, fitness for a particular purpose, or non-infringement. Descriptions of, or references to, products or publications does not imply endorsement of that product or publication. While every effort is made by Academic Journals to see that no inaccurate or misleading data, opinion or statements appear in this publication, they wish to make it clear that the data and opinions appearing in the articles and advertisements herein are the responsibility of the contributor or advertiser concerned. Academic Journals makes no warranty of any kind, either express or implied, regarding the quality, accuracy, availability, or validity of the data or information in this publication or of any other publication to which it may be linked.

ARTICLES

Review

- Review on bioactive potential of marine microbes** 4683
V. Vasanthabharathi and S. Jayalakshmi

Research Articles

- Siderophore mediated antagonism of fluorescent Pseudomonads against soil borne plant pathogenic fungi in West Bengal, India** 4689
A. Adhikari, K. Sarker, M. De Roy, I. Bhattacharya, T. Mandal, P. K. Dasmohapatra and S. Dutta
- Validation of the acaricidal properties of materials used in ethno-veterinary control of cattle ticks** 4701
Busani Moyo and Patrick Julius Masika
- Effect of different ferric fertilizers on planting *Morchella conica* fruiting yields and analyses of the microflora and bioactivities of its grown soil** 4707
Xue Li, Zizhu Chen, Chenchen Peng, Yu Han, Jianguang Xu, Zhangfu Long
- Analysis of an outbreak of *Klebsiella pneumoniae* by the DiversiLab system and pulsed-field gel electrophoresis** 4717
Jing Li, Zhi Dong Hu and Cui E. He
- Towards efficient crude oil degradation by *Pseudomonas* sp. strain-O2: Application of Plackett-Burman design for evaluation of cultivation conditions** 4722
Mahmoud M. Berekaa
- Real time polymerase chain reaction for identification of *Candida* species isolated from blood stream infections in non-neutropenic cancer patients** 4730
Rasha A. Alm El-Din, Hanan El-Bassat, Mohamed El-Bedewy and Mohamed A. Alm El-Din

Review

Review on bioactive potential of marine microbes

V. Vasanthabharathi* and S. Jayalakshmi

CAS in Marine Biology, Faculty of Marine Sciences, Annamalai University, Tamilnadu, India.

Accepted 26 July, 2013

There is a need for new drugs especially novel antibiotics, anticancer compound, antibiotic against multi drug-resistant bacteria. The discovery and development of new drugs from marine microbes played a significant role over the last few decades. The world oceans comprise a rich diversity of microbial life with current estimates reaching over a million different species. Marine microbes are important from both ecological and biotechnological point of view. Marine microbial bioactive compounds have attracted increasing attention from microbiologists, taxonomists, ecologists, agronomists, chemists and evolutionary biologists.

Key words: Bioactive molecules, bacteria, fungi.

INTRODUCTION

Marine microbes having immense genetic and biochemical diversity have become a rich source of novel effective drugs. It is surprising to find that many bioactive compounds, reported from marine invertebrates are produced by their microbial symbionts. Competition among microbes for space and nutrients in the marine environment is a driving force behind the production of such precious antibiotics and other useful pharmaceuticals. Interestingly, microorganisms associated with marine invertebrates are proved as valuable candidates for drug discovery program.

Like bacteria, marine fungi are also reported to be potential source of bioactive substances. For example, sorbicilactone-A, a novel type alkaloid was reported from sponge (*Ircinia fasciculata*) associated fungus, *Penicillium chrysogenum*. This compound showed therapeutic potential in human trials. Polyketide synthases (PKSs) are a class of enzymes that are involved in the biosynthesis of secondary metabolites like erythromycin (Lamela et al., 1989; Parkes et al., 1994; Hentschel et al., 2003).

Since the discovery of penicillin in 1929, a number of natural products are reported to have biological activities that are in use today as antibiotics, antitumour agents

and agrochemicals (Davidson et al., 2001). The development of the fungal metabolite, Mevinolin for the treatment of high serum cholesterol and the bacterial metabolite FK-506 as an immunosuppressant illustrate the vast natural diversity of microbial natural products (Faulkner, 1992). In spite of such successes in drug discovery from terrestrial microorganisms, marine microorganisms have received only very little attention on account of the non-culturability of the majority of marine bacteria (Gaino and Sara, 1994).

Marine toxins such as tetrodotoxin and saxitoxin are potent and specific sodium channel blockers and pharmacological studies on these toxins have played a major role in developing the concept of sodium channels in general and membrane channels in particular (Gustafson et al., 1989). There is an increasing evidence that several anti-cancer compounds isolated from sponges or other marine invertebrates are actually produced by bacteria associated with these invertebrates. Bryostatins were initially isolated from a bryozoan, *Bugula neritina*. Putative type 1 polyketide synthase genes had been found in bacteria from colonies of *B. neritina* producing bryostatin and these genes were absent in bacteria associated with colonies of *B. neritina*

*Corresponding author. E-mail: bharathigene@rediffmail.com.

that did not produce bryostatin (Holst et al., 1994). Dolastatin isolated from the Indian Ocean seahare *Dolabella auricularia* has also been isolated from marine cyanobacteria (Hugenholtz and Pace, 1996) like *Lyngbya majuscula* (Ireland, 1993).

MARINE BACTERIA AND ACTINOMYCETES

About 20% of bacteria from marine sponge and coral in different coastal areas of the China Sea showed positive antimicrobial activity (Zheng et al., 2005). In the study of Li et al. (2007), some bacteria were isolated from the sponges *Stelletta tenuis*, *Halichondria rugosa* and *Dysidea avara* which showed pronounced broad-spectrum of antimicrobial activities and enzymatic potentials. Brammavidhya and Usharani (2013) extracted and optimized the bioactive compound and it act against the human pathogen which was isolated from *Hyattella cribriformis* associated *Bacillus cereus* SBS02.

The major antimicrobial metabolite isolated from *Pseudoalteromonas piscicida* NJ6-3-1 with wide antimicrobial spectrum was identified as norharman (a beta-carboline alkaloid) (Zheng et al., 2005). Cyclo-L-Phenylalanyl-L-Proline (Cyclo-L-Ph- L-Pro) was isolated from *Alcaligenes faecalis* A72 associated with the sponge *S. tenuis* which showed moderate inhibitory activity against *Staphylococcus aureus* (Li et al., 2008). Bioassay-guided fractionation of the CHCl₃ extract of the fermentation broth of a sponge *Mycale plumose*-derived actinomycete *Saccharopolyspora* spp. nov., led to the isolation of two known prodigiosin analogs and undecyl prodigiosin. These compounds exhibited significant cytotoxic activities against cancer cell lines: P388, HL60, A-549, BEL-7402 and SPCA4 (Liu et al., 2005). Revathy et al. (2013) studied the antioxidant and enzyme inhibitory potential of *Streptomyces* sp. VITMSS05 strain, isolated from Marakkanam, southern coast of India.

Bacillamides have been proven to inhibit the growth of red-tide algae such as *Cochlodinium polykrikoides* (Jeong et al., 2003). A novel thiazole alkaloid, neobacillamide A together with a known related one, bacillamide C were isolated from *Bacillus vallismortis* C89 associated with the South China Sea sponge, *Dysidea avara* (Yu et al., 2009). Studies made by the scientists at the Scripps Institute of Oceanography showed that marine bacteria are capable of producing unusual bioactive compounds that are not observed in terrestrial sources (Fenical, 1993; Fenical and Jensen, 1993). Pandey et al. (2013) studied the diversity of beta-glucosidase inhibitors producing marine bacteria. They observed that the marine sponge *Aka coralliphaga* associated bacteria had produced beta-glucosidase inhibitors as compared to other associated microbes. Mithun and Rama (2012) isolated and identified *Micrococcus luteus* sp. from marine soil samples

collected from Bay of Bengal coast of Machilipatnam, Krishna district, Andhra, India. This strain showed promising results against cancer cell line namely HCT 15 and MES-SA.

Thermo-stable proteases, lipases, esterases and starch and xylan degrading enzymes have been actively sought and in many cases are found in bacterial and archaeal hyper thermophilic marine microorganisms (Bertoldo and Antranikian, 2002). An unusual Gram-positive bacterium from deep-sea sediment produced a series of natural products, for example macrolactin A-F of an unprecedented C₂₄ linear acetogen origin (Gustafson, 1989). The major metabolite, macrolactin A inhibited B16-F10 murine melanoma cells in *in vitro* assays, showing significant inhibition of mammalian Herpes simplex virus (type I and II) and protecting T lymphocytes against human immunodeficiency virus (HIV) replication (Carte, 1996).

On the other hand, a microbial metabolite obtained from *Alteromonas* spp. isolated from the tissues of sponge had anti HIV activity due to the production of reverse transcriptase inhibitor. Some *Vibrio* species have been found to produce a variety of extra cellular proteases. *Vibrio alginolyticus* produced six proteases including an unusual detergent-resistant, alkaline serine exoprotease, this marine bacterium also produced collagenase, an enzyme with a variety of industrial and commercial applications, including the dispersion of cells in tissue culture studies (Graham et al., 1980; Osama and Koga, 1995). Bioactive compound, Cis-sativenediol extracted from *Aeromonas* sp and *Rhodopseudomonas* sp. which was collected from coastal water of Thoothukudi, Thiruchendur and Kanyakumarai Tamil Nadu, India act against *Enterobacter aerogenes*, *Pseudomonas aeruginosa*, *Streptococcus mutans*, *Staphylococcus epidermidis* and *S. aureus* (Ashadevi et al., 2011).

Pyridinium, a compound with antimicrobial activity was isolated from marine actinomycete, *Amycolatopsis alba* var. nov. DVR D4 (Dasari et al., 2012). Actinomycetes from marine sediment have anticancer activity against breast cancer cell lines MCF-7 and MDA-MB-231 (Ravikumar et al., 2012). The cytotoxicity and antioxidant activity of 5-(2,4-dimethylbenzyl)pyrrolidin-2-one (DMBPO) extracted from marine *Streptomyces* VITSVK5 sp. from sediment samples collected from the Marakkanam coast, Bay of Bengal, India was studied (Saurav and Kannabiran, 2012). Anthracyclines namely 4,6,11-trihydroxy-9-propyltetracene-5,12-dione and 10β-carbomethoxy-7,8,9,10-tetrahydro-4,6,7α,9α,11-pentahydroxy-9-propyltetracene-5,12-dione have cytotoxic activity against the HCT-8 human colon adenocarcinoma cell line, which were isolated from *Micromonospora* sp. associated with the tunicate *Eudistoma vancouveri* (Sousa et al., 2012). Sivasankar et al. (2013) isolated and optimized the L-asparaginase from marine *Streptomyces* sp. which showed the antibacterial activity.

MARINE FUNGI

A compound, (S)-2,4-dihydroxy-1-butyl(4-hydroxy) benzoate and another compound, fructigenines A exhibiting cytotoxic activity against FT210 cells, were isolated from fungus *Penicillium auratiogriseum* derived from sponge *Mycale plumose* (Xin et al., 2005). Xie et al. (1996) demonstrated that two antifungal trichothecenes, including roridin A and roridin D produced by the fungus *Myrothecium* spp. isolated from the marine sponge *Axinella* spp. in the South China Sea, could be potential inhibitors against the plant pathogen *Sclerotinia sclerotiorum*. The sponge *Acanthella cavernosa*-associated fungus, *Letendreaea helminthicola*, produced two antifouling compounds such as 3-methyl-N-(2-phenylethyl) butanamide and carbo methoxycarbonyl -D-Pro-D-Phe benzyl ester (Yang et al., 2007).

Three quinazoline alkaloids, aurantiomides A, B, and C were isolated from *Penicillium auratiogriseum* SP0-19 associated with sponge *Mycale plumose* among which aurantiomides B and C showed moderate cytotoxicities against HL-60, P388 and BEL-7402 and P388 cell lines, respectively (Xin et al., 2007).

Beside sponges, some pharmaceutical metabolites have been isolated from marine microbial symbionts associated with sea squirts, marine bryozoans and molluscs. 5 α ,8 α -epidioxy-23-methyl-(22E,24R)-ergosta-6,22-dien-3 β -ol with cytotoxic activity was isolated from the fungus *Penicillium stoloniferum* QY2-10, associated with a sea squirt (Xin et al., 2007). Six new ergo sterols obtained from a marine *Bugula* spp. derived fungus *Rhizopus* spp. showed activities against P388 and HL-60, A549 and BEL-7402.

Seven new prenylated indole diketopiperazine alkaloids, including spirotryprostatins derivatives of fumitre-morgin B, and 13-oxoverruculogen, have been isolated from the holothurian-derived fungus *Aspergillus fumigatus*. These compounds showed anticancer activity to four cancer cell lines, MOLT-4, A549, HL-60 and BEL-7420 (Wang et al., 2008). Lasiodiplodin, which could inhibit the *in vitro* growth of *S. aureus*, *Bacillus subtilis* and *Fusarium oxysporum*, were isolated from the mycelium extracts of an endophytic fungus obtained from the brown algae species collected from Zhanjiang sea area by Yang et al. (2006).

When compared with the small molecular metabolites of marine microbial symbionts, the investigation of pharmaceutical big molecule metabolites is rare. The gene cloning, purification, properties, kinetics and antifungal activity of chitinase from marine *Streptomyces* spp. DA11 associated with the South China Sea sponge *Craniella australiensis* were investigated by Han et al. (2009).

Association of fungi with marine animals ranges from saprotrophic, symbiotic to parasitic. Saprotrophic fungi have been isolated from the surface, guts and fluids of holothurians or the sea cucumbers (Pivkin, 2000).

Several fungi isolated from invertebrates have been found to produce interesting secondary metabolites. A novel group of platelet activating factor (PAF) antagonists, phomactins A, B, B1 and B2 were isolated from culture broth of *Phoma* isolated from the shell of crab *Chionoecetes opilio* collected from the coast of Fukui, Japan (Sugano et al., 1991). Two lipophilic tripeptides from *Penicillium fellutanum* live inside the gastrointestinal tract of a marine fish and three quinazoline derivatives from *Aspergillus fumigatus* from gastrointestinal tract of the fish, *Pseudolabrus japonicas* from the coast of Japan (Kobayashi and Ishibashi, 1993). An endolithic fungus *Ostracoblabe implexa* living inside the shells of rock oyster *Crassostrea cucullata* from the coast of Goa was reported (Raghukumar and Lande, 1988). Fungi belonging to the class of Trichomycetes are found in the guts of marine arthropods, isopods, decapods and amphipods in a symbiotic association (Misra and Lichtwardt, 2000). Endolithic fungi in coral skeleton are common (Alsumard et al., 1995; Ravindran et al., 2001; Golubic et al., 2005).

A basidiomycetous yeast *Cryptococcus* spp. associated with *Pocillopora damicornis* coral skeleton was shown to produce a transient cryoprotective effect, selectively enhancing the survival of skeletogenic cell types (Coulon et al., 2004) - which gave some detailed characteristic of this cryoprotective effect, some values would be very interesting. *Colecobasidium* species was isolated from necrotic patches in several massive corals in the Andamans (Raghukumar and Raghukumar, 1991).

Biological activities of microbes are mainly focused in the areas of antibiotic and anticancer properties, but other selective activities include cell cycle inhibition, antagonism of platelet activating factor, antiviral activity, neuritogenic activity, phosphatase inhibition and kinase inhibition and radical scavenging activities are less attended (Bugni and Ireland, 2004). Among the eukaryotic microscopic fungi, the inhibitory compounds capability of imperfect fungi, the ascomycetes and several other filamentous and endophytic fungal species are the most significant (Shearer and Maivan, 1988). Several compounds that inhibit the growth of a large spectrum of saprophytic and phytopathogenic fungi were isolated from basidiomycetes (Anke, 1989, 1995).

A marine-derived *Penicillium chrysogenum* isolated from the sponge, *Ircinia fasciculata* produced a sorbicillin derived alkaloid sorbicillacton A (Bringmann et al., 2003) which showed selective cytostatic activity against lympho blasts and was able to protect human T cells against HIV-1. Preclinical evaluation of this compound has recently been initiated. Many of these fungi have novel carbon skeletons thus providing further evidence of the potential of marine-derived fungi (Proksch et al., 2003).

The first crystalline fungal product obtained from *Penicillium glaucoma* was chemically myco phenolic acid and it was discovered in 1896 (Berdy, 2005). Microbial

secondary metabolites regulate growth processes, replications, and/or exhibited some kind of regulatory, inhibitory or stimulatory responses in prokaryotic and eukaryotic cells even at a minimal concentration. Marine-derived isolate of *Aspergillus versicolor* (MST-MF495) yielded many known fungal metabolites like sterigmatocystin, violaceol I, violaceol II, diorcinol, (-) cyclophenol and viridicatol, along with a new alkaloid, cottoquinazoline A, and two new cyclopentapeptides, cotteslosins A and B (Fremlin et al., 2009). Moreover, the great practical and historical importance of beta-lactams (penicillins, cephalosporins), the cyclosporin, and various statins (mevinolin, compactin, lovastatin, pravastatin, atorvastatin) which are all fungus derived compounds are well known (Donadio et al., 2002).

Several compounds from marine fungi have been screened for antifungal activities, and a number of compounds have been characterized with regard to their inhibitory activities and chemical structures. There has been a sharp increase in fungal infections among patients suffering from HIV, receiving cancer and immune-therapy (Anaissie, 1992; Li et al., 1998; Barrett, 2002). Recent research showed that compounds like Hypoxysordarin, a new sordarin derivative, isolated from the facultative marine fungus *Hypoxylon croceum* are promising.

Terrestrial fungi produce a variety of chemically diverse natural compounds with interesting biological activities. Until recently, only modest attention has been given to metabolites from marine (marine derived) fungi. To date, a number of new compounds from marine fungi have been described and reviewed (Biabani and Laatsch, 1998; Faulkner, 2000, 1992; Jensen and Fenical, 2002; Proksch et al., 2003; Bugni and Ireland, 2004). These documented discoveries clearly showed that marine fungi are of high profile source of structurally unique and biologically active natural compounds as compared to terrestrial fungi. Fungi associated with sponges are the single most potential source of new marine fungal compounds and display diverse biological activities (Jensen and Fenical, 2000; Bugni and Ireland, 2004). These fungi account for the largest number of total described marine fungal compounds and produce highest number of novel metabolites (Bugni and Ireland, 2004).

Encouraged by the idea of "Drugs from the Sea", the chemists have identified many bioactive compounds with novel structures from marine bio-resources in recent years (Faulkner, 2000; Marris, 2006). Among them, marine derived fungi have contributed an important proportion. Many marine fungal strains were isolated, screened and reported to produce novel antimicrobial compounds like alkaloids, macrolides, terpenoids, peptide derivatives and other types (Bugni and Ireland, 2004; Saleem et al., 2007). These marine fungal derived compounds have provided us with new choices to fight infectious diseases. For example, the marine fungus *Pestalotia* spp. isolated from the surface of the brown

alga *Rosenvingea* spp. was able to produce a new chlorinated benzophenone compound pestalone, which showed potent antibiotic activity against multi drug resistance *Staphylococcus aureus* (MRSA), revealing its potential as a new antibiotic. Current studies indicate that complex interactions exist between the host and their epi/endophytic fungi, e.g. the host provides organic nutrition and epi/endophytes act as chemical guards (Tan and Zou, 2001; Bugni and Ireland, 2004). As compared to free-living marine fungi, the epi/endophytic marine fungi have drawn more interest of natural product chemists in search for novel antimicrobial or other active compounds. According to Bugni and Ireland (2004) among the new compounds reported from marine derived fungi till 2002, most of them are produced by epi/endophytes isolated from plants, invertebrates and woody substrates.

CONCLUSION

The world oceans comprise a rich diversity of microbial life with current estimates reaching over a million different species. The vast metabolic diversity of marine microorganisms is underpinned by novel enzymatic functions. While marine microorganisms have already proven to be a rich source of biologically active compounds (bioactives), recent large-scale surveys indicated the existence of thousands of other yet undescribed protein families. Chemically mediated interactions and communications between the microorganisms and their eukaryotic hosts are also likely to have a significant impact on the composition and function of surface consortia. These factors shape the composition and properties of the surface community. The ocean represents a rich resource of even more novel compounds with great potential as pharmaceutical, nutritional supplements, cosmetics, agrichemicals and enzymes, where each of these marine bioproducts have a strong potential market value.

REFERENCES

- Alsumard LC, Golubic T, Hutchings P (1995). Microbial endoliths in skeletons of live and dead corals: *Porites lobata* (Moorea, French Polynesia). *Mar. Ecol.* 117:149-157.
- Anaissie E (1992). Opportunistic mycoses in the immune compromised host: experience at a cancer center and review. *Clin. Infect. Dis. Suppl.* 14:43-53.
- Anke T (1989). Basidiomycetes: a source for new bioactive secondary metabolites. *Prog. Ind. Microbiol.* 27:51-66.
- Anke T (1995). The antifungal strobilurins and their possible ecological role. *Can. J. Bot.* 73:940-945.
- Ashadevi NK, Rajendran R, Karthik Sundaram S (2011). Isolation and characterization of bioactive compounds from marine bacteria. *Indian J. Nat. Resour.* 2:59-64.
- Barrett (2002). From natural products to clinically useful antifungals. *Biochim. Biophys. Acta.* 1587:224-233.
- Berdy J (2005). Bioactive microbial metabolites. *J. Antibiot.* 58:1-26.
- Bertoldo C, Antranikian G (2002). Starch hydrolyzing enzymes from thermophilic archaea and bacteria. *Curr. Opin. Chem. Biol.* 6:151-160.

- Biabani Farooq MA, Laatsch H (1998). Advances in chemical studies on low-molecular weight metabolites of marine fungi. *J. Prakt. Chem.* 340:589-607.
- Brammavidhya S, Usharani G (2013). Bioactive potential of sponge associated *Bacillus cereus* SBS02 isolated from *Hyattella cribriformis*. *Int. J. Res. Environ. Sci. Technol.* 3(2):61-64
- Bringmann G, Lang G, Mühlbacher J, Schaumann K, Steffens S, Rytik P, Hentschel G, Morschhäuser J, Müller WEG (2003). Sorbicillactone A: a structurally unprecedented bioactive novel-type alkaloid from a sponge-derived fungus. *Prog. Mol. Subcell. Biol.* 37:231-253.
- Bugni TS, Ireland CM (2004). Marine-derived fungi: a chemically and biologically diverse group of microorganisms. *Nat. Prod. Rep.* 21:143-163.
- Carte BK (1996). Biomedical potential of marine natural products. *J. Biosci.* 46:271-286
- Coulon D, Sinclair CS, Hill RT, Tambutte S, Puvarel S, Ostrander GK (2004). A basidiomycete isolated from the skeleton of *Pocillopora damicornis* (Scleractinia) selectively stimulates short-term survival of coral skeletogenic cells. *Mar. Biol.* 144:583-592.
- Dasari VR, Muthyala MK, Nikku MY, Donthireddy SR (2012). Novel Pyridinium compound from marine actinomycete, *Amycolatopsis alba* var. nov. DVR D4 showing antimicrobial and cytotoxic activities *in vitro*. *Microbiol. Res.* 167:346-351.
- Davidson SK, Allen SW, Lim GE, Anderson CM, Haygood MG (2001). Evidence for the biosynthesis of bryostatins by the bacterial symbiont *Candidatus Endobugula Sertula* of the Bryozoan *Bugula Neritina*. *Appl. Environ. Microbiol.* 67:4531-4537.
- Donadio S, Carrano L, Brandi L, Serina S, Soffientini A, Raimondi E, Montanini N, Sosio MC, Gualerzi O (2002). Targets and assays for discovering novel antibacterial agents. *J. Biotechnol.* 99:175-185.
- Faulkner DJ (1992). Biomedical uses for natural marine chemicals. *oceanus* 35:29-35.
- Faulkner DJ (2000). Highlights of marine natural products chemistry (1972-1999). *Nat. Prod. Rep.* 17:1-6.
- Fenical W (1993). Chemical studies of marine bacteria: developing a new resource. *Chem. Rev.* 93:1673-1683.
- Fenical W, Jensen PR (1993). Marine microorganisms: A New Biomedical Resource. In: *Marine biotechnology*. (Eds) Attaway D and Zaborsky, Plenum Press: New York. pp. 419-457.
- Fremlin LJ, Piggott AM, Lacey E, Capon RJ (2009). Cottoquinazoline A and Cotteslosins A and B, metabolites from an Australian marine-derived strain of *Aspergillus versicolor*. *J. Nat. Prod.* 72:666-670.
- Gaino E, Sara M (1994). An ultrastructural comparative study of the eggs of 2 species of *Tethya* (Porifera, Demospongiae). *Invertebr. Reprod. Dev.* 26:99-106.
- Golubic S, Radke G, Alsumard TLC (2005). Endolithic fungi in marine ecosystem. *Trends Microbiol.* 13:229-235.
- Graham CR, David RW, Frank TR (1980). Peptone induction and rifampin-insensitive collagenase production by *Vibrio alginolyticus*. *J. Bacteriol.* 142:447-454.
- Gustafson K, Roman M, Fenical W (1989). The microlactins, a novel class of antiviral and cytotoxic macrolides from deep-sea marine bacterium. *J. Am. Chem. Soc.* 111:7519-7524.
- Han Y, Yang B, Zhang F, Miao X, Li Z (2009). Characterization of antifungal chitinase from marine *Streptomyces* sp. DA11 associated with South China sea sponge *Craniella australiensis*. *Mar. Biotechnol.* 11:132-140.
- Hentschel U, Bringmann G, Lang G, Mühlbacher J, Schaumann K, Steffens S, Rytik PG, Morschhäuser J, Müller WE (2003). Sorbicillactone A: a structurally unprecedented bioactive novel-type alkaloid from a sponge-derived fungus. *Prog. Mol. Subcell. Biol.* 37:231-253.
- Holst PB, Anthoni U, Christophersen C, Neilsen PH (1994). Marine alkaloids, 15. Two alkaloids, flustramine E and debromoflustramine B, from the marine bryozoan *Flustra foliacea*. *J. Nat. Prod.* 57:997-1000.
- Hugenholtz P, Pace NR (1996). Identifying microbial diversity in natural environment: a molecular phylogenetic approach. *Trends. Biotechnol.* 14:190-197.
- Ireland CM, Copp BR, Foster MD, McDonald LA, Radisky DC, Swersey JC (1993). Biomedical potential of marine natural products. In: *Marine Biotechnology Vol. I: Pharmaceutical and bioactive natural products*. (Eds) D.H. Attaway and O.R. Zaborsky, Plenum Press, New York. pp. 1-43.
- Jensen PR, Fenical W (2000). Marine organisms and drug discovery: Current status and future potential. In: *Drugs from sea*. (Eds) Fuestini N.S., Kruger publisher, Basel. pp. 6-29.
- Jensen PR, Fenical W (2002). Secondary metabolites from marine fungi. In: *Fungi in marine environments* (Eds) Kevin DH, Hong Kong. pp. 293-315.
- Jeong S, Ishida K, Ito Y, Okada S, Murakami M (2003). Bacillamide, a novel algicide from the marine bacterium, *Bacillus* sp. SY-1, against the harmful dinoflagellate, *Cochlodinium polykrikoides*. *Tetrahedron Lett.* 44:8005-8007.
- Kobayashi J, Ishibashi M (1993). Bioactive metabolites of symbiotic marine microorganisms. *Chem. Rev.* 93:1753-1769.
- Lamela M, Anca J, Villar R, Otcro J, Calleja JM (1989). Hypoglycemic activity of several seaweed extracts. *J. Ethno. Pharmacol.* 27:35-43.
- Li HY, Matsunaga S, Fusetani N (1998). Antifungal metabolites from marine sponges. *Curr. Org. Chem.* 2:649-682.
- Li Z, Hu Y, Liu Y, Huang Y, He L, Miao X (2007). 16S rDNA clone library-based bacterial phylogenetic diversity associated with three South China Sea sponges. *World J. Microbiol. Biotechnol.* 23:1265-1272.
- Liu R, Cui C, Duan L, Gu Q, Zhu W (2005). Potent *in vitro* anticancer activity of metacycloprodigiosin and undecylprodigiosin from a sponge-derived actinomycetes *Saccharopolyspora* sp. nov. *Arch. Pharm. Res.* 28:1341-1344.
- Marris E (2006). Marine natural products: Drugs from the deep. *Nature* 443:904-905.
- Misra JK, Lichtwardt RW (2000). Illustrated genera of trichomycetes: Fungal symbionts of insects and other arthropods. (Eds) Science Publishers Inc, Enfield, New Hampshire, 155
- Mithun VSL, Rama Chandra Rao CSV (2012). In vitro studies on anti-cancer activity of anti-cancerous compound producing marine bacteria against on cancer cells by MTT assay. *J. Chem. Biol. Phys. Sci. Sec. B.* 3:276-282.
- Osama R, Koga T (1995). An investigation of aquatic bacteria capable of utilizing chitin as the sole source of nutrients. *Lett. Appl. Microbiol.* 21:288-291.
- Sivasankar P, Sugesh S, Vijayanand P, Sivakumar K, Vijayalakshmi S, Balasubramanian T, Mayavu P (2013). Efficient production of L-asparaginase by marine *Streptomyces* sp. isolated from Bay of Bengal, India Vol. 7(31):4015-4021.
- Pandey S, Ayinampudi S, Soumya SD, Dipti PS, Lipsa C (2013). Diversity of marine bacteria producing beta-glucosidase inhibitors. *Microb. Cell Fact.* 12:35 2-5.
- Parkes RJ, Cragg BA, Bale SJ, Getliff JM, Goodman K, Rochelle PA, Fry JC, Weightman AJ, Harvey SM (1994). A deep bacterial biosphere in Pacific Ocean sediments. *Nature* 371:410-413
- Pivkin MV (2000). Filamentous fungi associated with holothurians from the sea of Japan, off the Primorye coast of Russia. *Biol. Bull.* 198:101-109.
- Proksch P, Ebel R, Edrada RA, Wray V, Steube K (2003). Bioactive natural products from marine invertebrates and associated fungi. In: *Sponges (Porifera)*, (Eds) Müller WEG, Springer, Berlin Heidelberg New York. pp. 117-138.
- Raghukumar C, Raghukumar S (1991). Fungal invasion of massive corals. *Mar. Ecol.* 12:251-260.
- Raghukumar C, Lande V (1988). Shell disease of rock oyster *Crassostrea cucullata*. *Dis. Aquat. Org.* 4:77-81.
- Ravikumar S, Fredimoses M, Dnanadesigan M (2012). Anticancer property of sediment actinomycetes against MCF-7 and MDA-MB-231 cell lines. *Asian Pac. J. Trop. Biomed.* 2:92-96.
- Revathy T, Jayasri MA, Suthindhiran K (2013). Anti-oxidant and enzyme-inhibitory potential of marine *Streptomyces*. *Am. J. Biochem. Biotechnol.* 9(3):282-290.
- Ravindran J, Raghukumar C, Raghukumar S (2001). Fungi in *Porites lutea*: association with healthy and diseased corals. *Dis. Aqua. Org.* 47:219-228.
- Saleem M, Ali MS, Hussain S, Jabbar A, Ashraf M, Lee YS (2007). Marine natural products of fungal origin. *Nat. Prod. Rep.* 24:1142-1152.

- Saurav K, Kannabiran K (2012). Cytotoxicity and antioxidant activity of 5-(2,4-dimethylbenzyl) pyrrolidin-2-one extracted from marine *Streptomyces* VITSVK5 spp. Saudi J. Biol. Sci. 19:81-86.
- Shearer CA, Maivan ZH (1988). *In vitro* hyphal interactions among wood and leaf inhabiting ascomycetes and fungi imperfecti from fresh water habitats. Mycologia 80:31-37.
- Sousa TS, Jimenez PC, Ferreira EG, Silveira ER, Braz-Filho R, Pessoa ODL, Costa-Lotufo LV (2012). Anthracyclinones from *Micromonospora* sp. J. Nat. Prod. 75:489-493.
- Sugano M, Sato A, Iijima Y, Oshima T, Furuya K, Kuwano H, Hata T, Hanzawa H (1991). Phomactin A: A novel PAF antagonist from a marine fungus *Phoma* sp. J. Am. Chem. Soc. 113:5463-5464.
- Tan RX, Zou WX (2001). Endophytes a rich source of functional metabolites. Nat. Prod. Rep. 18:448-459.
- Wang F, Fang Y, Zhu T, Zhang M, Lin A, Gu Q, Zhu W (2008). Seven new prenylated indole diketopiperazine alkaloids from holothurians derived fungus *Aspergillus umigatus*. Tetrahedron 64:7986-7991.
- Xie H, Pasternak JJ, Glick BR (1996). Isolation and characterization of mutants of the plant growth-promoting rhizobacterium *Pseudomonas putida* GR12-2 that over produce indole acetic acid. Curr. Microbiol. 32:67-71.
- Xin Z, Fang Y, Du L, Zhu T, Duan L, Chen J, Gu Q, Zhu W (2007). Aurantiomides A-C, quinazoline alkaloids from the sponge-derived fungus *Penicillium aurantiogriseum* SP0-19. J. Nat. Prod. 70:853-855.
- Xin ZH, Zhu WM, Gu QQ, Fang YC, Duan L, Cui CB (2005). A new cytotoxic compound from *Penicillium auratiogriseum*, symbiotic or epiphytic fungus of sponge *Mycale plumose*. Chin. Chem. Lett. 16:1227-1229.
- Yang LH, Miao L, Lee OO, Li X, Xiong H, Pang K, Virjmoed L, Qian P (2007). Effect of culture conditions on antifouling compound production of a sponge-associated fungus. Appl. Microbiol. Biotechnol. 74:1221-1231.
- Yang R, Li C, Lin Y, Peng G, She Z, Zhou S (2006). Lactones from a brown alga endophytic fungus (No. ZZ36) from the South China sea and their antimicrobial activities. Bioorg. Med. Chem. Lett. 16:4205-4208.
- Yu L, Li Z, Peng C, Li Z, Guo Y (2009). Neobacillamide A, a novel thiazole-containing alkaloid from the marine bacterium, *Bacillus vaiismortis* C89, that was associated with South China Sea sponge *Dysidea avara*. Helvetica Chimica Acta 92:607-612.
- Zheng L, Chen H, Han X, Lin W, Yan X (2005). Antimicrobial screening and active compound isolation from marine bacterium NJ6-3-1 associated with the sponge *Hymeniacidon perleve*. World J. Microbiol. Biotechnol. 21:201-206.

Full Length Research Paper

Siderophore mediated antagonism of fluorescent Pseudomonads against soil borne plant pathogenic fungi in West Bengal, India

A. Adhikari^{1,2#}, K. Sarker^{2,3*#}, M. De Roy⁴, I. Bhattacharya², T. Mandal¹, P. K. Dasmohapatra³ and S. Dutta^{2*}

¹Department of Biotechnology, O. I. S. T., Vidyasagar University, Midnapore, India.

²Department of Plant Pathology, BCKV, Mohanpur, Nadia, India.

³Department of Microbiology, Vidyasagar University, Midnapore, India.

⁴ADA (Polba block), Hoogly, W. B.; Present add: STL, PORS, Berhampore, West Bengal, India.

Accepted 8 September, 2013

The rhizobacterial isolates from different agro ecological region of West Bengal, used in this study, were identified as fluorescent pseudomonads based on biochemical and molecular characterization. Six effective antagonist isolates were selected for further studies of secondary metabolite (siderophore) production with respect to bacterial growth at different time intervals (6 to 96 h), the nature of siderophore production (hydroxamate and/or catecholate type), effect of iron supplementation at its different concentration on siderophore production and antagonistic potentiality. Observations of higher level of growth and siderophore production, for most of the isolates, were recorded after 72 h of incubation. The maximum zone of inhibition and siderophore production was recorded at 0.5 µM and the least at 10 µM iron concentration. With the gradual increase in iron supplementation in king's B medium, the siderophore production as well as *in vitro* antagonistic potentiality decreased in terms of enhancement of diameter of fungal mycelium and reduction of percent inhibition of pathogen by fluorescent pseudomonads in dual culture assay method.

Key words: Fluorescent pseudomonads, siderophore, ferric ion, *Sclerotium rolfsii*, *Rhizoctonia solani*.

INTRODUCTION

Many genera of plant root-inhabiting bacteria have potential as biological control agents (Weller, 1988). The genus *Pseudomonas* belongs to the γ-subclass of the Proteobacteria and includes mostly fluorescent Pseudomonads as well as a few non-fluorescent species. Particularly fluorescent *Pseudomonas* spp. has received much attention because of their suggested involvement in natural disease suppressiveness of certain soils (Weller et al., 2002). Iron-regulated metabolites are mainly referred to as siderophores. The siderophores are high-affinity iron chelators in bacteria, enabling uptake of Fe

(Braun, 2001). Iron is essential for the growth of almost all organisms but, in many environments, the amount of free iron is below 10^{-7} M, which is the required concentration by most bacteria for growth (Ratledge and Dover, 2000). Bacteria have developed several strategies for the acquisition, solubilization and transportation of iron (Guérinot, 1994). The most efficient mechanism of iron acquisition is the secretion of high affinity iron-chelating compounds, so-called siderophores. The siderophores chelate iron in the extracellular environment and the resulting ferric siderophore complex is recognized by

*Corresponding authors. E-mail: subrata_mithu@yahoo.co.in, kanish.sarker@gmail.com.

#Authors contributed equally to this work.

Table 1. Origins and different crop rhizospheres of the isolates.

Fluorescent pseudomonad isolates	Location	Region	Crop rhizosphere
MFP9	Coochbehar	Terai	Maize
CHFP23	Nadia	Gangetic alluvial	Chilli
TFP16	West Midnapore	Lateritic	Tomato
FPK7	Bagdaha, North 24Pgs	Gangetic alluvial	Pointed gourd
FPK8	Bagdaha, North 24Pgs	Gangetic alluvial	Pointed gourd
FPK10	Habra, North 24 Pgs	Gangetic alluvial	Cucumber

siderophore-specific membrane receptors, enabling uptake of Fe by the bacterial cells (Braun, 2001). Many bacteria produce more than one type of siderophore, as shown for *Enterobacter cloacae*, *Mycobacterium smegmatis* and *Pseudomonas* spp. (Adilakshmi et al., 2000; Mercado-Blanco et al., 2001). There are several reports where the effective role of siderophore on antagonistic potentiality under *in vitro* and *in vivo* conditions, has clearly been discussed (Pal and Gokarn, 2010; Adhikari et al., 2013; Subashri et al., 2013). Although considerable structural variation exists among the several dozen siderophores chemically characterized at the present time, most can be classified as hydroxamates or catechols (Neiland, 1981). The present study on siderophore producing fluorescent pseudomonads was concentrated on the screening for their siderophoregenesis, siderotyping and siderophore mediated antibiosis under *in-vitro* conditions against pathogenic fungi with different concentration of iron supplement.

MATERIALS AND METHODS

Isolation of bacteria

Soil samples were collected from different crop rhizospheres from different agro-ecological region of West Bengal (Table 1). Roots were shaken vigorously to remove loosely adhering rhizospheric soil. To isolate rhizospheric bacteria, 10 g of rhizospheric soil were shaken in 90 ml sterile phosphate buffer saline (PBS) and were four fold diluted and plated on King's B (KB) medium. A total of 100 bacterial isolates were isolated and fluorescent colonies, screened through UV light ($k = 365$ nm), with different morphological characteristics were subcultured by repeated streaking on KB medium. On the basis of siderophore production property five highly and one moderately siderophore producing isolates were selected for further studies.

Characterization of siderophore producing bacteria

Bacterial identification was done by biochemical analysis on the basis of the tests of oxidase, arginine dihydrolase, denitrification, gelatine hydrolysis, levan production, and acid production from trehalose, utilization of tryptophane and L-tartrate coupled with amplification of the *Pseudomonas* genus specific 618 bp fragment of 16S rDNA genes by PCR using the primers PA-GS-F and PA-GS-R corresponding to the 5' (5'-GACGGGTGAGTAATGCCTA-3')

end and 3' end (5'-CACTGGTGTTCCTTCTATA-3') of the 16S rDNA gene (Spilker et al., 2004). Amplification was confirmed by analyzing 5 μ L of each PCR reaction mixture on a 1% agarose gel (Sambrook et al., 1989).

Screening of Siderophore producing bacteria

Qualitative assay

Siderophore production by all the isolates was tested qualitatively by Chrome Azural S (CAS) plate assay (Schwyn and Neilands, 1987). The isolates were spotted over cetrimide agar and incubated for 48 h at 28°C. After incubation, a thin layer of CAS reagent in 0.7% agar was spread on the bacterial growth and plates were again incubated for 24 h at 28°C, formation of yellow orange color zone around the colonies in plate assay indicated the siderophore production.

Quantitative assay

All the isolates were grown at 28 \pm 2°C on a rotary shaker in King's B broth for three days and centrifuged at 10,000 rpm for 10 min and the supernatant was collected. The pH of the supernatant was adjusted to 2.0 with 1 N HCl and equal quantity of ethyl acetate was added in a separating funnel, mixed well and ethyl acetate fraction was collected. 5 mL of ethyl acetate fraction was mixed with 5 mL of Hathway's reagent (The reagent was prepared by adding 1 mL of 0.1 M ferric chloride in 0.1 N HCl to 100 mL of distilled water, and to this 1 mL of 0.1 M potassium ferricyanide was added). The absorbance for dihydroxy phenols was read at 700 nm in a Spectrophotometer (Reeves et al., 1983). A standard curve was prepared using different concentration of dihydroxy benzoic acid at 700 nm. The quantity of siderophore synthesized was expressed as μ mol benzoic acid mL⁻¹ of culture filtrate.

Siderophore typing

Hydroxamate type of siderophore

Csaky's test: Hydroxamate type of siderophore was determined by hydrolyzing 1 ml supernatant of overnight grown culture with 1 ml of 6 N H₂SO₄ in boiling water bath for 6 h or 130°C for 30 min. Further, this hydrolyzed sample was buffered by adding 3 ml of sodium acetate solution. To this, 0.5 ml iodine was added and allowed to react for 3-5 min. After completion of reaction, the excess iodine was destroyed with 1 ml of sodium arsenate solution. Finally 1 ml of α - Naphthylamine solution was added and allowed to develop the colour. Wine red colour formation indicates production of hydroxamate type of siderophore (Gillan et al., 1981).

Neilands spectrophotometric assay

The hydroxamate nature of siderophore was detected by Neilands spectrophotometric assay (Jalal and Vander Helm, 1990; (Neilands, 1981) where a peak between 420-450 nm on addition of 2% aqueous solution of FeCl₃ to 1 mL of supernatant indicated presence of Ferrate hydroxamate.

Catecholate type of siderophore

Arnow's Test

To 1 ml of cell-free supernatant, 1 ml of nitrite-molybdate reagent with 1 ml NaoH solution was added. Finally, 1 ml of 0.5 N HCl was added and allowed to develop colour. Yellow colour formation indicates production of catecholate type siderophore (Arnow, 1937).

Spectrophotometric assay

Catecholate nature of siderophore was detected by the method of Jalal and Vander Helm (1990) using spectrophotometric assay where a peak at 495 nm on addition of 2% aqueous solution of FeCl₃ to 1 mL of supernatant indicated the presence of siderophores of catecholate nature.

Temporal variation of bacterial growth and siderophore production

All the six fluorescent pseudomonads were grown in king's B medium by submerged culture method in 50 mL conical flasks and incubated at 28±2°C on a rotary shaker at 150 rpm. Samples were withdrawn in duplicate at definite time intervals from 6 h up to 96 h. One set was used for growth measurement at 600 nm and the other for estimation of siderophore at 700 nm (Reeves et al., 1983).

Influence of iron on production of siderophore

In order to determine the concentration of iron at which siderophore biosynthesis is minimized in fluorescent pseudomonads under study, all the cultures were grown in king's B media, externally supplemented with different concentrations of iron (FeCl₃. 6H₂O) viz., 0.5, 1, 1.5, 2.5, 5 and 10 µM. Following the incubation at 28±2°C for 72 h at 120 rpm, siderophore contents were estimated at 700 nm. To remove traces of iron, glasswares were cleaned with 6 M HCl and with double distilled water.

In vitro antagonistic effect on mycelial growth of phytopathogen under different iron limiting conditions

An agar plug (7 mm diameter) taken from actively growing fungal culture such as *Rhizoctonia solani* and *Sclerotium rolfsii* were placed on the surface of the plate-enhancing medium supplemented with FeCl₃ at different concentration that is 0.5, 1, 1.5, 2.5, 5, 10 µM. Simultaneously, fluorescent pseudomonas isolates were streaked 3 cm away from the agar plug at both sides towards the edge of Petri plates (Kumar et al., 2000). Plates inoculated with fungal agar plugs alone were used as control. The plates were incubated at 28±2°C until fungal mycelia completely covered the agar surface in control plate. Observations on mycelial growth of test pathogens were recorded and percent inhibition of pathogen growth were calculated:

$$\%I = 100(C-T)/C$$

Where, I = inhibition of mycelial growth, C = growth of pathogen in the control plate (cm) and T = growth of pathogen in dual cultures (cm).

Statistical analysis

All the isolates were clustered with respect to their siderophoregenesis at different ferric ion concentration, zone of inhibition, inhibition percentage, temporal growth measurement and siderophore production at different time intervals by critical difference at 5% probability level using DMRT by SPSS v.10.

RESULTS AND DISCUSSION

Characterization of siderophore producing bacteria

Biochemical characterization

Among the bacterial isolates, five highly and one moderately siderophore producing isolates were confirmed as fluorescent pseudomonads based on biochemical tests such as gram reaction, oxidase, arginine dihydrolase, denitrification, gelatine hydrolysis, levan production, and acid production from trehalose, utilization of tryptophane and L-tartrate. Microscopic observation showed that all the six isolates were rod shaped, motile, gram negative, and fluorescent in the presence of UV light (Rao et al., 1999; Srivastava et al., 1999). All the isolates exhibited positive reaction in catalase, oxidase, arginine di hydrolase and negative in TABAC hypersensitivity test (Bossis et al., 2000). All the five isolates except FPK 10 were positive in gelatin liquefaction test; isolates MFP9 and FPK7 showed positive in trehalose test; MFP9 was positive in levan production and FPK7, FPK8 showed denitrification activity; whereas, the isolate TFP16 exhibited positive reaction in all the cases except TABAC hypersensitivity (Table 2).

Molecular characterization

PCR assays employing genus specific primer pair PA-GS-F/PA-GS-R produced 618 bp DNA fragments of the predicted size for all the six isolates (Figure 1). With the presence of the 618 bp amplicon the six putative siderophore producing isolates were confirmed as *Pseudomonas sp.* (Spilker et al., 2004). All the isolates produced diffusible fluorescent pigment which was confirmed in the presence of UV light, which indicated that all the isolates belonged to fluorescent pseudomonads.

Qualitative assay for siderophore production

All the six isolates were subjected to screening for side-

Table 2. Different biochemical tests performed to identify the isolates.

Fluorescent pseudomonad isolates	Fluorescence	Arginine	Oxidase	catalase	Tabac	Gelatine	Trehalose	Levan	Denitrification	L-ara	L(+)tart
MFP9	+	+	+	+	-	+	+	+	-	NA	NA
CHFP23	+	+	+	+	-	+	-	-	-	NA	NA
TFP16	+	+	+	+	-	+	+	+	+	+	+
FPK7	+	+	+	+	-	+	+	-	+	NA	NA
FPK8	+	+	+	+	-	+	-	-	+	NA	NA
FPK10	+	+	+	+	-	-	-	NA	NA	NA	NA

'+' indicates positive; '-' indicates negative; 'NA' indicates not applicable; 'L-ara' indicates L- arabinose; 'L (+) tart' indicates L-tartarate.

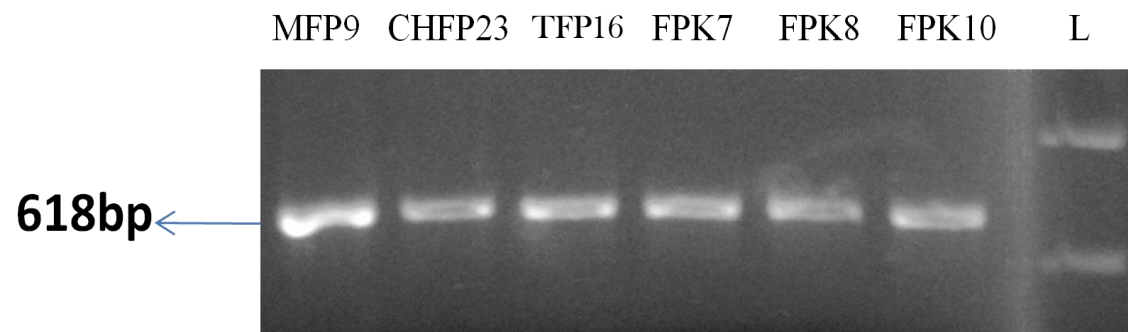


Figure 1. Agarose gel electrophoresis of PCR amplified genus specific 618 bp 16S rDNA amplicons; Lanes are named with respective isolates; L: 100 bp molecular marker.

rophore production. Isolates CHFP23, FPK8, FPK10 produced a strong reaction with the CAS reagent (Table 3). After incubating for a period of 24 h in the dark at 28°C, a change in color from blue to orange was observed in the agar plate (Figure 2). The color change from blue to orange resulted by siderophoretic removal of Fe from the dye.

Quantitative assay for siderophore production

All the six isolates were subjected to screening for

siderophore production. Three isolates, viz., FPK8, FPK10, CHFP23, were found to be highly siderophore producer such as 15.20 (\pm 0.33), 13.9 (\pm 0.30) and 11.55 (\pm 0.21), μ M/ml respectively, whereas, moderately siderophore production was obtained from the isolates TFP16 and FPK7 and the least siderophore production was observed in MFP9 (Table 4).

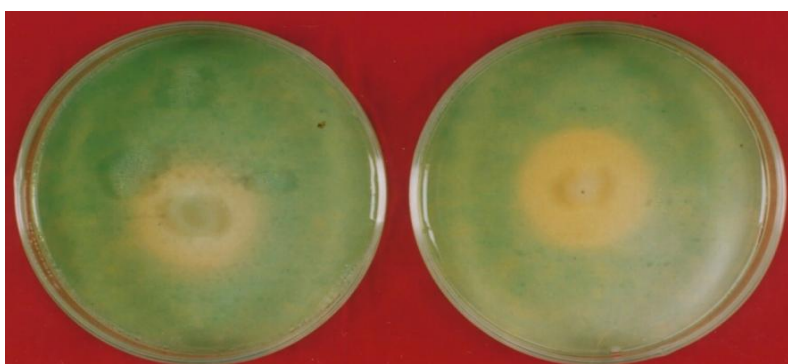
The siderophores have very high affinity for ferric iron and form ferric-siderophores complex making it unavailable to other organisms but the producer organism can utilize these complexes

via a specific receptor in their outer cell membrane (Buyer and Leong, 1986). Fluorescent pseudomonads produce several siderophores such as pyoverdine, pyochelin (Dave and Dube, 2000) and are important to agriculture because they can reduce crop yield losses caused by bacteria and fungi in the root environment. The protective activity is related to the production and excretion of siderophores which efficiently chelate the iron in the root environment. This iron deficiency leads to an impaired growth of the deleterious micro-organisms.

Table 3. Siderophore production under *in vitro* condition by six different fluorescent pseudomonads.

Fluorescent pseudomonad isolates	Siderophore production in CAS agar plate assay
MFP9	+
CHFP23	+++
TFP16	++
FPK7	++
FPK8	+++
FPK10	+++

'+' indicates weak producer; '++' indicates medium producer and '+++ indicates good producer.

**Figure 2.** Siderophore production by bacterial isolates in CAS plate assay.**Table 4.** Siderophore production under *in vitro* condition by six different fluorescent pseudomonads.

Fluorescent pseudomonad isolates	Siderophore production (μM benzoic acid/mL)
	Mean \pm SD
MFP9	2.04 \pm 0.13
CHFP23	11.55 \pm 0.21
TFP16	9.87 \pm 0.25
FPK7	6.98 \pm 0.23
FPK8	15.20 \pm 0.33
FPK10	13.9 \pm 0.30

Average mean \pm SD of three replicates, $P < 0.05$.

Assay for siderophore typing

The type of siderophore was determined by Csaky and Arnow assay, where isolates TFP16 and FPK8 has shown hydroxamate type of siderophore production that is formation of wine red colour in supernatant. Further spectrophotometric analysis of the culture, TFP16 and FPK8 in King's B medium showed a sharp peak between 420 to 450 nm (Figure 3a) and clearly indicated the presence of siderophores of ferrate hydroxamate nature.

Catechol type of siderophore was determined by Arnow's test where the formation of yellow colour was

observed in the supernatant of the isolates MFP9, CHFP23, FPK7, FPK10. A peak seen between 490 to 505 nm confirmed the presence of siderophores of catechol nature (Figure 3b and 3c) (Ali and Vidhale, 2011). Evidence for the production of catechol or hydroxamate siderophores by different bacterial species has been shown previously (Hohlneicher et al., 1995; Howard et al., 2000; Gull and Hafeez, 2012).

Siderophores usually form a stable, hexadentate, octahedral complex with Fe^{3+} preferentially compared to other naturally occurring abundant metal ions. Siderophores are usually classified by the ligands used to

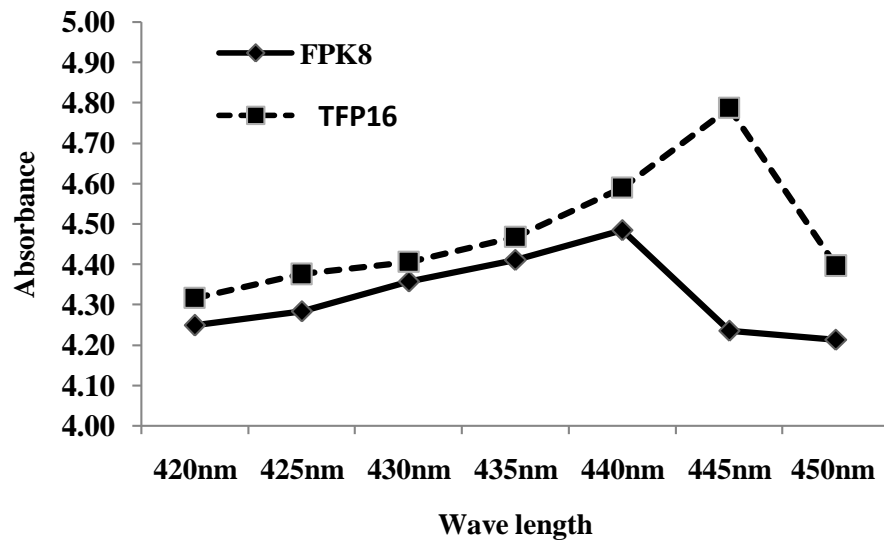


Figure 3a. Absorption characteristics FPK8 and TFP16 extract confirming hydroxamate nature of siderophores. Scanning of absorption characteristics were between 400-550 nm; a sharp peak between 420 and 450 nm confirmed hydroxamate nature of siderophore of FPK8 and TFP16.

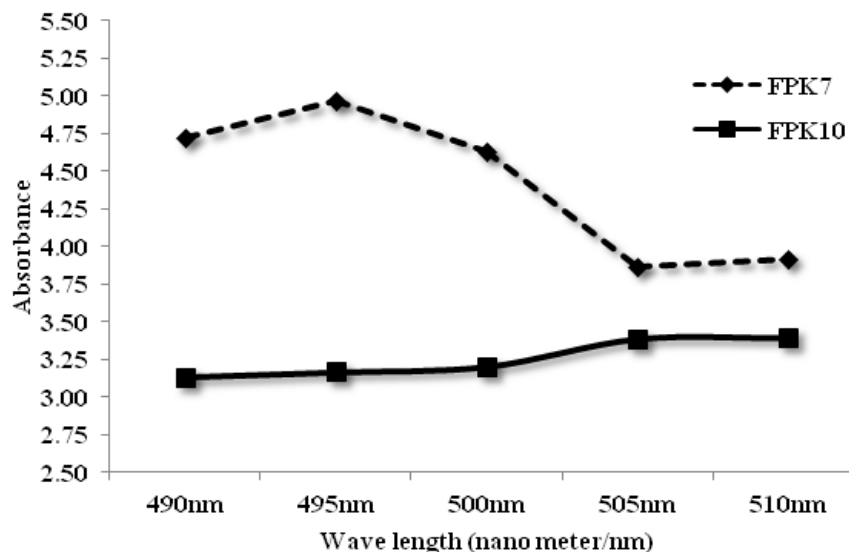


Figure 3b. Absorption characteristics FPK7 and FPK10 extract confirming catecholate nature of siderophores. Scanning of absorption characteristics were between 400-550 nm; a sharp peak between 490 and 510 nm confirmed catecholate nature of siderophore of FPK7 and FPK10.

chelate the ferric iron. The major groups of siderophores include the catecholates (phenolates), hydroxamates and carboxylates (derivatives of citric acid) (Hofte, 1993).

Temporal variation of bacterial growth and siderophoregenesis

In the growth and siderophore production, as depicted in

Figure 4a to 4f, an exponential phase from 6 to 48 h was observed regarding the isolates MFP9, CHFP23, TFP16 and FPK8; stationary phase was initiated after 48 h. In the case of the isolates FPK7 and FPK10, growth was ceased after 72 h. Threshold level of siderophore production was observed after 12 h of incubation. Syeed et al. (2005) also found similar result that siderophore production started after 12 h of incubation, while working

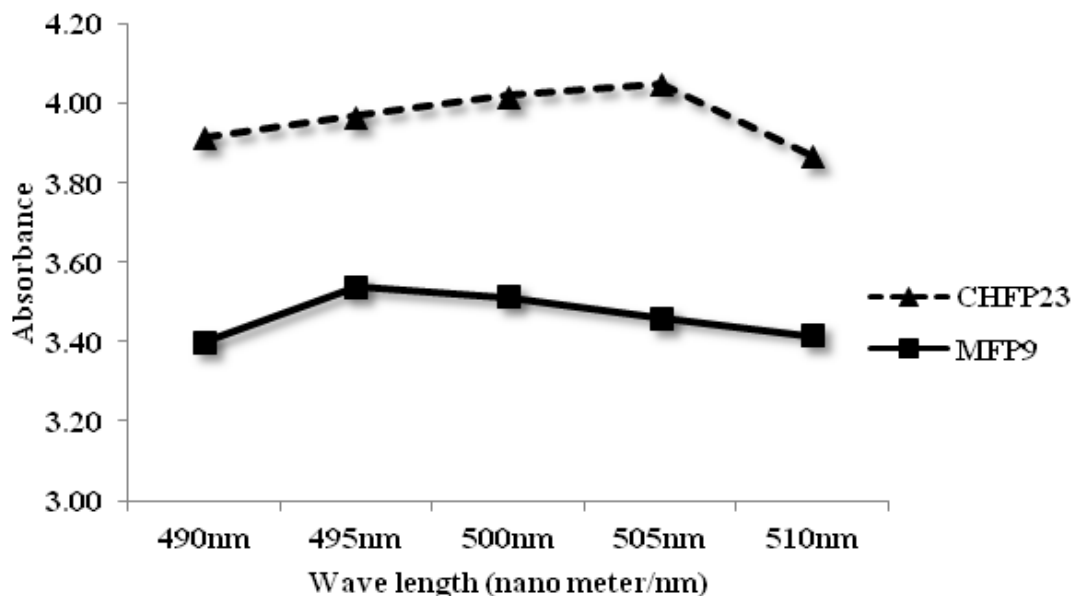


Figure 3c. Absorption characteristics MFP9 and CHFP23 extract confirming catechol nature of siderophores. Scanning of absorption characteristics were between 400-550 nm; a sharp peak between 490 and 510 nm confirmed catechol nature of siderophore of MFP9 and CHFP23.

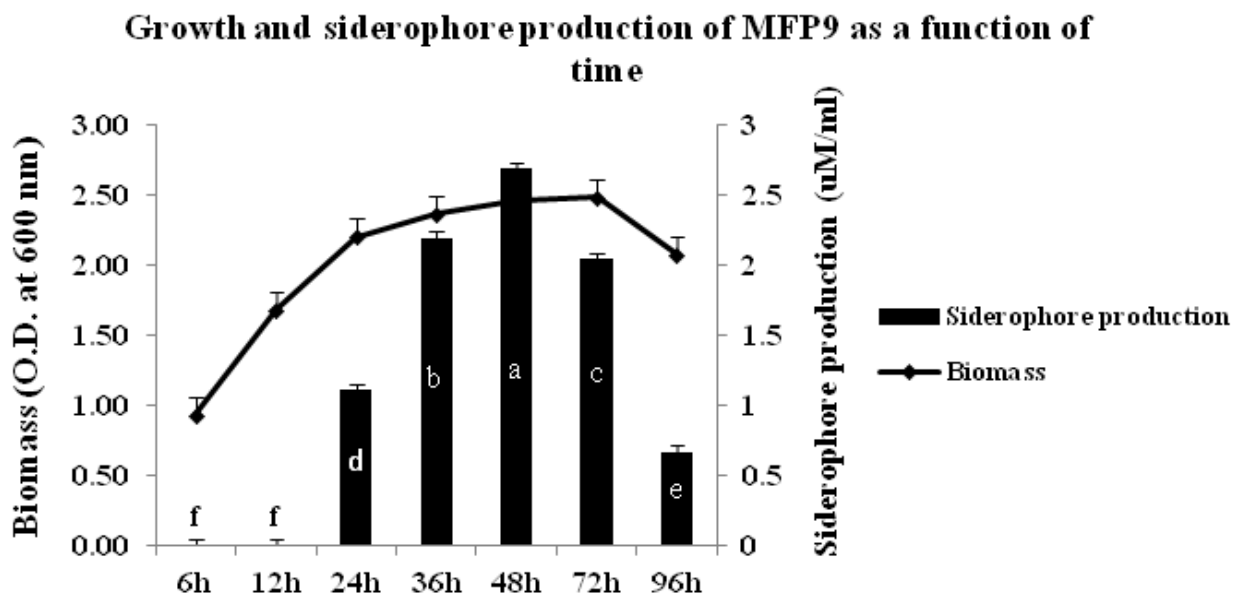


Figure 4a. Temporal growth measurement and siderophore production of MFP9 as a function of time. Vertical bar shows critical difference values. Values are mean of three replications. A common letter (letters on the bars) means they are not significantly different ($p=0.05$) by DMRT.

with two fluorescent pseudomonads. For all the five isolates except MFP9, production of siderophore was increased up to 72 h and declined thereafter, for MFP9 it was after 48 h. Thus, maximum siderophore yield was observed at the end of log phase growth (Sharma and Johri, 2003).

Influence of iron on siderophoregenesis

In order to assess the influence of iron concentration on siderophore release of six different isolates, King's B media supplemented with different concentration of ferric ion was used. After 72 h of incubation, maximum side-

Growth and siderophore production of CHFP23 as a function of time

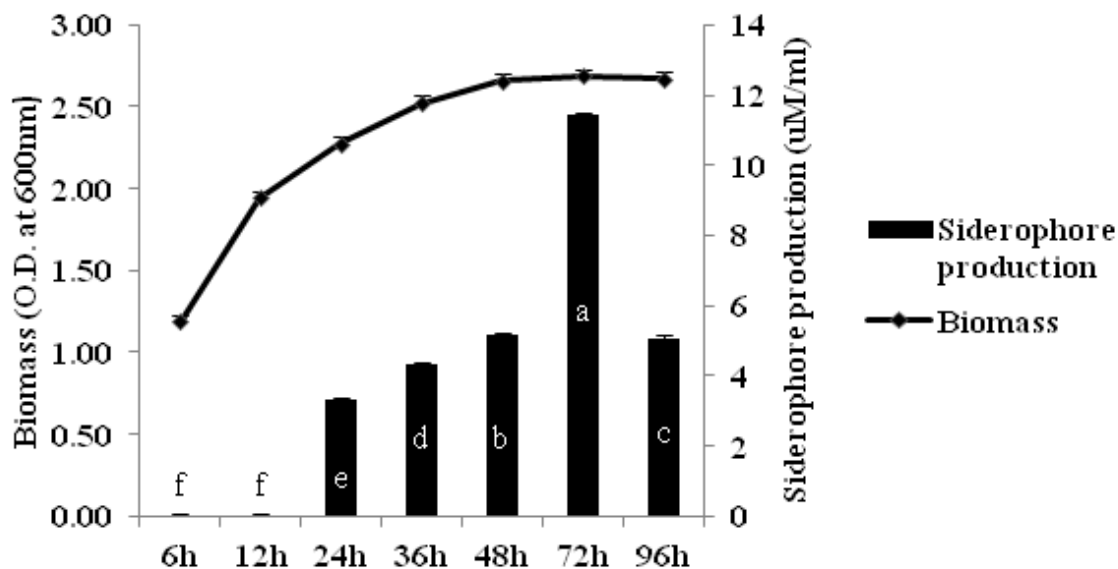


Figure 4b. Temporal growth measurement and siderophore production of CHFP23 as a function of time. Vertical bar shows critical difference values. Values are mean of three replications. A common letter (letters on the bars) means they are not significantly different ($p=0.05$) by DMRT.

Growth and siderophore production of TFP16 as a function of time

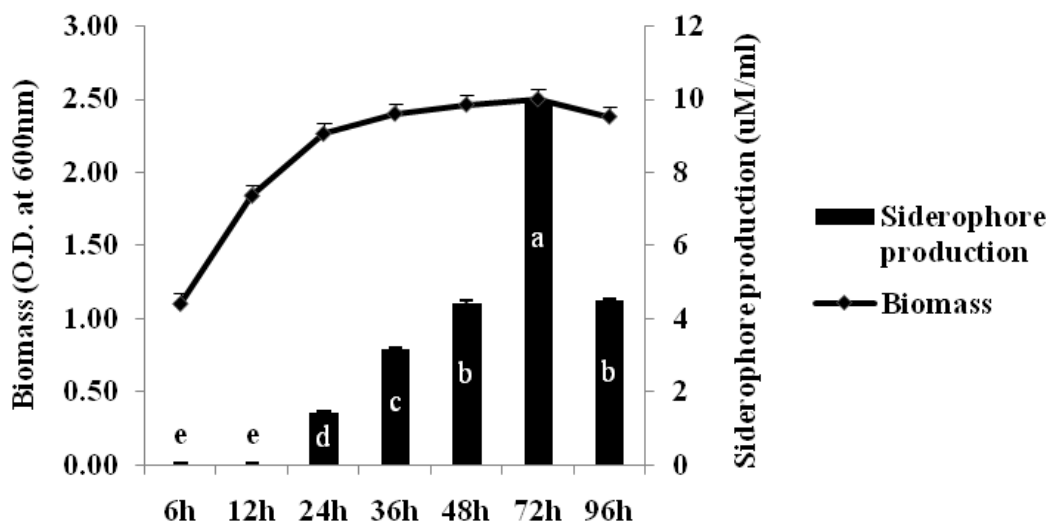


Figure 4c. Temporal growth measurement and siderophore production of TFP16 as a function of time. Vertical bar shows critical difference values. Values are mean of three replications. A common letter (letters on the bars) means they are not significantly different ($p=0.05$) by DMRT.

rophore productions for all the isolates under study were obtainable at $0.5 \mu\text{M Fe}^{3+}$ concentration and the lowest siderophore level was recorded at $10.0 \mu\text{M Fe}^{3+}$ (Table

5). Siderophores are iron-specific compounds which are synthesized and secreted under iron stress condition and for each isolate under study siderophore production was

Growth and siderophore production of FPK7 as a function of time

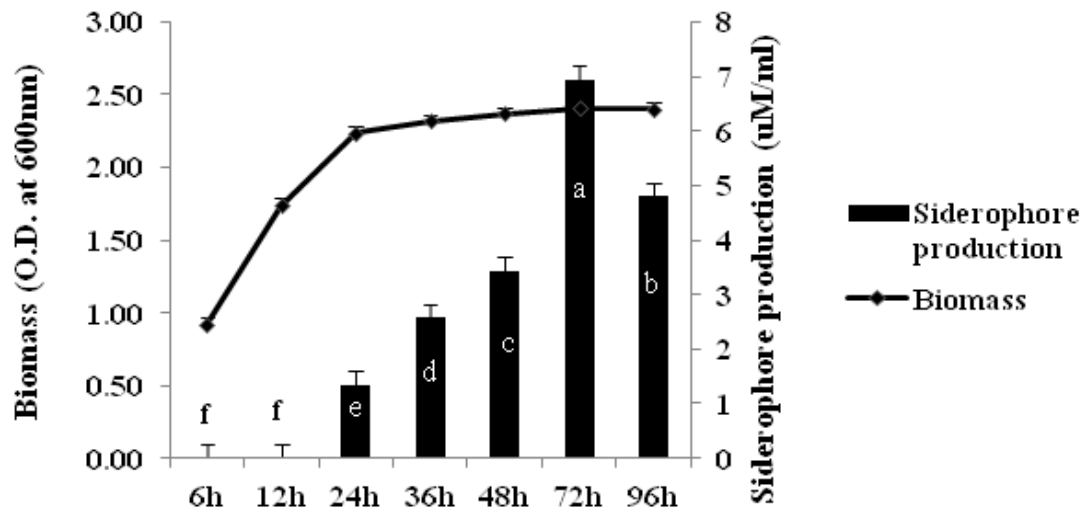


Figure 4d. Temporal growth measurement and siderophore production of FPK7 as a function of time. Vertical bar shows critical difference values. Values are mean of three replications. A common letter (letters on the bars) means they are not significantly different ($p = 0.05$) by DMRT.

Growth and siderophore production of FPK8 as a function of time

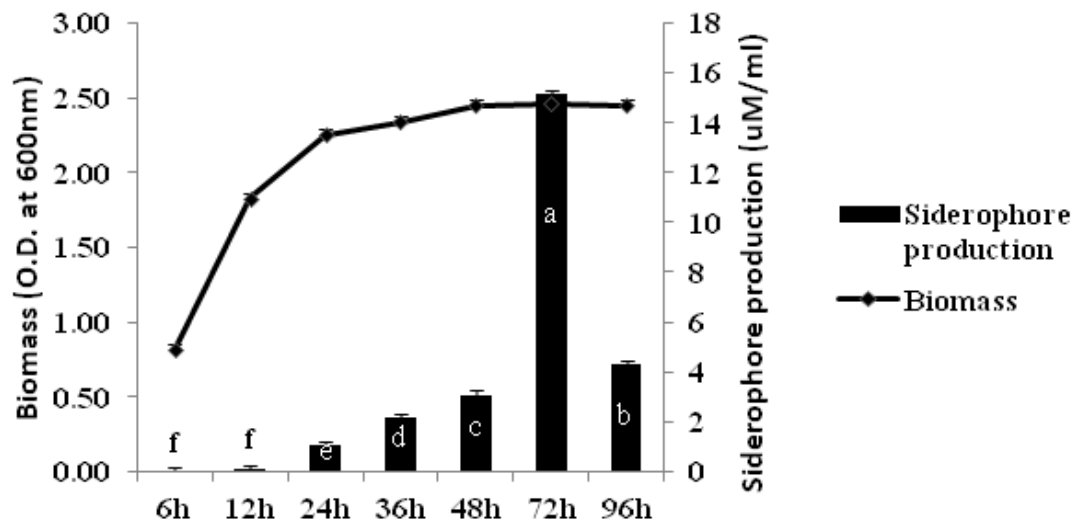


Figure 4e. Temporal growth measurement and siderophore production of FPK8 as a function of time. Vertical bar shows critical difference values. Values are mean of three replications. A common letter (letters on the bars) means they are not significantly different ($p = 0.05$) by DMRT.

gradually repressed with the increasing concentration of iron (0.5, 1, 1.5, 2.5, 5, 10 μM FeCl_3) (Budzikiewicz, 1993; Rachid and Ahmed, 2005; Sayyed et al., 2005).

The universal repressor 'Fur' acts together with iron

under iron rich conditions. When the intra-cellular environment is iron rich, the Fe^{2+} -Fur complex bind to promoter containing a 'Fur box' and represses transcription of the genes involving in iron uptake. Moreover,

Growth and siderophore production of FPK10 as a function of time

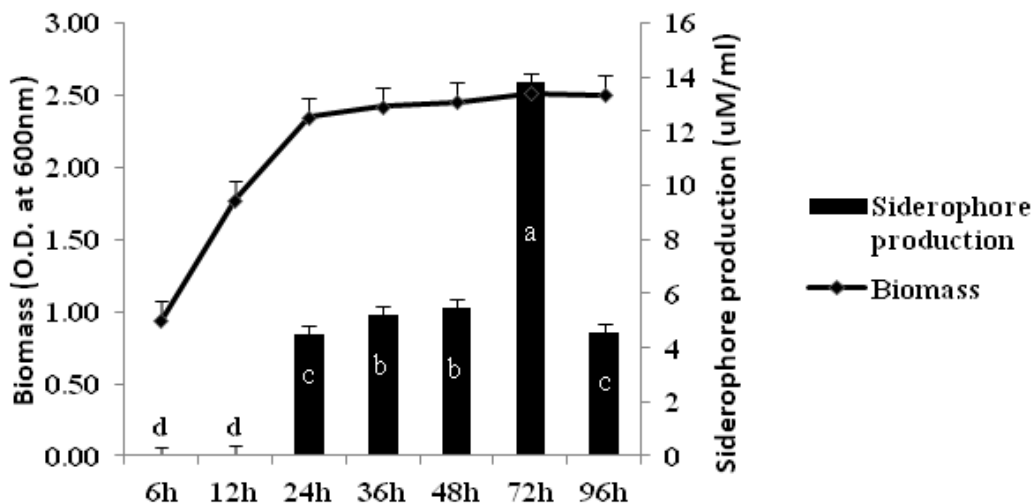


Figure 4f. Temporal growth measurement and siderophore production of FPK10 as a function of time. Vertical bar shows critical difference values. Values are mean of three replications. A common letter (letters on the bars) means they are not significantly different ($p=0.05$) by DMRT.

Table 5. Influence of different ferric ion concentration on siderophoregenesis (μM benzoic acid/mL).

Fluorescent pseudomonad isolates	Siderophoregenesis (μM benzoic acid/mL)				
	0.5 μM FeCl_3	1.5 μM FeCl_3	2.5 μM FeCl_3	5 μM FeCl_3	10 μM FeCl_3
MFP9	0.53 ^e	0.20 ^c	0.00 ^e	0.00 ^e	0.00 ^e
CHFP23	6.81 ^b	5.92 ^a	5.71 ^a	4.48 ^a	2.62 ^a
TFP16	5.84 ^c	5.62 ^a	5.16 ^b	3.04 ^c	1.95 ^b
FPK7	4.76 ^d	4.49 ^b	3.73 ^d	3.02 ^c	1.60 ^c
FPK8	11.33 ^a	4.46 ^b	4.25 ^c	3.53 ^b	2.45 ^a
FPK10	5.18 ^{cd}	4.46 ^b	3.53 ^d	2.17 ^d	1.15 ^d
SEM (\pm)	0.216	0.198	0.104	0.115	0.080
CD ($p=0.05$)	0.683	0.624	0.329	0.364	0.255

Values are mean of three replications. A common letter (on the superscript of each value) means they are not significantly different ($p=0.05$) by DMRT.

there are several reports of *Pseudomonas aeruginosa* and *Vibrio anguillarum* regarding positive regulatory mechanisms of siderophore producing genes and iron uptake systems by a combination of positive regulatory proteins and cognate siderophore but the details are under study (Chen et al., 1996; Crosa, 1997; Heinrichs and Poole, 1996; Brandel et al., 2012).

Invitro antagonistic effect on mycelial growth of phytopathogen under different iron limiting conditions

All the isolates were tested for *in vitro* siderophore mediated antagonism at different iron (Fe^{3+}) concentration

against *R. solani* and *S. rolfsii*. All the six isolates showed percent inhibition, to some extent, at all the different iron concentrations. Although, the present study argued that percent inhibition and or zone of inhibition decreased with increasing iron concentration when all the other parameters such as growth temperature, incubation period, defined media compositions are constant (Kumar et al., 2002; Sharma and Johri, 2003).

At 0.5 μM iron concentration, the percent inhibition of mycelial growth of *S. rolfsii* and *R. solani* for FPK8 were 71.1 and 73.3% respectively, which are the maximum among all the isolates, whereas, the same was 52.2% at 10 μM concentration of iron. The similar phenomenon is true with regard to the other isolates (Table 6a and b). It is apparent from the data that the siderophore mediated

Table 6a. Siderophore mediated *in vitro* antagonism against *S. rolfsii* at different iron concentration.

Fluorescent pseudomonad isolates	Different concentration of ferric ion									
	0.5 (µM)		1.5 (µM)		2.5 (µM)		5 (µM)		10 (µM)	
	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)
MFP9	1 ^c	63.3 ^d	0.6 ^b	58.9 ^c	0.2 ^d	52.2 ^e	0.0 ^d	44.4 ^e	0.0 ^b	38.9 ^e
CHFP23	1.5 ^a	67.8 ^c	0.9 ^a	60.0 ^c	0.7 ^a	55.6 ^c	0.5 ^a	51.1 ^c	0.2 ^a	43.3 ^c
TFP16	1.2 ^b	57.8 ^e	1.0 ^a	55.6 ^d	0.4 ^c	53.3 ^d	0.2 ^c	46.7 ^d	0.0 ^b	41.1 ^d
FPK7	0.7 ^d	71.1 ^a	0.5 ^b	68.9 ^a	0.2 ^d	64.4 ^a	0.0 ^d	58.9 ^a	0.0 ^b	52.2 ^a
FPK8	0.6 ^d	68.9 ^b	0.5 ^b	66.7 ^b	0.5 ^b	61.1 ^b	0.3 ^b	56.7 ^b	0.2 ^a	50.0 ^b
FPK10	0.3 ^e	53.3 ^f	0.2 ^c	47.8 ^e	0.0 ^e	44.4 ^f	0.0 ^d	37.8 ^f	0.0 ^b	33.3 ^f
SEM (±)	0.057	0.254	0.054	0.409	0.019	0.043	0.011	0.136	0.017	0.107
CD (p = 0.05)	0.181	0.803	0.172	1.288	0.060	0.136	0.037	0.430	0.054	0.338

Values are mean of three replications. A common letter (on the superscript of each value) means they are not significantly different (p= 0.05) by DMRT.

Table 6b. Siderophore mediated *in vitro* antagonism against *R. solani* at different iron concentration.

Fluorescent pseudomonad isolates	Different concentration of ferric ion									
	0.5 (µM)		1.5 (µM)		2.5 (µM)		5 (µM)		10 (µM)	
	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)	Zone of inhibition (cm)	Inhibition (%)
MFP9	1.3 ^b	64.4 ^e	1.2 ^b	61.1 ^e	1.0 ^a	58.9 ^e	0.7 ^b	56.7 ^d	0.4 ^b	47.8 ^c
CHFP23	1.6 ^a	75.6 ^a	1.3 ^a	73.3 ^a	+0.9 ^a	71.1 ^a	0.7 ^b	67.8 ^a	0.5 ^a	52.2 ^a
TFP16	1.3 ^b	67.8 ^d	1.2 ^b	64.4 ^d	1.0 ^a	62.2 ^d	0.8 ^a	58.9 ^c	0.4 ^b	50.0 ^b
FPK8	0.8 ^c	73.3 ^b	0.6 ^d	72.2 ^b	0.4 ^b	65.6 ^b	0.2 ^d	60.0 ^b	0.0 ^d	52.2 ^a
FPK7	0.8 ^c	72.2 ^c	0.7 ^c	67.8 ^c	0.5 ^b	63.3 ^c	0.4 ^c	58.9 ^c	0.2 ^c	52.2 ^a
FPK10	0.4 ^d	55.6 ^f	0.2 ^e	54.4 ^f	0.0 ^c	51.1 ^f	0.0 ^e	43.3 ^e	0.0 ^d	34.4 ^d
SEM (±)	0.012	0.176	0.013	0.136	0.045	0.133	0.021	0.170	0.021	0.047
CD (p = 0.05)	0.039	0.556	0.042	0.430	0.141	0.421	0.067	0.536	0.068	0.148

Values are mean of three replications. A common letter (on the superscript of each value) means they are not significantly different (p= 0.05) by DMRT.

antagonism may be one of the important characters for the fluorescent pseudomonas under study.

Conclusion

Based on these results, it can be concluded that the production of different and potent antifungal hydroxamate and catecholate siderophore by fluorescent pseudomonads against *R. solani* and *S. rolfsii* are being reported and could be effective bio-control agent against these phytopathogens. Reduction of antagonism by the addition of iron indicated that siderophores were inhibitory to fungal growth and showed a significant effect. Though different mechanisms might be responsible for the inhibition of *R. solani* and *S. rolfsii* but siderophore pro-

duction might be the main bio-control mechanism associated with the antagonistic potentiality of the rhizobacterial isolates of West Bengal.

ACKNOWLEDGEMENT

We, hereby, express our sincere thanks to AICRP on Vegetable Crops, Directorate of Research, B.C.K.V., Kalyani, West Bengal, India.

REFERENCES

- Adhikari A, Dutta S, Nandi S, Bhattacharya I, DeRoy M, Sarkar G, Mandal T (2013). Antagonistic potentiality of native rhizobacterial isolates against root rot disease of okra, incited by *Rhizoctonia solani*. Afr. J. Agric. Res. 8(4):405-412.

- Adilakshmi T, Ayling PD, Ratledge C (2000). Mutational analysis of a role for salicylic acid in iron metabolism of *Mycobacterium smegmatis*. J. Bacteriol. 182:264-271.
- Ali SS, Vidhale NN (2011). Evaluation of Siderophore produced by Different clinical Isolate *Pseudomonas aeruginosa*. Int. J. Microbiol. Res. 3(3):131-135.
- Arnou LE (1937). Colorimetric determination of the components of 3,4-dihydroxyphenylalanine tyrosine mixtures. J. Biol. Chem. 118:531-537.
- Bossis E, Lemanceau P, Latour X, Gardan L (2000). The taxonomy of *Pseudomonas fluorescens* and *Pseudomonas putida*: current status and need for revision. Agronomie 20:51-63.
- Brandel J, Humbert N, Elhabiri M, Schalk IJ, Mislin GLA, Gary AMA (2012). Pyochelin, a siderophore of *Pseudomonas aeruginosa*: Physicochemical characterization of the iron(III) copper(II) and zinc (II) complexes. Dalton. Trans. 41:2820-2834.
- Braun V (2001). Iron uptake mechanisms and their regulation in pathogenic bacteria. J. Med. Microbiol. 291:67-79.
- Budzikiewicz H (1993). Secondary metabolites from fluorescent *Pseudomonas*. FEMS. Microbiol. Rev. 104:209-228.
- Buyer JS, Leong J (1986). Iron transport mediated antagonism between plant growth promoting and plant deleterious *Pseudomonas* strains. J. Biol. Chem. 261:791-794.
- Chen Q, Wertheimer AM, Tolmasky ME, Crosa JH (1996). The AngR protein and the siderophore anguibactin positively regulate the expression of iron-transport genes in *Vibrio anguillarum*. Mol. Microbiol. 22:127-134.
- Crosa JH (1997). Signal transduction and transcriptional and posttranscriptional control of iron-regulated genes in bacteria. Microbiol. Mol. Biol. Rev. 67:319-336.
- Dave BP, Dube H (2000). Detection and chemical characterisation of siderophores of rhizobacterial fluorescent pseudomonads. Ind. Phytopathol. 53:97-98.
- Gillan AH, Lewis AG, Andersen RS (1981). Quantitative determination of hydroxamic acids. Anal. Chem. 53:841-844.
- Guerinot ML (1994). Microbial iron transport. Annu. Rev. Microbiol. 48:743-772.
- Gull M, Hafeez FY (2012). Characterization of siderophore producing bacterial strain *Pseudomonas fluorescens* Mst 8.2 as plant growth promoting and biocontrol agent in wheat. Afr. J. Microbiol. Res. 6(33):6308-6318.
- Heinrichs DE, Poole K (1996). PchR, a regulator of ferripyochelin receptor gene (*fptA*) expression in *Pseudomonas aeruginosa*, functions both as an activator and as a repressor. J. Bacteriol. 178:2586-2592.
- Hofte M (1993). Iron chelation in plants and soil microorganisms. (Barton, L.L. and Hemming, B.C. (eds.): Academic Press, San Diego.
- Hohlneicher U, Hartmann R, Taraz K, Budzikiewicz H (1995). Pyoverdine, ferribactin, azotobactin-a new triade of siderophores from *Pseudomonas chlororaphis* ATCC 9446 and its relation to *Pseudomonas fluorescens* ATCC 13525. Zeitschrift fur Naturforschung C. 50:337-344.
- Howard D, Rafie HR, Tiwari A, Faull KF (2000). Hydroxamate siderophores of *Histoplasma capsulatum*. Infect. Immun. 68:2338-2343.
- Jalal MAF, Vander Helm D (1990). Handbook of microbial iron chelates: Isolation and spectroscopic identification of fungal siderophores: Winklemann, G. (ed.), Pergamon Press Oxford.
- Kumar G, Sharma RC, Rai SN (2000). Biocontrol of banded leaf and sheath blight of maize by peat based *Pseudomonas fluorescens* formulation. Ind. Phytopathol. 53:190-192.
- Kumar RN, Arasu VT, Gunasekaran P (2002). Genotyping of antifungal compounds producing plant growthpromoting rhizobacteria, Mercado-Blanco J, Van der Drift KMG, Olsson PE, Thomas-Oates JE, Van Loon LC, Bakker PAHM (2001). Analysis of the *pmsCEAB* gene cluster involved in biosynthesis of salicylic acid and the siderophore pseudomonine in the biocontrol strain *Pseudomonas fluorescens* WCS374. J. Bacteriol. 183:1909-1920.
- Pseudomonas fluorescens*. Curr Sci 82:1463-1466.
- Neilands JB (1981). In: Development of iron chelator for clinical use Martell AE, Anderson WJ, Badman DG (eds.), Elsevier: Microbial iron transport compounds (siderophores) as chelating agents: North Holland:Amsterdam.
- Pal RB, Gokarn K (2010). Siderophores and Pathogenicity of Microorganisms. J. Biosci. Tech. 1 (3):127-134.
- Rachid D, Ahmed B (2005). Effect of iron and growth inhibitors on siderophores production by *Pseudomonas fluorescens*. Afr. J. Biotechnol. 4(7):697-702.
- Rao ChVS, Sachan IP, Johri BN (1999). Influence of fluorescent pseudomonads on growth and nodulation of lentil (*Lens esculentus*) in *Fusarium* infested soil. Ind. J. Microbiol. 39:23-28.
- Ratledge C, Dover LG (2000). Iron metabolism in pathogenic bacteria. Annu. Rev. Microbiol. 54:881-941.
- Reeves M, Pine L, Neilands JB, Bullows A (1983). Absence of siderophore activity in *Legionella* spp. grown in iron deficient media. J. Bacteriol. 154:324-329.
- Sambrook J, Fritsh EF, Maniatis T (1989). Molecular cloning: Laboratory manual.2nd ed. Cold Spring Harbor laboratory press. Cold Spring Harbor: NY
- Sayed RZ, Badguar MD, Sonawane HM., Mhaske MM, Chincholkar SB (2005). Production of microbial iron chelators (siderophores) by fluorescent pseudomonads. Ind. J. Biotechnol. 4:448-490
- Schwyn B, Neilands JB (1987). Universal chemical assay for the detection and determination of siderophores. Anal. Biochem. 160:47-56.
- Sharma A, Johri BN (2003). Growth promoting influence of siderophore-producing *Pseudomonas* strains GRP3A and PRS9 in maize (*Zea mays* L.) under iron limiting conditions. Microbiol. Res. 158:243-248.
- Spilker T, Coenye T, Vandamme P, LiPuma JJ (2004). PCR-Based Assay for Differentiation of *Pseudomonas aeruginosa* from Other *Pseudomonas* Species Recovered from Cystic Fibrosis Patients. J. Clin. Microbiol. 2074-2079.
- Srivastava R, Johri BN, Sharma A (1999). Colonization of wheat (*Triticum aestivum* L.) root by fluorescent pseudomonas GRP3 and PRS9. Ind. J. Microbiol. 39:205-210.
- Subashri R, Raman G, Natarajan S (2013). Biological control of pathogens and plant growth promotion potential of fluorescent Pseudomonads. Bact Agrobiol: Disease Management. 77-110.
- Weller D, Raaijmakers JM, McSpadden Gardener BB, Thomashow LS (2002). Microbial populations responsible for specific soil suppressiveness to plant pathogens. Annu. Rev. Phytopathol. 40:309-348.
- Weller DM (1988). Biological control of soilborne plant pathogens in the rhizosphere with bacteria. Annu. Rev. Phytopathol. 26:379-407.

Full Length Research Paper

Validation of the acaricidal properties of materials used in ethno-veterinary control of cattle ticks

Busani Moyo^{1*} and Patrick Julius Masika^{2*}

¹Department of Livestock and Pasture Sciences, Faculty of Science and Agriculture, University of Fort Hare, P/Bag X 1314, Alice 5700. South Africa.

²Agriculture and Rural Development Research Institute, Faculty of Science and Agriculture, University of Fort Hare, P/Bag X 1314, Alice 5700. South Africa.

Accepted 26 July, 2013

Ticks are vectors of tick-borne diseases, cause teat damage and tick-worry. They are commonly controlled using conventional acaricides, which are expensive to the resource-limited farmers, making them to resort to alternative tick control materials. The objective of this study was to validate the acaricidal properties of various ethno-veterinary materials used by rural farmers in the Eastern Cape Province of South Africa. *In vitro* repellency and contact bio-assay models were carried out to determine the repellency and acaricidal properties of *Ptaeroxylon obliquum*, *Aloe ferox* Mill, *Lantana camara* L., *Tagetes minuta*, used engine oil and Jeyes fluid on *Rhipicephalus sanguineus* ticks. The optimum repellency of Jeyes fluid at concentrations of 76.8 and 100% lasted for 6 and 7 h, respectively. Tabard the reference product lasted for 4 h. *P. obliquum* (40%) repelled the ticks for 40 min. For the contact bio-assay, used engine oil, *T. minuta* oil (50%), Ektoban[®] and Jeyes fluid (76.8%) caused tick mortality of more than 86%. This study reveals that the materials rural farmers use to control ticks vary in their efficacy. Jeyes fluid and used engine oil have acaricidal effect as the conventional acaricides whereas extracts of *A. ferox*, *L. camara* and *T. minuta* are not.

Key words: Acaricides, ethno-veterinary materials, repellency, *Rhipicephalus sanguineus*

INTRODUCTION

Ticks are one of the major factors that contribute to animal health problems and exert the greatest limitation in cattle production, by causing serious debility, morbidity, mortality and production losses in cattle (Okello-Onen et al., 2004; Moyo and Masika, 2009). They also transmit pathogens such as *Anaplasma* spp., *Babesia* spp., *Theileria* spp. and *Ehrlichia* species (Bram, 1983; L'Hostis and Seegers, 2002). The common tick species in the Eastern Cape Province of South Africa are *Rhipicephalus appendiculatus*, *Rhipicephalus evertsi* *evertsi*, *Rhipicephalus (Boophilus) decoloratus* and *Rhipicephalus sanguineus* (Marufu et al., 2010).

Ticks are commonly controlled using conventional aca-

ricides which are applied as dips or sprays and pour-on at various frequencies. Some of the conventional acaricides have repellency or acaricidal properties. Use of repellent acaricides for animal protection against ticks, constitute an important prophylactic component of tick-borne disease (TBD) management strategy (Dautel, 2004). Same applies to the contact acaricides which are chemical agents meant to kill ticks and are largely toxic through contact action (Boden and West, 1998). Some of the acaricides have long residual effect while others degrade rapidly after application (Brimecombe, 2006). Unfortunately, these chemicals are expensive, not readily available to the resources-limited farmers. In addition, the

undegradable residues contained in conventional acaricides may pollute the products (milk and meat) and the environment, and ticks have developed resistance to them (Okello-Onen and Rutagwenda, 1997). This has caused farmers to turn to low cost alternatives such as ethno-veterinary materials namely used engine oil, Jeyes fluid, *Aloe ferox* Mill and *Ptaeroxylon obliquum* (Moyo and Masika, 2009; Moyo et al., 2009).

The alternative materials like used engine oil are highly complex mixture, containing compounds distilled from petroleum, for example aliphatic hydrocarbons, aromatic hydrocarbons, polycyclic aromatic hydrocarbons [PAHs]), as well as metals such as aluminium, chromium, lead, manganese (Delistraty and Stone, 2007). Also, the Jeyes fluid a commercial disinfectant which is registered for use on non-living materials contains carbolic acid 13% m/m identified as phenol (6H5OH) and sodium hydroxide (1%) (Chem. Alert Report, 2005). However, some of these alternatives like Jeyes fluid and used engine oil are equal or more toxic than the conventional acaricides and are not the best option to be used in tick control in livestock (Aitken and Barrett, 2007; Delistraty and Stone, 2007).

In Zimbabwe for example, ethno-veterinary medicine is gaining recognition at the expense of conventional drugs especially because of its greater accessibility, lower costs and apparent effectiveness (Mwale et al., 2005) and little work on validation have been done. Many small-scale farmers are known to use ethno-veterinary practices for the control of ticks (Njoroge and Bussmann, 2006).

The purpose of this study was to validate the acaricidal properties of *P. obliquum*, *Aloe ferox*, *Lantana camara*, *Tagetes minuta*, used engine oil and Jeyes fluid on *R. sanguineus* ticks.

MATERIALS AND METHODS

Plant material collection

The leaves of *A. ferox* Mill, Voucher number BM01-037/2007, *L. camara* L Voucher No. BM01-039/2007, *T. minuta* L Voucher No. BM01-040/2007 and the bark of *P. obliquum* (Thunb) Radlk Voucher No. BM01-038/2007 were collected before flowering within Amathole District Municipality (30°15' 15" S and 22°15' 36" E), South Africa.

Preparation of the plant materials

Aqueous extraction of dried plant material

Fresh plant material (leaves of *L. camara*, *T. minuta*, *A. ferox* and the fresh bark of *P. obliquum*) were collected, air dried under shade for six weeks and then milled into powder using a hammer mill with 1 mm pore size sieve according to the farmers' description. 100 g of the respective plant materials were mixed with 1000 ml of distilled water and soaked overnight. The extracts were filtered using Whatman No.1 filter paper. The filtrate was then lyophilized using a freeze drying system Xerotec (model Cd 052, Kenmore international, Italy) for 72 h, yielding 10, 9, 11 and 12 g of extracts of *L. camara*, *T. minuta*, *A. ferox* and *P. obliquum*, respectively. The resulting extracts was stored in capped bottles and kept in the refri-

erator until use. The extracts were reconstituted using distilled water to make different concentrations of 10, 20 and 30% (w/v).

Aqueous extraction of fresh plant material

Fresh leaves of *L. camara*, *T. minuta* and *A. ferox* and the fresh bark of *P. obliquum* were collected and thoroughly washed using distilled water. Plant material of 40, 60 and 80 g were placed in 200 ml of distilled water and macerated for 1 min using an electric Sunbeam Deluxe glass blender model SGB-150, Johannesburg, South Africa.

Generally, the farmers crush the plant materials using mortar and pestle. The mixture was stored at room temperature overnight and later strained using a muslin cloth. The concentration percentages of extracts were determined using weight per volume (Jayasinghe, 1975) to obtain a 20, 30 and 40% (w/v) extract. The plant extracts were prepared as per information given by the farmers (Moyo and Masika, 2009).

Extraction of oil

Quantities (200 g) of leaves for *L. camara*, *A. ferox*, *T. minuta* and the bark of *P. obliquum* were mixed with 5 L of distilled water separately subjected to hydro distillation for 3 h using a cleveger-type apparatus (Soffiera vetro, Sassari, Italy). With the exception of *T. minuta*, the others yielded negligible quantities of oil. The *Tagetes* oil was diluted to concentrations of 3.125, 6.25, 12.5, 25, 50 and 100% using olive oil, to find out which concentration has the acaricidal activity.

Extraction of aloe leaf exudates

The aloe leaf exudates were obtained by cutting the aloe leaves and the exudates was collected, air dried and stored in bottles. It was reconstituted to the following concentrations: 1.8, 3.5, 7.5 and 15% using distilled water. Different extraction methods and concentrations of materials were used to assist in the screening of materials having acaricidal properties.

Preparation of non-plant materials

Jeyes fluid (Adcock Ingram, Bryanston, South Africa), a household disinfectant was diluted using distilled water to make the following concentrations: 0.6, 1.2, 2.4, 4.8, 9.6, 19.2, 38.4, 76.8 and 100%. Used engine oil was used undiluted. A commercial acaricides, Ektoban® (cypermethrin 2.5% w/v and cypermethrin 2.5% w/v - Novartis, Johannesburg, South Africa) registered for the control of ticks (Swan, 2001) was also used. A standard insect repellent material, Tabard (35% diethyltoluamide, Acorn (Pty) Ltd, strubens Valley, South Africa) which is effective against fleas, ticks, flies and gnats (Acorn South Africa (Pty) Ltd, 2009, personal communication) was used as a positive control.

Ticks

Adult and nymph stages of *R. sanguineus* were obtained from Clinvet International, Bloemfontein, South Africa. This tick species was selected for the study because it affects both cattle and dogs. In addition, it was the only tick species that was cultured having same age and from the same environment. The ticks were held in small transport vials supported with moist Whatman filter paper and fresh green grass. They were kept at 75% relative humidity and a temperature of 25 ±1°C, to produce an environment conducive for the survival of ticks (Thorsell et al., 2005).

***In vitro* repellency bioassay**

The repellency method described by Thorsell et al. (2005) was used in this bioassay. The nymphal stage of *R. sanguineus* tick species was used to test the repellency properties of ethno-veterinary materials (Chungsamarnyart et al., 2003). Two filter papers (Whatman No. 1) were placed inside a Petri-dish with an inner diameter of 9.5 cm. Tabard (diethyltoluamide 350 mg) an insect repellent was used as positive control while distilled water was used as negative control. Aliquots of 0.5 ml of the test solutions were applied along the periphery of the filter papers. The filter papers were then air dried for 2 min. Each test was replicated three times.

Six nymphs were placed at the centre of each of the treated filter papers and their movement with regards to avoiding the treated area was observed. If the nymph continued its motion beyond the periphery of the treated area within 5 min, the tick was indicated as non-repelled; conversely, if the nymph reversed its direction before reaching the periphery of the treated area the tick was considered as repelled. The Petri dishes were uncovered and exposed to the air from the start to the end of the experiment. The test was repeated after 0.33, 0.67, 1, 2, 3, 4, 5, 6, 7, 8 and 9 h after treatment. At each occasion, the number of nymphs avoiding the treated area was recorded. The repellency was expressed as number of nymphs avoiding treated area to the total number of nymphs at each occasion. Thus, 6 nymphs avoiding the treated area out of a total of 6 is recorded as 100% repellency. The repellency was calculated according to Thorsell et al. (2005) as follows:

$$R = \frac{a}{n} \times 100$$

Where R is the repellency; a = the number of nymphs avoiding the treated area; and n = the total number of nymphs put in the centre of filter paper at each occasion.

Contact bioassay on ticks

The dipping method was used for *in vitro* bioassay (Pirali-Kheirabadi et al., 2007). Adult *R. sanguineus* ticks were divided into groups of five and each group was immersed in a specific concentration of test samples ranging from 10, 15, 20, 30 to 40% for one minute. Distilled water and acaricide (Ektoban®) were used as negative and positive control, respectively. After dipping in the respective materials, each group was placed into separate Petri dishes containing moist Whatman No. 1 filter papers measuring 62.63 cm², with pieces of green grass to provide an environment conducive for tick survival (Thorsell et al., 2005). The tick samples were incubated for 7 days at 25°C and 75% relative humidity in the dark as described by Pirali-Kheirabadi et al. (2007) with slight modifications. Each treatment was replicated three times. The Petri dishes were examined on an hourly basis for the first 6 h after treatment, and thereafter every 24 h in the morning; to count and remove the dead ticks (Pamo et al., 2005). The tick mortality rate was calculated according to Abott (1925) and Chungsamarnyart et al. (2003) as follows:

$$\text{Corrected mortality (\%)} = \left(1 - \frac{T}{C}\right) \times 100$$

Where: T is the number of ticks remaining alive after treatment; C is the number of ticks remaining alive in the control group.

Ticks were considered alive if they exhibited normal behavior (movement looking for the host) when breathed upon or physically

stimulated with a wooden stick. For each time point, if ticks were incapable of moving, maintaining a normal posture, leg coordination, being upright themselves, or showing any sign of life, they were considered moribund or dead (Panella et al., 2005).

Statistical analysis

***In vitro* repellency bioassay**

The percentage tick repellency was calculated according to Thorsell et al. (2005). Mean repellency for all materials used was calculated.

Contact bio-assay

The following model was used to analyze tick mortality caused by treatment materials.

$$Y_{ij} = \mu + T_i + \epsilon_{ij}$$

Where Y_{ij} = response effect (mortality) due to treatment; μ = overall mean (constant); T_i = effect due to treatment ($i = 1, 2, 3, \dots$ and 32); ϵ_{ij} = residual error.

The collected data was then analyzed using PROC GLM for repeated measures (SAS, 2003). P-values < 0.05 were regarded as significant. Duncan test was used to compare differences between treatment means.

RESULTS

***In vitro* repellency bioassay**

With the exception of *P. obliquum*, the other plant extracts did not show any tick repellency effect. Only *P. obliquum* (40%) and Jeyes fluid (9.6 to 100% concentration) showed repellency as shown in Table 1. The repellency duration was shorter for *P. obliquum* (40 min) than Jeyes fluid >9.6% (1-8 h). The Jeyes fluid at 76.8 and 100% concentration was effective (repellent) for 7 and 8 h, while the positive control, Tabard was effective for 4 h after application. Extracts of *L. camara* (20 and 40%), *A. ferox* (20 and 40%), *T. minuta* (20 and 40%), *T. minuta* oil, used engine oil, olive oil and *A. ferox* leaf exudates (1.8, 3.5, 7.5 and 15%) showed no repellency. The repellency of Jeyes fluid was effective from a concentration of 2.4 to 100%.

Contact bio-assay

The plant extracts did not yield any acaricidal properties. Used engine oil, Ektoban®, *T. minuta* oil (25 and 50%) and Jeyes fluid (19.2, 38.4, 76.8, and 100%) showed variable acaricidal efficacy with the following mortalities: 93, 87, 13, 100, 26.7, 66.7, 86.6 and 100%, respectively within the first 24 h after application as shown in Table 2. *A. ferox* leaf exudates immobilized the ticks for the first 12 h and later they became active.

DISCUSSION

In all the experiments, it was clear that the nymphs displayed

Table 1. Tick repelling activity of *P. obliquum* and Jeyes fluid at different concentrations.

Material	Tick repellency (%)										
	20 min	40 min	1 h	2 h	3 h	4 h	5 h	6 h	7 h	8 h	9 h
<i>P. obliquum</i> (40%)	100	94	0	0	0	0	0	0	0	0	0
Jeyes fluid (9.6%)	100	100	72	0	0	0	0	0	0	0	0
Jeyes fluid (19.2%)	100	100	100	94	22	0	0	0	0	0	0
Jeyes fluid (38.4%)	100	100	100	100	78	22	0	0	0	0	0
Jeyes fluid (76.8%)	100	100	100	100	100	100	100	89	33	0	0
Jeyes fluid (100%)	100	100	100	100	100	100	100	100	83	28	0
Tabard (35%)	100	100	100	100	100	94	56	6	0	0	0

Table 2. Least square means for acaricidal effect of some ethno-veterinary remedies.

Material	Tick mortality (%)													Total tick mortality (%)
	1 h	2 h	3 h	4 h	5 h	6 h	1 day	2 days	3 days	4 days	5 days	6 days	7 days	
Used engine oil	73	0	0	13	7	0	93	0	0	0	0	0	7	100 ^a
<i>T. minuta</i> oil (50%)	60	40	-	-	-	-	100	-	-	-	-	-	-	100 ^a
<i>T. minuta</i> oil (100%)	100	-	-	-	-	-	100	-	-	-	-	-	-	100 ^a
Jeyes fluid (38.4%)	6.7	60	0	0	0	0	66.7	0	0	0	0	0	0	66.7 ^c
Jeyes fluid (76.8%)	73.7	13.3	0	0	0	0	86.6	0	0	0	0	0	0	86.6 ^b
Jeyes fluid (100%)	100	-	-	-	-	-	100	-	-	-	-	-	-	100 ^a
Ektoban	87	0	0	0	0	0	87	13	-	-	-	-	-	100 ^a

^{a,b,c}Means in the column with different superscript letters differ significantly, $p < 0.05$.

their natural host-seeking behavior during the tests. They moved around the filter paper and even crossed the treated areas for the extracts that did not show repellency.

The extracts of *P. obliquum* at 40% concentration had a short-lasting protection that provided 94% repellency at 40 min post-application against the nymph. This short repellency period may not be adequate to protect livestock against ticks. Tabard stick, our positive control, provided greater than 94% repellency for 4 h which was greater than the repellency period of *P. obliquum*. The

bark of *P. obliquum* has been reported to contain saptaoerxylon (an acid saponin), pyrogall, resins and an alkaloid (Archer and Reynolds, 2001) with resins reported to have repellency activities (Pontes et al., 2007).

It is speculated that the resin oil in the *P. obliquum* was in small quantities which made the repellency to last for a short period of time or it was volatile in nature making it to lose its repellency. Therefore, further research is necessary to find ways of improving the duration period of repellency. The results of the contact

bioassay are in contrast with those of Archer and Reynolds (2001), who indicated that the powder of the bark added to water kills cattle ticks. The variation could be attributed to the tick species and the age of the tree from where the bark was collected and growth environment which can influence the amount of compounds in the plant. The results obtained in this study indicate that essential oil of *T. minuta* had no repellency properties, however, it differs from the studies of Nchu et al. (2012) which showed some tick repellency against *Hyalomma rufipes*. This could

be attributed to differences in tick species used in the study.

Jeyes fluid showed some acaricidal efficacy both through contact and repellency in the study which compared well with the positive control, Ektoban® acaricide. The active ingredient of Jeyes fluid (carbolic acid) was once used as carbolic dip to control ticks more than 50 years ago (Malesela, 2004). Moreover, the carbolic acid is poisonous to tissues especially when applied directly to muscle and nerves, causing paralysis to the nerves which later paralyse the heart (Henriette, 2008) hence death of organisms.

However, the veterinarians do not recommend its use because carbolic dip has very narrow safety margins (Malesela, 2004; Aitken and Barrett, 2007). Carbolic dip caused some poisoning to animals dipped (Linklater et al., 1982; Aitken and Barrett, 2007). Moreso, its exposure in the animal skin have pathological effect (Chem Alert Report, 2005). The above reasons could have contributed to its withdrawal from the list of registered acaricides. In addition, Jeyes fluid is toxic to aquatic organisms, it alters the water pH (Irwin et al., 1997).

Jeyes fluid is locally available in shops and farmers maintain that it was cheaper as compared to the conventional acaricides and also kill ticks as reported by Malesela (2004) and Moyo et al. (2009). The residual effect of Jeyes fluid to consumers is not known. Therefore, its use should be avoided or used with caution and more research needs to be done on its toxicity.

The efficacy of used engine oil through contact bioassay compared well with the positive control conventional registered, Ektoban® acaricide in agreement with Moyo et al. (2009). This was in contrast with the field efficacy findings by Dreyer et al. (1998) where it had an average efficacy of 38.1%. The variation could be due to tick species or the field condition. The high mortality caused by used engine oil is ascribed to its action as a physical acaricide clogging the spiracles and causing the tick to suffocate (Dreyer et al., 1998). Therefore, direct contact between the oil and the tick is necessary for effective result (mortality). Mbaty et al. (2002) and Masika et al. (1997) have documented the wide use of used engine oil. However, no validation was done. The toxic components in it, could be the ones that caused the death of the ticks.

The disadvantage of using used engine oil is that it has toxic components such as lead chromium, copper and zinc which have residual effect on plant and animal tissue making them unsafe for human consumption as cited by Moyo and Masika (2009). Used engine oil can contaminate the environment and water bodies resulting in the death of aquatic and soil microbes (Delistraty and Stone, 2007). This aspect needs investigation since little/no work has been done in this regard. However, many farmers in the Eastern Cape Province depend on government pensions (Dreyer, 1997), they cannot afford to buy conventional acaricides as such, they resort to using used engine oil to control ticks on their cattle.

The efficacy of *T. minuta* oil was comparable to that of the reference conventional acaricide, Ektoban®, which is very effective against ticks. This makes *T. minuta* oil an alternative acaricide that can be used by farmers. However, the cost of the oil may be prohibitive too. The acaricidal effect could be attributed to the compound terpenes contained in *T. minuta* oil, which has been reported to have aphicidal and insecticidal properties (Sarin, 2004; Seyoun et al., 2007; Tomova et al., 2005). The effect of this oil on ticks, in this study concurs with its efficacy on other insects. The recommended concentration of *T. minuta* oil to be used as an acaricide could be 50% level. This is more economic than the 100% concentration which uses more essential oil.

Use of boiled *T. minuta* leaves kill ticks (Njoroge and Bussmann, 2006), as such farmers in the study area could benefit from boiling the leaves which is cheaper than distilling oil from the plants. The other plant extracts exhibited variable results ranging from no effect to lower mortality. Therefore, they cannot be used as alternative acaricides in tick control.

The extracts of *L. camara* at 40% concentration had a tick mortality of 27%, which was inadequate for reducing tick burdens in livestock. Our findings however are in contrast with Okello-Onen et al. (2004) and Moyo et al. (2009) where its efficacy was 96.6 and 58% for all the tick species, respectively. The difference in the observations could be due to difference in the environment where the plant was collected and age of the plant. This affects the chemical composition of the plant. Also, this variation could be ascribed to the fact that acaricidal efficacy depends on the tick species.

The results of this study have revealed that the materials rural farmers use as acaricides vary in their efficacy to control ticks. Jeyes fluid and used engine oil are as effective as the conventional acaricides whereas others are not effective at all. Jeyes fluid (38.4, 76.8 and 100%), *T. minuta* oil (50%) and used engine oil had the most acaricidal properties. However, they also need to be validated in *in vivo* experiments. Despite being effective, used engine oil and Jeyes fluid have potential toxic effects on animals and are also environmental contaminants.

ACKNOWLEDGEMENTS

The authors acknowledge the financial support from the National Research Foundation of South Africa. They also extend their thanks to ARDRI staff and the Department of Livestock and Pasture Sciences at the University of Fort Hare in South Africa for their support.

REFERENCES

- Abott WS (1925). A method for computing the effectiveness of an insecticide. *J. Econ. Entomol.* 18:265-267.

- Aitken I, Barrett T (2007). Diseases of sheep, <http://books.google.co.za/book?isbn=140513143>.
- Archer R, Reynold Y (2001). *Ptaeroxylon obliquum*_(Thunb) Radlk. South Africa.National Biodiversity institute. (<http://bold.cf.ac.uk/Bot.DermFolder/BotDermR/RUTA.html>).
- Boden E, West GP (1998). Black's veterinary Dictionary.19th edition. <http://books.google.co.za/books?isbn=038921017x>.
- Bram RA (1983). Tick-borne livestock diseases and their vectors. 1. The global problem. Wild Anim. Rev. 36:1-5.
- Brimecombe RD (2006). Voltametric analysis of pesticides and their degradation: A case study of amitraz and its degradants. M. Sc. Thesis, Rhodes University.
- Chem Alert Report (2005). [http://www.waiwhetu.co.nz/msds/jd/jeys%20fluid%20\(nz\).pdf](http://www.waiwhetu.co.nz/msds/jd/jeys%20fluid%20(nz).pdf).
- Chungsamarnyart N, Jansawan W, Tassanawat T, Thiapayarak T (2003). Efficacy of herbal Acaricide;K U Natural Extocide against the Tick, Flea, Lice of dogs. 28th World Congress of the World small animal veterinary Association (October 24-27, 2003, Bangkok, Thailand).
- Dautel H (2004). Test systems for tick repellents. Int. J. Med. Microbiol. 293:suppl. 37:182-188.
- Delistraty D, Stone A (2007). Dioxins, metals, and fish toxicity in ash residue from space heaters burning used motor oil. Chemosphere 68:907-914.
- Dreyer K (1997). Occurrence and control of parasites on cattle in urban and peri-urban environments with specific reference to ticks. M.Sc. Thesis, University of Orange Free State.
- Dreyer K, Fourie LJ, Kok DJ (1998). The efficacy of used engine oil against ticks on cattle. Onderstepoort J. Vet. Res. 65:275-279.
- Henriette K (2008). <http://www.henriettesherbal.com-copyright1995-2008HenrietteKress>.
- Irwin RJ, Mouwerick MV, Stevens L, Seese MD, Basham W (1997). Environmental contaminants encyclopedia. (<http://www.fws.gov/caribbean/es/PDF/contaminants/coaltar.pdf>).
- Jayasinghe DM (1975). Bhaysha Kalpana Paribhashava: in Ayurveda Pharmacopoeia, vol. 1. Department of Ayurveda, Sri Lanka, Colombo, Sri Lanka. p. 30.
- L'Hostis M, Seegers H (2002). Tick-borne parasitic diseases in cattle: current knowledge and prospective risk analysis related to the ongoing evolution in French cattle farming systems. Vet. Res. 33:599-611.
- Linklater A, Angus KW, Mitchell B, Spence JA, Rowland AC, Hunter AR (1982). Pneumonia in sheep associated with dipping in carbolic dips. Vet. Rec. 110:33-36.
- Malesela JS (2004). Assessing implementation of veterinary Extension on control of cattle parasites, in Moretele District, North West Province, South Africa. (unpublished Thesis, University of Pretoria).
- Marufu MC, Chimonyo M, Dzama K, Mapiye C (2010). Seroprevalence of tick-borne diseases in communal cattle reared on sweet and sour rangelands in a semi-arid area of South Africa. Vet. J. 184(1):71-76.
- Masika PJ, Sonandi A, van Averbeke W (1997). Tick control by small-scale cattle farmers in the central Eastern Cape Province, South Africa. J. S. Afr. Vet. Assoc. 68:45-48.
- Mbati PA, Hlatshwayo M, Mtshali MS, Mogaswane KR, deWaal TD, Dipeolu OO (2002). Tick and tick borne diseases of livestock belonging to resource poor farmers in the Eastern Free State of South Africa. Springer Netherlands. 28:217-224.
- Moyo B, Masika PJ (2009). Tick control methods used by resource-limited farmers and the effect of ticks on cattle in rural areas of the Eastern Cape Province, South Africa. Trop. Anim. Health Prod. 41:517-523.
- Moyo B, Masika PJ, Dube S, Maphosa V (2009). An *in-vivo* study of the efficacy and safety of ethno-veterinary remedies used to contro cattle ticks by rural farmers in the Eastern Cape Province of South Africa. Trop. Anim. Health Prod. 41:1569-1576.
- Mwale M, Bhebhe E, Chimonyo M, Halimani TE (2005). Use of herbal plants in poultry health management in the Mushagashe small-scale commercial farming area in Zimbabwe, Int. J. Appl. Res. Vet. Med. 3:163-170.
- Nchu F, Magano SR, Eloff JN (2012). In vitro anti-tick properties of the essential oil of *Tagetes minuta* L. (Asteraceae) on *Hyalomma rufipes* (Acari: Ixodidae). Onderstepoort J. Vet. Res. 79(1):E1-5.
- Njoroge GN, Bussmann RW (2006). Herbal usage and informant consensus in ethnoveterinary management of cattle diseases among the kikuyus (Central Kenya). J. Ethnopharmacol. 108:332-339.
- Okello-Onen J, Kerwegi SA, Ollila D, Ssekitto CCMB, Osinde C (2004). Participatory Evaluation and Improvement of ethno-veterinary practices for tick control in Lango and Teso Farming System. Final technical report.
- Okello-Onen J, Rutagwenda T (1997). A Study on the status of East coast-fever, causes of calf mortality and abortion in Ankole cattle in ankole ranching scheme, Mbarara District, Uganda. Final Report, GTZ project, Sanga, Mbarara. p. 36.
- Pamo TE, Tendonkeng F, Kana JR, Payne VK, Boukila B, Lemoufouet J, Miegoue E, Nanda AS (2005). A study of the acaricidal properties of an essential oil extracted from the leaves of *Ageratum houstonianum*. Vet. Parasitol. 125:319-323.
- Panella NA, Dolan MC, Karchesy JJ, Xiong Y, Peralta-Cruz J, Khasawneh M, Montener JA, Maupin GO (2005). Use of novel compounds for pest control, Insecticidal and Acaricidal activity of Essential Oil Components from Heartwood of Alaska Yellow Cedar. J. Med. Entomol. 42:352-358.
- Pirali-Kheirabadi K, Razzaghi-Abyaneh M (2007). Biological activities of chamomile (*Matricaria chamomile*) flowers's extract against the survival and egg laying of the cattle fever tick (Acari Ixodidae) J. Zhejiang Univ. Sci. B. 8:693-696.
- Pontes WJT, de Oliveira Jose CS, Camara Claudio AG, Lopes Adelmo CHR (2007). Composition and acaricide Activity of Resin's Essential Oil of *Protium batianum daly* against two spotted spider mite (*Tetranychus urticae*): JEOR(http://findarticles.com/p/articles/im_qa4091/is_200707/ai_n19434104).
- Sarin RC (2004). Insecticidal activity of callus culture of *Tagetes erecta*, Fitoterapia 75:62-64.
- Seyoun A, Kabirru EW, Lwande W, Killen DF, Hassanali A, Knols BG (2002). Repellency of live potted plants against *Anopheles gambiae* from human baits in semi-field experimental huts. Am. J. Trop. Med. Hyg. 67:191-195.
- Statistical Analyses System (SAS) (2003). Statistical analysis system user's guide (5 th Edition), version 6, (SAS Institute Inc., Raleigh, North carolina, USA).
- Swan G (2001). Index of veterinary specialities, Vol. 38. No. 3, September- November 2000.
- Thorsell W, Mikiver A, Tunon H (2005). Repelling properties of some plants materials on the tick *Ixodes ricinus* L. Phytomedicine 13:132-134.
- Tomova BS, Waterhouse JS, Doberski J (2005). The effect of fractionated *Tagetes* oil volatiles on aphid reproduction Entomol. Experimentalis. et Applicata 115:153-159.

Full Length Research Paper

Effect of different ferric fertilizers on planting *Morchella conica* fruiting yields and analyses of the microflora and bioactivities of its grown soil

Xue Li¹, Zizhu Chen, Chenchen Peng¹, Yu Han², Jianguang Xu², Zhangfu Long^{1*}

¹Key Laboratory of Bio-resources and Eco-environment (Ministry of Education), College of Life Sciences, Sichuan University, Chengdu 610064, P.R. China.

²Sichuan Tongfeng Science and Technology Co. Ltd, Chengdu 610072, P.R. China.

Accepted 19 April, 2013

The aims of this study were to determine whether six ferric fertilizers influence the development of fruit bodies of *Morchella conica* identified by analysis of the internal transcribed spacer (ITS), its grown soil microflora and enzymatic activities. We successfully cultivated one kind of *Morchella* in the wheat planting field, its partial ITS rDNA in the present study was identified as *M. conica* by GenBank and Morchella MLST database as a reference. Six ferric fertilizers were applied to the planting area. Four soil enzymatic activities (including soil urease, polyphenol oxidase, invertase, and protease) as well as soil microflora showed significant differences in the development stages of *M. conica* especially during the primordium differentiation stage. Results suggest that ferric fertilizer not only influenced the *Morchella* grown soil enzymatic activities and microorganisms, but also increased the *Morchella* yields while such effects differ from the species of the ferric fertilizer, especially groups with ammonium ferric sulfate [NH₄Fe(SO₄)₂·12H₂O], Fe-EDDHA and Tongfeng nutrients. So, this work highlights the importance of further attempts to resolve important aspects of the morel commercial cultivation regarding optimizing nutrition plan and figuring out the relationships between genotypes and their fertilities of fruiting body.

Key words: *Morchella conica*, identification, ferric fertilizer, microflora, enzyme activity.

INTRODUCTION

In the world's 1.5 million estimated fungi species (Hawksworth, 2004), few genera are as synonymous with epicurean cuisine as true morels (Du et al., 2012b), highly prized for their medicinal and nutritional qualities (Kanwal and Reddy, 2012). Fruiting bodies of morel fungi (*Morchella* spp., phylum Ascomycota) are highly restricted to tem-

perate regions of the Northern Hemisphere in a few weeks each spring (O'Donnell et al., 2011). Due to the increasing popularity and commercial demand, wild morels are harvested commercially and exported extensively from China, India, Turkey, Mexico, and the United States (Pilz et al., 2007; Taskin et al., 2010).

*Corresponding author. E-mail: Lzf0028@163.com.

Abbreviations: **ITS**, Internal transcribed spacer; **CTAB**, hexadecyltrimethyl-ammonium bromide; **BLAST**, Basic Alignment Search Tool; **MLST**, multilocus sequence typing; **BS**, bootstrap; **PDA**, potato dextrose agar; **NCBI**, National Center for Biotechnology Information; **DGGE**, denaturing gradient gel electrophoresis.

The first successfully growing morel fruit body under controlled condition was carried out by cultivating in a specialized indoor facility using patented technology within the US (Ower et al., 1986), included outdoors in Yunnan Province of China (Zhao et al., 2009).

Morels' very short harvesting season and high price have encouraged researchers to study the biology and ecology of the species (Goldway et al., 2000). Many researchers have attempted to raise morels in controlled conditions; nevertheless, ascocarps reaching the market are mostly collected from the wild (Ower et al., 1986; Goldway et al., 2000; Masaphy, 2005; Guler and Ozkaya, 2009; Masaphy, 2010). Their modes of reproduction, biology, nutritional sources, life cycle, are unusual and complex (David et al., 2007). Recently, a remarkable finding is that the majority of the Esculenta clade appeared to be adapted to mixed temperate deciduous hardwood forests with the Elata clade's preference to an evergreen coniferous biome (Du et al., 2012a; Donoghue, 2008). By all odds, it is important showing the morels growing conditions.

Previous reports are almost focused on their mycelial fermentation or officinal functions, little on their cropping stages investigation. The mating systems of *Morchella* species are unknown (Buscot, 1993), and whether heterokaryosis (mating of two compatible strains) followed by meiosis occurs in natural populations of morels is also unknown (Volk and Leonard, 1989, 1990). Attention has focused on antioxidant activities of produced compounds, content of phenolics and flavonoids, autofluorescence and various metals (Gursoy et al., 2009; Kalač 2010). Many studies have demonstrated that minerals and heavy metals play an important role in the metabolic processes, during the growth and development of mushrooms in appreciable concentration (Claudia et al., 2010) and wild-growing mushrooms can accumulate great concentrations of toxic metallic elements and metalloids such as mercury, cadmium, lead, copper or arsenic and zinc (Vetter, 2004; Nevcihan et al., 2009; Zhu Fangkun et al., 2011). Reports pointed out that the accumulation of metals in fungi found to be affected by element environment such as organic matter amount, temperature, humidity, pH, metal concentrations in soil and fungal factors such as species of microfungi, morphological part of fruiting body, development stages and age of mycelium, biochemical composition, and interval between the fructifications affect metal accumulation in macrofungi (Mendil et al., 2005; Nevcihan et al., 2009).

It has also been shown that Fe was abundant in seven *Morchella* species, follow by calcium and magnesium, respectively (Nevcihan et al., 2009). Iron can stimulate lipid peroxidation by the fenton reaction, and also accelerates peroxidation by decomposing lipid hydroperoxides into peroxy and alkoxy radicals that can abstract hydrogen and perpetuate the chain reaction of lipid peroxidation (Halliwell, 1991). As the most effective pro-oxidants, ferrous

ions are commonly found in food systems (Yamaguchi et al., 1998).

Based on our previous study, we successfully cultivated *Morchella* in the wheat planting field. Up to now, the formation of fruit-bodies of morchella is still mysterious and their commercial production is not mature. Commercially cultivation of morel has also been confronting with series of difficulties. Many triggers for fruiting appear to differ by species to some extent. We still do not know which vital factor determines that the transformation from sclerotium of *Morchella* to primordium of fruit bodies, such as soil microorganisms or microelements or other compounds, even the differences existing in genomics between fertile and infertile morchella strains.

In the present paper, our aims are to identify the morels using internal transcribed spacer (ITS) data with *Morchella* MLST as a reference and the effect of the different ferrous treatments on soil characteristics and the morel yields, in order to determine the possibility of optimal morels growing conditions.

MATERIALS AND METHODS

Strains and chemical agents

pMD18-T Vector Ligation Kit, DL2000 DNA Marker was purchased from TaKaRa Technology (Dalian) Co., Ltd. DNA Gel Extract Kit was purchased from MBI Fermentas. *Morchella conica* strain Mc-5 (commercially cultivated for five years), *Escherichia coli* DH5 α and BL21 (DE3) strains were preserved in author's Laboratory. All other reagents except the noted agents used in present study were of analytical grade.

Molecular identification of *Morchella* strain and its sclerotia observation by scanning electron microscopy (SEM)

Total genomic DNA of *M. conica* was followed by a hexadecyl-trimethyl-ammonium bromide (CTAB) protocol (O'Donnell et al., 1997; Taşkin et al., 2010) when the mycelium was freeze-dried before the CTAB extraction step. The internal transcribed spacer (ITS) rDNA sequence was amplified using the total DNA as template while the primer pair of ITS1 and ITS4 was used as the primer pair, upstream primer (ITS1: 5'-TCCCTTACTCTTCTAACCTCTCCTCT-3') and downstream primer (ITS 4: 5'-TCCTCCC TCTTATTCTATATCTC-3'). Polymerase chain reaction (PCR) products were purified with the DNA Extraction Kit (MBI Ferments). The rDNA fragment was T-A cloned into the pMD18-T vector (Takara Shuzo Co., Ltd., Dalian, China) and propagated in *E. coli* (strain DH5 α).

The positive clone was screened by Clone-PCR method (Gussow and Clackson, 1989; Sathé et al., 1991). The ITS sequences of the isolates was determined by Shanghai Jikang Biotech Company (Shanghai, China), then used as templates to search for homologous sequences in GenBank using the Basic Alignment Search Tool (BLAST) and *Morchella* MLST database as a reference (Robert et al., 2011). The obtained sequences were compared one to another by the computer-assisted method of Higgins and Sharp (1988). The phylogenetic relationships within the Morchellaceae using a aligned nucleotide positions in Paup via one thousand Maximum parsimony bootstrap (BS).

Experimental strain of *M. conica* Mc5 was grown in potato dextrose agar (PDA) and its sclerotia was harvested and fixed for 2 h at 4°C by using 2.5% (v/v) glutaraldehyde in 0.1 M sodium cacodylate buffer. Secondary fixation was performed in 1% osmium tetroxide in dH₂O after the samples were washed three times with 0.1 M cacodylate buffer (pH 7.4), and were then treated with 1% (w/v) osmium tetroxide, washed with 5% (w/v) sucrose in cacodylate buffer, and subsequently dehydrated in a graded ethanol series (from 30 to 100%). The samples were examined on a JEOL JSM-7500F Scanning Electron Microscopy after the works for critical point drying and gold coating were successfully carried out and the images were recorded.

Cultivation Experimental groups sprayed with ferric fertilizers

First fruit bodies pieces collected for pure *M. conica* culture isolation, sprayed with 70 % ethanol, were grown well in the PDA (potato 20 g/L, glucose 2g/L, agar 2g/L) test-tubes, then sub-culture of isolates were removed in another new sterilized medium which consists of wheat 70%, vermiculite 5%, pure soil 15% and chaff 10%, natural pH value and cultured under 15-18°C since mid-October for over 15 days, when morel sclerotia were formed well.

We choose Longxing Town (located in Chongzhou, Sichuan Province, China) where hardwood trees grow naturally and soil is sandy loam or humus without much clay. The climate is subtropical with annual precipitation and average temperature of 1012 mm and 17.8°C. Each section was 10x0.6 m². Each treatment was established with three replicates. Seeding of *Morchella* spawn was carried out during early November and about 1.5 bottle (750 mL/bottle) spawn were used per square meter (m²) when the planting field tamped and leveled up firmly. About twenty days later after seeding in December, six experimental groups, including Iron II ethylenediamine di(*o*-hydroxyphenylacetic) acid (abbreviated as Fe-EDDHA, provided by Sichuan Tongfeng Science and Technology Co. Ltd, Chengdu, China), ferrous sulfate (FeSO₄·7H₂O), ferric sulfate [Fe₂(SO₄)₃·9H₂O], ammonium ferric sulfate [NH₄Fe(SO₄)₂·12H₂O], citrate ferric, Tongfeng nutrition agents (abbreviated as nutrients, also provided by Sichuan Tongfeng Science and Technology Co. Ltd, Chengdu, China) with six trace elements (including with Fe, Zn, Mn, Mo, B, Cu, and its ferric content was about 2%), were separately sprayed onto the soil sampling districts grown with *M. conica*, and all the nutrition concentration of different ferric fertilizers was 100 mg/m² when *Morchella* mycelium grown well in the field before sclerotia formation, while the same volume of distilled water was sprayed as the control. Primordia formation started as temperatures increase above 10°C (a range between 6-15°C), lasting for one month since late January. The fruiting bodies appear approximately in the early February, obviously influenced by the soil temperature, and the fruiting season is no better than 20 days, compared with wild morels in ten days in other reports (Mihail et al., 2007; Wurtz et al., 2005).

Enumeration of major soil microbial population groups

Soil samples were randomly collected aseptically in ultra-violet (UV) sterilized plastic bags from each groups to a depth of 10–30 cm during sclerotia formation, primordia formation and fruit-body growth separately in late November, late January and early February. The soil samples were kept on ice during transport to the laboratory and stored at 4°C before microbial enumeration. The enumeration of the soil microflora was done by the dilution plate method (Nair and Subba-Rao, 1977). Soil sampling (10 g) was done from three replicated sites and emulsified in 90 mL sterilized water, aseptically. A serial decimal dilution was made from this suspension up to 1:10⁻¹⁰.

1 mL of each dilution was used to inoculate plates in triplicate containing specific growth media for different microorganism. The total colony forming units (cfu) of bacteria, fungi and actinomycetes were recorded on Gauze No. 1 agar (for actinomycetes, Ye et al., 1983), Martin's rose bengal streptomycin agar agar (for fungi, Martin, 1950) and Jensen's agar (for bacteria, Jensen, 1951) media, respectively. There were nine plates for each sample, and the plates were incubated at 28°C for 2–4 days for bacteria, 5–8 days for fungi, and 11–14 days for actinomycetes; after, microbial population was calculated and expressed as 10ⁿ cfu/g air dried soil, where 10ⁿ was dilution factor.

Determination of enzymatic activity for *Morchella* grown soil

Soil samples were collected on the same time as the enumeration of major soil microbial population. The samples for the determination of enzyme activities were sieved (1 mm) after being air-dried at room temperature for 7 days and stored at 4°C prior to analysis. The studied soil enzymes were invertase, urease, polyphenol oxidases and proteases. Invertase activity in soil was assayed by measuring the release of reducing sugar (Papa et al., 2010). Soil urease activity was determined by the method of Tabatabai and Bremner (1972) and protease activity by the method of Ladd and Butler (1972) using gelatin as the substrate. Polyphenol oxidase activity was measured by absorbance at 420 nm using the method described by Kar and Mishra (1976) and pyrogallol used as the substrate.

Statistical analysis

All the experimental results were means±SD and statistical significances of differences among treatments were determined by using the Statistical Package for the Social Sciences (SPSS) (version 13), and followed by comparisons at significance level of 0.05 (P<0.05).

RESULTS AND DISCUSSION

Morchella strain identification and its sclerotia observation by SEM

The edible mushroom *Morchella esculenta* Pers. forms sclerotia, whose importance in its life cycle was demonstrated by Ower et al. (1986). Amir et al. (1995) compared nine inhibitors affecting translocation and sclerotia formation in *M. esculenta* and they found that the use of nikkomycin inhibited sclerotia formation, without affecting translocation to the sclerotia. Therefore, it is very important to qualify the formation of sclerotia for the experimental strain. Results of scanning electron microscopy (SEM) had shown that *Morchella* sclerotia were formed by tube-shape mycelium and looked like granules (Figure 1). From the photos shown in Figure 1, we found that the sclerotia of *M. conica* were about 15±3.24 µm in diameter, and its mycelium about 10.25±2.13 µm in diameter. Some mycelia were observed to create plasma bridges and such results were reported by others (Guler and Ozkaya, 2008; Arkan 1992). Volk and Leonard (1990) reported that one area of neglect is the cytology of various development stages of the *Morchella* life cycle: the vegetative hyphae, sclerotia and fruiting bodies. Guler and Ozkaya, (2008)

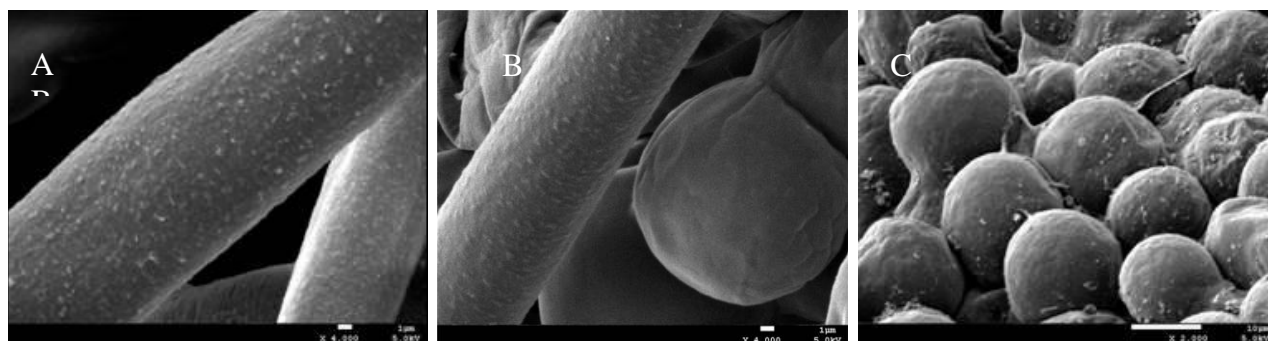


Figure 1. Mycelium and sclerotia observation by Scanning emission microscope for *Morchella conica*. Figure 1A, mycelium of *M. conica* observed by SEM with the mycelium shaped as a kind of tube coated with small particles. Barrel-shaped cell in sclerotia. Bar= 1 μ m. Figure 1B, sclerotia of *M. conica* were about 15 ± 3.24 μ m in diameter, and its mycelium about 10.25 ± 2.13 μ m in diameter. Bar= 1 μ m. Figure 1C, round, closely packed basal sclerotia.

investigated the effects of various carbon sources for *M. conica* sclerotium formations under in vitro conditions and found sclerotia cells were generally very thick and spherical. It was thought that the angle of hypha growth was something collected with sclerotia in culture medium, but this needs further confirmation.

Former species-level molecular systematic markers employed within this genus were limited to isozymes (Gessner et al., 1987; Royle and May, 1990; Wipf et al., 1996), restriction fragment polymorphisms of the LSU rDNA (Bunyard et al., 1994, 1995), ITS region of the nuclear rDNA (Buscot et al., 1996), and phylogenetic analyses of the ITS rDNA sequence data (Wipf et al., 1999; Kellner et al., 2005). Using multilocus DNA sequence data and phylogenetic species recognition (Dettman et al., 2003; Taylor et al., 2000), researcher investigated DNA sequences from marker loci, including ITS rDNA, and partial sequences of LSU rDNA, RPB1, RPB2, and EF-1 alpha to study morels species limits and to assess evolutionary relationships (O'Donnell et al., 2011; Tas-kın et al., 2010, 2012; Du et al., 2012a). Du's research also found that ITS sequence data has limited utility in differentiating species within the species-rich *Elata* sub-clade in the identified three lineages within *Morchella* (Du et al. 2012b) and most *Morchella* species appear to exhibit continental endemism and provincialism (O'Donnell et al., 2011). Because two-thirds of the *Morchella* ITS rDNA sequences identified to species in GenBank are misidentified and the ITS rDNA partition still was the most length variable (Du et al., 2012a), the *Morchella* strain in the present study was identified by *Morchella* MLST database in preference to NCBI GenBank numbers. Therefore, the studied strain was authenticated by analyzing its partial ITS rDNA sequences (750bp, Wipf et al., 1996) and the homologies analyses with *Morchella* MLST database with GenBank as a reference. Results are shown in (Figure 2).

The ITS sequences of experimental strain matched *M.*

conica, with best matches characterized by minimum 98.5 % homologies to DQ257343 (*M. conica*, China) and maximum 100% homologies to AJ544198 (*M. conica*, North Israel; Kellner et al., 2007) with NCBI. 13 homologous sequences within *Elata* clade were analysed, it is apparent that all of the black species of *Morchella* grouped closely. It shares 98.71 homologies to that of JF908365 (*M. conica*, Italy) within BioloMICS software (Robert et al., 2011), significantly, which shares 100% homologies to that of AJ544198 (*M. conica*, North Israel), shares over 99% homologies to EF080999 (99.9%; *M. conica*, China), AJ544194 (99.9%; *M. conica*, Germany), GQ265899 (99.7%; *Morchella costata*, China), AM269501 (99.7%; *M. conica*, Poland). These results obviously agree that variation may occur between geographically isolated populations of the same species (Bunyard et al., 1994).

ITS rDNA sequences give some information to analyses the commercial cultivation strain. Conducting these sequences, either through the species identification or possibly establish species DNA barcoding, we can better contribute to more suitable breeding in future. From above results, we found a very interesting phenomenon, that the present study strain of *M. conica* are just the same as another strain sampled from North Israel when their ITS rDNA sequences were compared, which suggested disjunct distribution during their evolutionary relationships.

Enumeration of major microbial populations from grown soil for *M. conica*

Up to now, we know little about the changes of the microorganisms from the cultured morel soils, especially treated with microelements such as ferric, zinc, among others. Microorganisms require iron for growth. Under iron-limited conditions, low molecular weight compounds called siderophores are synthesised by microbes (Varma and Chincholkar, 2007). Siderophores chelate ferric ions

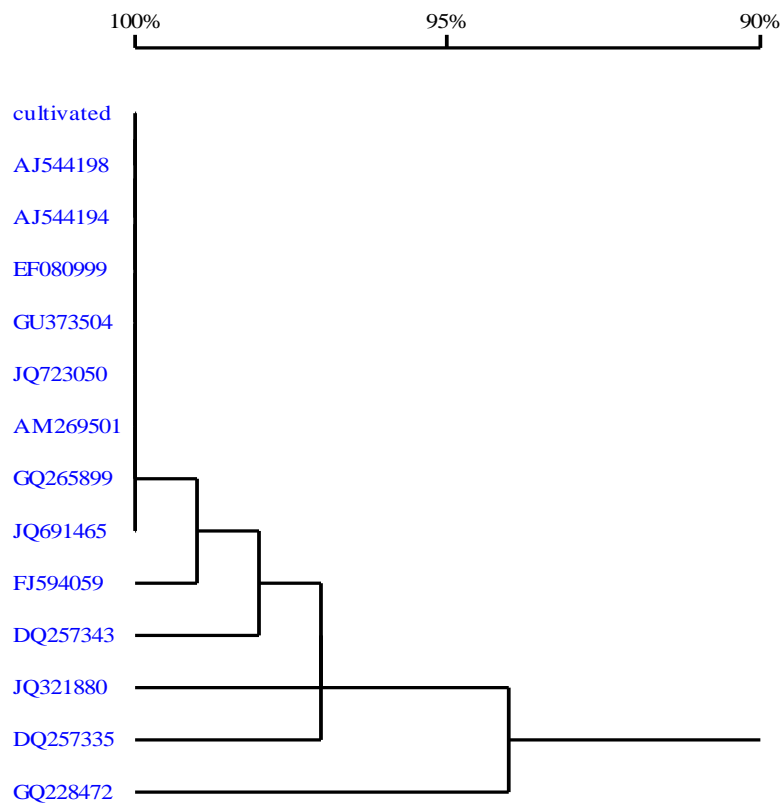


Figure 2. Cultivated strain of *Morchella conica* compared with other reported *Morchella* spp. for ITS1 sequences. AJ544198 (*Morchella conica*, Israel), AJ544194 (*Morchella conica*, Germany), EF080999 (*Moechalla conica*, China), GU373504 (*Morchella esculenta*, Finland), JQ723050 (*Morchella importuna*, China), AM269501 (*Morchella conica*, Poland), GQ265899 (*Morchella costata*, China), JQ691465 (*Morchella vulgaris*, Germany), FJ594059 (*Morchella conica*, Poland), DQ257343 (*Morchella conica*, China), JQ321880 (*Morchella populiphila*, China), DQ257335 (*Morchella angusticeps*, China), GQ228472 (*Morchella elata*, India).

and transport it to the cell via receptor-mediated way. As iron is an indispensable growth factor, microbes producing siderophores can capture it from the others. Iron-deprivation causes decreasing metabolic activity and stop cell growth. *Pseudomonas* species synthesise numerous iron-chelators, e.g. pyoverdine and pyochelin (Cox et al., 1981; Cox and Adams, 1985). These compounds play an important role in restricting the growth of fungi (Matthijs et al., 2007). In this study, the numbers of culturable microbial population in soil at different developing stages were measured and results (Table 1) showed that experimental groups were higher than the control in number of bacteria, fungi and actinomycetes ($P < 0.05$).

In detail, groups treated with ferric citrate and Fe-EDDHA during spore-formation stage show no significant differences as compared to the control group in bacteria number, but the number of fungi significantly increased and actinomycetes' number were significantly decreased. During

primordium differentiation stage, experimental groups except the groups treated with $\text{Fe}_2(\text{SO}_4)_3 \cdot 9\text{H}_2\text{O}$, were all significantly increased in the bacteria, fungi and actinomycetes' numbers, but the group treated with ammonium ferric sulfate ($\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$) during spore formation were higher than others (included the control group) in the microorganism number, and such tendency was found in soil microflora number during the primordium differentiation stage. It is interesting that numbers of soil microorganism showed a tendency of increasing from spore formation stage to primordium differentiation stage and reached the highest, and then decreasing the microflora numbers when the *M. conica* developed into the fruit-body growth stage, but numbers of fungi were relatively changed lower than others. Generally, the bacterial and fungi population increased significantly from spore-formation stage to the primordia formation stage and increased up, then decreased at fruit-body maturing stage while the actinomycetes did not show the same

order. Group with ammonium ferric sulfate [$\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$] often had the highest number compared to others at different stages. So, these results suggested that numbers of microorganisms of the grown morel soils differ in different ferric fertilizers. From these results, we also think that it is necessary to use the denaturing gradient gel electrophoresis (DGGE) technique with environmental samples to analyze the genetic diversity of microflora populations, especially in DGGE study of fungal communities (Vainio and Hantula, 2000; Anderson et al., 2003).

Determination of enzymatic activity for *Morchella* grown soil

Soils were sampled from three stages which morel grows. At all sites there was a significant treatment effect for each enzyme tested ($P < 0.05$). The data suggested that four soil enzyme activities increased with the morel growing and reaches a maximum at primordia formation stage. *M. conica* grown soil with the ferric fertilizers' disposal at different stages had different effects on four soil enzyme (urease, polyphenol oxidase, invertase and protease) (Table 2). From the results, we found that four soil enzymatic activities increased with the morel growing and reaches a maximum at primordium differentiation stage. Among the four soil enzymes, polyphenol oxidase, invertase and protease showed a higher activity than urease. Group with ammonium ferric sulfate showed higher urease activity than those of other groups, probably due to the organic material or the greater nitrogen inputs and sub-sequent stimulation of microbial activity. Experimental groups showed higher activities in soil enzymes when compared to the control on a large scale. The data suggests that four soil enzyme activities increased with the morel growing and reaches a maximum at primordia formation stage. Among the four soil enzyme, polyphenol oxidase, invertase and protease showed a higher activity than urease.

During spore-formation stage, soil urease activities showed a tendency with $\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O} > \text{FeSO}_4 \cdot 7\text{H}_2\text{O} > \text{Fe}_2(\text{SO}_4)_3 \cdot 9\text{H}_2\text{O} > \text{Fe-EDDHA}$ and Ferric citrate > Nutrients > ck, and significant difference were shown between experimental groups except Nutrients and control group ($p < 0.05$), especially the group treated with ammonium ferric sulfate [$\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$] had been increased 8 times than control ($p < 0.01$). Soil polyphenol oxidase activities of the experimental groups, except the group treated with Fe-EDDHA, showed significant differences when compared with that of the control. Invertase activities of the experimental groups were changed a little but not significantly different when compared with the control, while that of groups treated with $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ and Nutrients were significantly decreased. Soil protease activities of the

groups treated with Nutrients, $\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$ and Fe-EDDHA were significantly increased when compared with that of the control. These results concludes that *M. conica* grown soil's enzymatic activities were effected by different ferric fertilizers, however, ammonium ferric sulfate ($\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$) and Fe-EDDHA relatively showed higher soil enzymatic activities than others.

Urease catalyzes the hydrolysis of urea into ammonia or ammonium ion depending on soil pH, and carbon dioxide. Urease is the most prominent involved in soil N cycling in the four enzymes (Tabatabai and Bremner, 1972; Cookson, 1999). Among the different enzymes in soil, dehydrogenase, urease and phosphatases are important in the transformation of plant nutrients (Gao et al., 2010). We know very well that morels, like other edible macro-fungi, their coarse proteins, nitrogen uptake levels were influenced by soil urease and protease. Such results perhaps indicated increased soil urease activities was necessary in order to promote the growth of morel hyphae, accumulate the yield of morel fruiting bodies and lower yield of commercial cultivation of *M. conica* which is as a result of the lower level of nitrogen application.

Results of the commercial planting for *M. conica*

In this study, the epigeal fructifications (ascocarps) of *M. conica* are usually appeared in late February or in early March when soil temperature warms to above 10°C and this is in accordance with previous reports (Kanwal and Reddy, 2012; Masaphy, 2010; Goldway et al., 2000; Pilz et al., 2007; Ower et al., 1986). A number of studies point out that obtaining sclerotia are a necessary part of the process for domestication of the genus and commercial production (Buscot, 1994). Also, we found obvious stage of sclerotial formation in our cultivation period of *M. conica*.

In this study, each group fruiting season length (d) was also used to characterize morel fruit body production. Most groups seemed to harvest in 18-22 days, which showed fruiting season length was not significantly correlated with any fertilizer, still positively affected by the air temperature and soil temperature (Mihaila et al., 2007). The yield results are shown in Figures 3 and 4.

In our experiment, most experimental groups, except the group with ferric sulfate [$\text{Fe}_2(\text{SO}_4)_3 \cdot 9\text{H}_2\text{O}$], increased the yields of *M. conica*; especially the groups with Fe-EDDHA and ammonium ferric sulfate [$\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$] resulted in higher yields of fruit-bodies than other experimental groups and higher than that of control group, 1.77 and 1.88 times of the control yields, respectively. We found that during primordial formation of morel, groups treated with ferric fertilizers appeared faster than the control, approximately three or four days earlier, with a highest soil microbial biomass and enzymatic activities. Maybe *M. conica* is responding to the different strategies

Table 1. Numbers of major soil microbial population groups from the *Morchella* grown soil.

Group	Bacteria (10 ⁶ cells/g dry soil)			Fungi (10 ⁴ cells/g dry soil)			Actinomycetes (10 ⁴ cells/g dry soil)		
	Sclerotia formation	Primordium differentiation	Fruit-body mature period	Sclerotia appearing	Primordium differentiation	Fruit-body mature period	Sclerotia appearing	Primordium differentiation	Fruit-body mature period
Citrate ferric	1.32±0.07 ^a	5.90±1.13 ^a	1.43±0.06 ^a	3.59±0.26 ^a	4.40±0.48 ^a	5.32±0.10 ^a	4.38±0.29 ^a	3.48±0.28 ^a	3.57±0.24 ^a
Fe-EDDHA	0.92±0.13 ^a	4.79±0.59 ^b	2.25±0.10 ^b	2.94±0.57 ^{ab}	5.34±0.15 ^b	1.14±0.06 ^b	1.53±0.85 ^b	2.78±0.12 ^b	4.60±0.07 ^b
Fe ₂ (SO ₄) ₃ ·9H ₂ O	2.63±0.34 ^b	3.26±0.21 ^c	3.94±0.06 ^c	2.50±0.73 ^b	4.56±0.21 ^a	1.64±0.13 ^c	3.75±0.06 ^c	6.30±0.23 ^c	3.76±0.10 ^a
Feso ₄ ·7H ₂ O	3.27±1.36 ^c	6.37±0.11 ^a	3.18±0.11 ^d	2.25±0.28 ^b	6.43±0.06 ^c	3.71±0.17 ^d	5.72±0.29 ^d	6.44±0.10 ^c	6.62±0.18 ^c
NH ₄ Fe(SO ₄) ₂ ·12H ₂ O	3.21±0.39 ^c	8.71±0.17 ^d	1.66±0.17 ^a	5.87±0.51 ^c	8.45±0.29 ^d	5.65±0.12 ^a	5.47±0.24 ^d	6.78±0.14 ^d	5.42±0.11 ^d
Nutrients	2.07±0.29 ^b	5.48±0.52 ^{ab}	1.48±0.10 ^a	6.54±0.12 ^c	6.68±0.13 ^c	1.36±0.35 ^{bc}	3.56±0.21 ^c	5.45±0.15 ^e	4.67±0.22 ^b
ck (pure water)	1.75±0.140 ^a	3.53±0.06 ^c	3.80±0.26 ^c	1.45±0.50 ^d	2.43±0.06 ^e	3.55±0.16 ^e	2.36±0.16 ^e	3.43±0.19 ^a	3.60±0.12 ^a

Mean values as M±SD. Within three mean values of each character, there is no significant differences between those contain same letters, and different letters in the same column denote significant differences among treatments according to ANOVA and LSD's multiple comparisons ($p < 0.05$).

Table 2. Soil enzymatic activities of the *Morchella* grown soil which treated by different ferric fertilizers.

Group	Sclerotia formation				Primordium differentiation stage				Fruit-body mature period			
	Urease	Polyphenol oxidase	Invertase	Protease	Urease	Polyphenol oxidase	Invertase	Protease	Urease	Polyphenol oxidase	Invertase	Protease
Citrate ferric	0.84±0.13a	3.55±0.55a	2.48±0.22a	3.34±0.10a	1.43±0.06a	6.52±0.59a	3.35±0.15a	5.44±0.10a	1.54±0.21a	6.35±0.02a	3.33±0.10a	4.53±0.15b
EDDHA.Fe	0.84±0.03a	2.36±0.36b	2.77±0.20c	4.53±0.03b	1.25±0.10b	4.20±0.62b	5.55±0.10b	6.47±0.61b	2.46±0.64b	3.55±0.15c	4.76±0.25b	3.33±0.01a
Fe ₂ (SO ₄) ₃ ·9H ₂ O	1.09±0.36c	4.32±0.10c	2.16±0.53e	3.57±0.32d	1.55±0.15a	6.47±0.75a	6.63±0.06c	5.44±0.06a	1.55±0.15d	5.42±0.15d	3.35±0.26a	3.49±0.50d
Feso ₄ ·7H ₂ O	1.23±0.20d	6.54±0.10d	1.36±0.61f	2.72±0.15e	3.14±0.20c	7.46±0.71c	3.34±0.78a	6.54±0.06d	3.41±0.06b	5.38±0.21d	2.76±0.44d	5.66±0.20e
NH ₄ Fe(SO ₄) ₂ ·12H ₂ O	4.28±0.04e	5.35±0.55e	2.92±0.01bc	4.22±0.10f	7.56±0.06d	7.33±0.10c	3.02±0.19d	2.56±0.10e	5.75±0.81c	6.36±0.46a	2.62±0.05e	3.32±0.06a
Nutrients	0.76±0.36b	3.51±0.35a	1.07±0.10d	6.36±0.49c	1.16±0.32b	4.19±0.21b	5.55±0.13b	7.48±0.53c	1.55±0.46a	3.44±0.10b	4.44±0.17c	2.44±0.15c
Ck (pure water)	0.53±0.11a	2.49±0.47b	2.74±0.08b	2.55±0.15g	3.20±0.26c	3.36±0.06d	2.70±0.08e	3.12±0.12f	2.45±0.25b	3.43±0.10b	2.67±0.21f	3.34±0.53a

Urease, [NH₃-N/ (mg g⁻¹24⁻¹)]; polyphenol oxidase, [gallic acid/(mg g⁻¹24⁻¹)]; invertase, [glucose/(mg g⁻¹24⁻¹)]; protease [amino acid/(mg g⁻¹24⁻¹)].

for growth and sclerotia formation. The result of better morel growth might be with the increasing soil microbial population due to fertilizer application, higher release of root exudates and larger amounts of root residue left in the soil, which positively influence microbial processes and development. This indicates that different ferric fertilizers could influence the primordial formation of

morel and affect the yield of fruiting bodies by its soil microbial population and enzymatic activities. In nature, morel fruiting body formation is associated with a broad range of environmental stress conditions, some of which have yet to be defined (Masaphy, 2010) and up to now, there is no report about morel fruiting body formation and yield is associated with microelements, such as

ferric, zinc, copper, molybdenum, among others. However, many literatures reported that microelements influenced the sclerotial formation and morels are rich in trace elements in their fruit-body (Kanwal and Reddy, 2012; Kalač, 2010; Amir et al., 1995). Fe-EDDHA is usually used to correct iron chlorosis in Strategy I plant, because of its great stability and solubility at high pHs (Rojas et al., 2008)



Figure 3. Artificial cultivation of morels. Figure 3A, milky sclerotia produced by *M. conica*, after incubated to puffed wheat soil media; Figure 3B, seed-planting. *Morchella* growing situation after hybridization, the plastic packages were equipped with the sterilized mixture including wheat grain, sawdust, chaff and vermiculite. They were used until the mycelium grown well; Figure 3C, morel growing period, is relative to certain temperature and humidity. Fruiting season length is not significantly correlated with any fertilizer; Figure 3D, fruit bodies in harvest time. The mature fruiting body reached 8–16 cm in length. The phenotypic features-fruiting body shape, color and size, are vary with, and affected by environmental conditions and developmental dynamics.

2008). In our study, it shows a higher effect on morel fruiting body than other ferric fertilizers did; the possible reasons may come from its great solubility. Nevertheless, the functional mechanisms of effects on different ferric fertilizers or other nutrition elements need to be further researched. In addition, we think the same microelement or mineral element may influence the nutrient uptake for edible fungi by different approaches, it is difficult to give a sole conclusion whether a nutrient element promotes or hinders the experimental fungi to growth or even influence its yield at last. The best bet to optimize the nutrition plan for planting edible fungi especially *Morchella* spp. is to select a lot of nutrients (including trace elements) and to analyses their actual efficacy.

Conclusion

This paper reports studies on the effect of ferric fertilization on microbial biomass, enzymatic activities and morel yield. Sequencing of ITS showed that the species is

closely related to *M. conica*. The results showed that among the six ferric fertilizers treatments, Fe-EDDHA, the ammonium ferric sulfate $[\text{NH}_4\text{Fe}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}]$ results in the highest amount of microbial biomass, enzymatic activities and morel yields, while the treatments of CK gave the opposite results.

China is rich in nearly 30 morels species with eleven newly discover nine species within the Esculenta clade (yellow morels) and two novel species within the Elata clade (black morels), Over the 30 species, 20 appear to be endemic. The annual export of dried morels has been increased fivefold over the past 5 years to 900,000 kg, averaging \$160 US dollars per kilogram (Du et al., 2012a). With their ever-increasing popularity, it is possible to purchase dried morels in local supermarkets in numerous countries throughout the year. Up to the present day, we have well known *M. conica* of its stable and effective planting condition. The study indicates ferric fertilizers making a big difference on the primordial formation of morel, further attempts regarding optimizing nutrition plan need to be carried out.

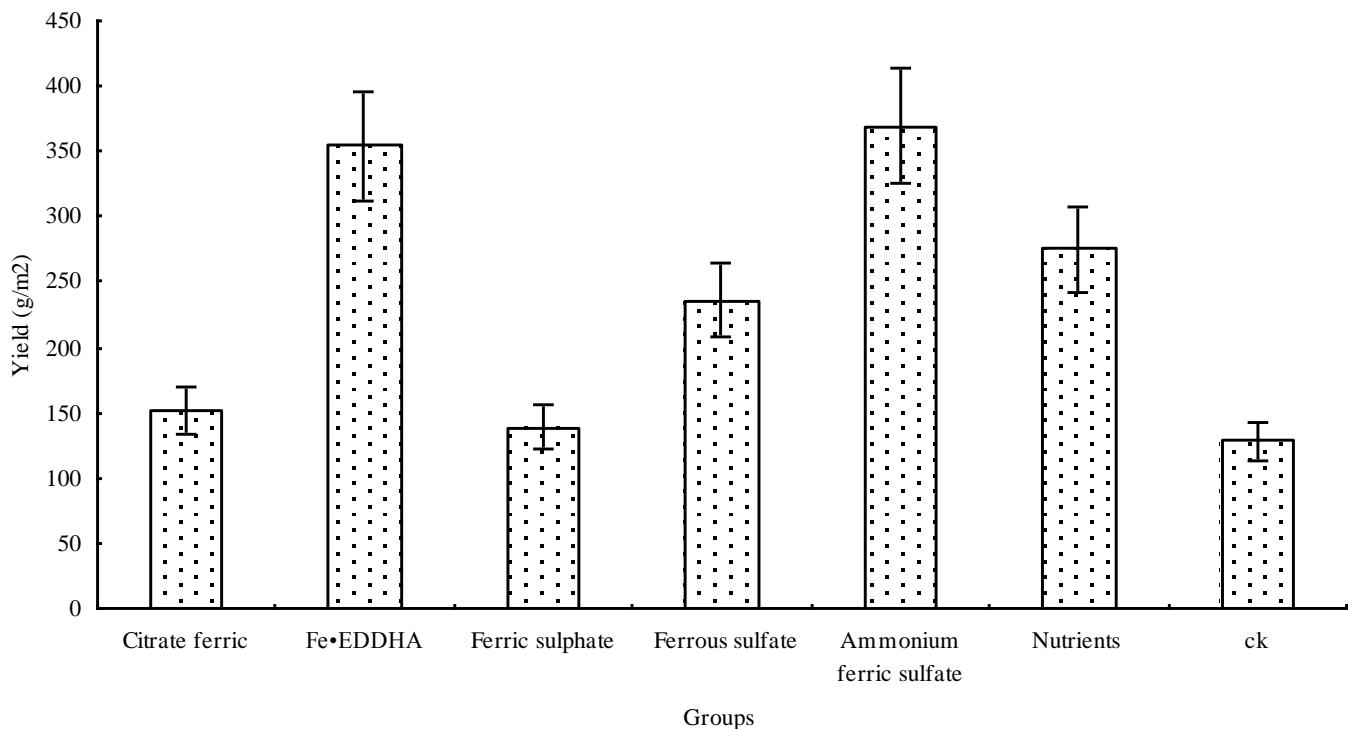


Figure 4. Yields of *Morchella conica* treated by different ferric fertilizer.

ACKNOWLEDGEMENTS

This work was partially supported by Sichuan Tongfeng Science and Technology Co. Ltd (No. 2010-01) and the Chengdu Municipal Key Research (10GGYB314NC-182).

REFERENCES

- Amir R, Levanon D, Hadar Y, Chet I (1995). Factors affecting translocation and sclerotia formation in *Morchella esculenta*. *Exp. Mycol.* 19:61-70.
- Anderson IC, Campbell CD, Prosser JI (2003). Diversity of fungi in organic soils under a moorland-Scots pine (*Pinus sylvestris* L.) gradient. *Environ Microbiol.* 5(11):1121-1132.
- Arkan O (1992). The heterokaryon formation and hyphal fusions at monoascospore cultures of *Morchella conica* Pers. *Doğa Tr. J. Biol.* 16:203-216.
- Bunyard BA, Nicholson MS, Royse DJ (1994). A systematic assessment of *Morchella* using RFLP analysis of the 28S ribosomal RNA gene. *Mycologia* 86(6):762-772.
- Bunyard BA, Nicholson MS, Royse DJ (1995). Phylogenetic resolution of *Morchella*, *Verpa*, and *Disciotia* [Pezizales: Morchellaceae] based on restriction enzyme analysis of the 28S ribosomal RNA gene. *Mycol. Res.* 19(3):223-233.
- Buscot F (1993). Mycelial differentiation of *Morchella esculenta* in pure culture. *Mycol. Res.* 97(2):136-140.
- Buscot F (1994). Ectomycorrhizal types and endobacteria associated with ectomycorrhizas of *Morchella elata* (Fr.) Boudier with *Picea abies* (L.) Karst. *Mycorrhiza* 4(5):223-232.
- Buscot F, Wipf D, Di Battista C, Munch JC, Botton B, Martin F (1996). DNA polymorphisms in morels I:PCR/RFLP analysis of the ribosomal DNA spacers and microsatellite-primed PCR. *Mycol. Res.* 100(1):63-71.
- Claudia Stihl, Anca Gheboianu, Cristiana Radulescu, Ion V. Popescu, Gabriela Busuioc, and Lulian B (2010). Studies Concerning the Accumulation of Minerals and Heavy Metals in Fruiting Bodies of Wild Mushrooms. *AIP Conference Proceedings*, 1387:282-287.
- Cookson P (1999). Special variation in soil urease activity around irrigated date palms. *Arid Soil Res. Rehabil.* 13:155-169.
- Cox CD, Adams P (1985). Siderophore activity of pyoverdine for *Pseudomonas aeruginosa*. *Infect. Immun.* 48(1):130-138.
- Cox CD, Rinehart KL, Moore ML, Cook JC (1981). Pyochelin: novel structure of an iron-chelating growth promoter from *Pseudomonas aeruginosa*. *Proc. Natl. Acad. Sci. USA* 78(7):302-308.
- David Pilz, Rebecca M, Susan A, Luis V, Shannon B, Tricia LW, Catherine GP, Erika M, Blaze B, Randy M, Jane ES (2007). Ecology and Management of Morels Harvested From the Forests of Western North America. General Technical Report. PNW-GTR-710, 2007(3).
- Donoghue MJ (2008). A phylogenetic perspective on the distribution of plant diversity. *Proc. Natl. Acad. Sci. U.S.A.* 105:11549-11555.
- Du XH, Zhao Q, O'Donnell K, Rooney AP, Yang ZL (2012a). Multigene molecular phylogenetics reveals true morels (*Morchella*) are especially species-rich in China. *Fungal Genet. Biol.* 49:455-469.
- Du XH, Zhao Q, Yang ZL, Hansen K, Taşkın H, Büyükalaca HS, Dewsbury D, Moncalvo J-M, Douhan GW, Robert VARG, Crous PW, Rehner SA, Rooney AP, Sink S, O'Donnell K (2012b). How well do ITS rDNA sequences differentiate species of true morels (*Morchella*)? *Mycologia* 104:1351-1368.
- Gao Y, Zhou P, Mao L, Zhi Y, Zhang C, Shi W (2010). Effects of plant species coexistence on soil enzyme activities and soil microbial community structure under Cd and Pb combined pollution. *J. Environ. Sci.* 22(7):1040-1048.
- Gessner RV, Romano MA, Schultz RW (1987). Allelic variation and segregation in *Morchella deliciosa* and *M. esculenta*. *Mycologia* 79(3):683-687.
- Goldway AR, Goldberg D, Hadar Y, Levanon D (2000). *Morchella conica* exhibiting a long fruit season. *Mycol. Res.* 104(8):1000-1004.
- Guler P, Ozkaya EG (2009). Morphological development of *Morchella*

- conica* mycelium on different agar media. J. Environ. Biol. 30(4):601-604.
- Guler P, Ozkaya EG (2008). Sclerotia structure of *Morchella conica* in agar media with different carbohydrates. Acta Alimentaria 37 (3):347-357.
- Gursoy N, Sarikurkcu C, Cengiz M, Solak MH (2009). Antioxidant activities, metal contents, total phenolics and flavonoids of seven *Morchella* species. Food Chem. Toxicol. 47(9):2381-2388.
- Gussov D, Clackson T (1989). Direct clone characterization from plaques and colonies by polymerase chain reaction. Nucleic Acids Res. 17(10):4000.
- Halliwell B (1991). Reactive oxygen species in living systems: source, biochemistry, and role in human disease. Am. J. Med. 91:1422.
- Hawksworth DL (2004). Fungal diversity and its implications for genetic resource collections. Stud. Mycol. 50:9-18.
- Higgins DG, Sharp PM (1988). CLUSTAL: a package for performing multiple sequence alignment on a microcomputer. Gene 73(1):237-244.
- Jensen HI (1951). Notes on the biology of *Azotobacter*. J. Appl. Microbiol. 14(1):89-94.
- Kalač P (2010). Trace element contents in European species of wild growing edible mushrooms: a review for the period 2000–2009. Food Chem. 122(1):2-15.
- Kanwal HK, Acharya K, Ramesh G, Reddy MS (2011). Molecular characterization of *Morchella* species from the Western Himalayan region of India. Curr. Microbiol. 62(4):1245-1252.
- Kanwal HK, Reddy MS (2012). The effect of carbon and nitrogen sources on the formation of sclerotia in *Morchella* spp. Ann. Microbiol. 62(1):165-168.
- Kar M, Mishra D (1976). Catalase, peroxidase, and polyphenoloxidase activities during rice leaf senescence. Plant Physiol. 57(2):315-319.
- Kellner H, Luis P, Buscot F (2007). Diversity of laccase-like multicopper oxidase genes in *Morchellaceae*: identification of genes potentially involved in extracellular activities related to plant litter decay. FEMS Microbiol. Ecol. 61(1):153-163.
- Kellner H, Renker C, Buscot F (2005). Species diversity within the *Morchella esculenta* group (Ascomycota: *Morchellaceae*) in Germany and France. Organ Divers. Evol. 5:101-107.
- Ladd JN, Butler JHA (1972). Short-term assays of soil proteolytic enzyme activities using proteins and dipeptide derivatives as substrates. Soil Biol. Biochem. 4(1):19-30.
- Martin JP (1950). Use of acid, rose bengal and streptomycin in the plate method for estimating soil fungi. Soil Sci. 69(3):215-232.
- Masaphy S (2005). External ultrastructure of fruit body initiation in *Morchella*. Mycol. Res. 109(4):508-512.
- Masaphy S (2010). Biotechnology of morel mushrooms: successful fruiting body formation and development in a soilless system. Biotechnol. Lett. 32(10):1523-1527.
- Mendil D, ULUÖZLÜ, ÖD, TÜZEN M, HASDEMİR E, SARI H (2005). Trace metal level in mushroom samples from Ordu, Turkey. Food Chemistry. 91:463-467.
- Mihail J, Hauser M, Cushman F, Liane Young R, Kang-Xing J (2007). A dissociation between moral judgments and Justifications. Mind Lang. 22:1-21
- Nair SK, Subba-Rao NS (1977). Microbiology of the root region of coconut and cacao under mixed cropping. Plant Soil 46(3):511-519.
- Nevcihan G, Cengiz S, Mustafa C, Halil SM (2009). Antioxidant activities, metal contents, total phenolics and flavonoids of seven *Morchella* species. Food Chem. Toxicol. 47:2381-2388.
- O'Donnell K, Rooney AP, Mills GL, Kuo M, Weber NS, Rehner SA (2011). Phylogeny and historical biogeography of true morels (*Morchella*) reveals an early Cretaceous origin and high continental endemism and provincialism in the Holarctic. Fungal Genet. Biol. 48:252–265.
- Ower R (1982). Notes on the development of the morel ascocarp. Mycologia 74(1):142-144.
- Ower R, Milles G, Malachowski J (1986). Cultivation of *Morchella*. US Patent No. 4594809.
- Papa S, Bartoli G, Pellegrino A, Fioretto A (2010). Microbial activities and trace element contents in an urban soil. Environ. Monit. Assess. 165(1-4):193-203.
- Pilz D, McLain R, Alexander S, Villarreal-Ruiz L, Berch S, Wurtz TL, Parks CG, McFarlane E, Baker B, Molina R, Smith JE (2007). Ecology and Management of Morels Harvested from the Forests of Western North America. General Technical Report PNW-GTR-710. Portland:US Department of Agriculture, Forest Service, Pacific Northwest Research Station. p. 161.
- Robert V, Szoke S, Jabas J, Vu D, Chouchen O, Blom E and Cardinali G (2011). BioloMICS software, Biological data Management, Identification, Classification and Statistics. Open Appl. Info. J. 5:87-98.
- Rojas CL, Romera FJ, Alcántara E, Pérez-Vicente R, Sarriego C, Garcaí-Alonso JI, Boned J, Marti G (2008). Efficacy of Fe(o,o-EDDHA) and Fe(o,p-EDDHA) Isomers in Supplying Fe to Strategy I Plants Differs in Nutrient Solution and Calcareous Soil. J. Agric. Food Chem. 56(22):10774-10778.
- Royse DJ, May B (1990). Interspecific allozyme variation among *Morchella* spp. and its inferences for systematics within the genus. Biochem. Syst. Ecol. 18(7-8):475-479.
- Sathe GM, O'Brien S, McLaughlin MM, Watson F, Livi GP (1991). Use of polymerase chain reaction for rapid detection of gene insertions in whole yeast cells. Nucleic Acids Res. 19(17):4775.
- Tabatabai MA, Bremner JM (1972). Assay of urease activity in soils. Soil Biol. Biochem. 4(4):479-487.
- Taşkin H, Büyükalaca S, Doğan HH, Rehner SA, O'Donnell K (2010). A multigene molecular phylogenetic assessment of true morels (*Morchella*) in Turkey. Fungal Genet. Biol. 47(8):672-682.
- Vainio EJ, Hantula J (2000). Direct analysis of wood-inhabiting fungi using denaturing gradient gel electrophoresis of amplified ribosomal DNA. Mycol. Res. 104(8):927-936.
- Varma A, Chincholkar S (2007). Microbial siderophores. Springer, Berlin.
- Vetter J (2004). Arsenic content of some edible mushroom species. Eur. Food Res. Technol. 219:71-74.
- Volk TJ, Leonard TJ (1989). Experimental studies on the morel. I. Heterokaryon formation between monoascospore strains of *Morchella*. Mycologia 81(4):523-531.
- Volk TJ, Leonard TJ (1990). Cytology of the life-cycle of *Morchella*. Mycol. Res. 94(3):399-406.
- Wipf D, Bedell JP, Munch JC, Botton B, Buscot F (1996). Polymorphisms in morels: isozyme electrophoretic analysis. Can. J. Microbiol. 42(8):819-82.
- Wipf D, Fribourg A, Munch JC, Botton B, Buscot F (1999). Diversity of the internal transcribed spacer of rDNA in morels. Can. J. Microbiol. 45(9):769-778.
- Wipf D, Munch J C, Botton B, Buscot F (1996). DNA polymorphism in morels: complete sequences of the Internal Transcribed Spacer of genes coding for rDNA in *Morchella esculenta* (Yellow Morel) and *Morchella conwa* (Black Morel). Appl. Environ. Microbiol. 62:3541-3543.
- Wurtz Tricia L, Wiita Amy L, Weber Nancy S, Pilz David (2005). Harvesting morels after wildfire in Alaska. Research Note RN-PNW-546. Portland, OR:U.S. Forest Service Pacific Northwest Research Station.
- Yamaguchi T, Takamura H, Matoba T, Terao J (1998). HPLC method for evaluation of the free radical-scavenging activity of foods by using 1,1-dicylhydrazyl. Biosci. Biotechnol. Biochem. 62:1201-1204.
- Ye WQ, Zhang WQ, Zhou PX, Xie JZ, Zhang YL (1983). Experimental methods of soil microbes. Science Press (in Chinese).
- Zhao Q, Xu ZZ, Cheng YH, Qi SW, Hou ZJ (2009). Artificial cultivation of *Morchella conica*. Southwest China J Agric. Sci 22:1690–1693 (in Chinese).
- Zhu Fangkun, Qu Li, Fan Wenxiu (2011). Assessment of heavy metals in some wild edible mushrooms collected from Yunnan Province, China. Environ. Monitor. Assess. 179(1-4):191-199.

Full Length Research Paper

Analysis of an outbreak of *Klebsiella pneumoniae* by the DiversiLab system and pulsed-field gel electrophoresis

Jing Li, Zhi Dong Hu* and Cui E. He

Department of Clinical Laboratory, Tian jin Medical University General Hospital, AnShan Road, HePing District, Tianjin, China.

Accepted 26 July, 2013

The aim of this study was to analyse the reason that five patients associated with bloodstream infections by *K. pneumoniae* in a surgical ward using Pulse Field Gel Electrophoresis (PFGE) and the repetitive-sequence-based PCR (rep-PCR) employing the DiversiLab system. In July 2010, eight isolates were collected from the same surgical ward of a hospital, and the specimens from sputum, blood and abdominal drainage fluid, respectively. The PFGE patterns after XbaI digestion and rep-PCR profiles produced by the DiversiLab system were determined for eight isolates. The Rep-PCR profiles produced by using the DiversiLab system showed that the eight isolates can be divided into two groups; the K8-02 as a group and the other seven strains is the other group. The eight strains of *K. pneumoniae* can be divided into two groups (A and B), seven strains were type A which the subtype A1 is the main-type (K8-01, K8-03, K8-04, K8-06, K8-08), and one is type B (K8-02). The PFGE and rep-PCR interpretations were concordant for the eight strains of *K. pneumoniae*. This data suggest that the DiversiLab system may be a reasonable alternative to PFGE for investigation and control of nosocomial infection outbreaks caused by *K. pneumoniae*, since it is easy to use, rapid and does not require highly skilled operators (Mazzariol et al., 2012).

Key words: *Klebsiella pneumoniae*, outbreak, pulse field gel electrophoresis (PFGE), rep-PCR, DiversiLab system.

INTRODUCTION

Klebsiella pneumoniae strain is one of the most important gram-negative pathogens causing hospital-acquired infection and community-acquired infection (Chen et al., 2013). In recent years, the infection and drug resistance of *K. pneumoniae* strain have been significantly increased; especially the growing in number of ESBLs-producing strains has become a major cause of death in patients with pneumonia. Therefore, rapid and accurate pathogen

traceable epidemiological method is extremely important for the blocking and control of hospital infection epidemic spread. Now the major genotyping method for *K. pneumoniae* strain is PFGE, which is recognized as the gold standard of bacteria typing method with the highest discriminatory power (Han et al., 2013). However, PFGE requires specialized equipment and is technically demanding, labor intensive, and relatively slow, as it may take 2

*Corresponding author. E-mail: huzhidong27@163.com. Tel: 86-22-6081-4202. Fax: 86-22-6081-4186.

Abbreviations: ESBLs, Extended spectrum beta-lactamase; CLSI, clinical and laboratory standards institute; PFGE, pulse field gel electrophoresis; rep-PCR, repetitive-sequence-based PCR; *K. pneumoniae*, *Klebsiella pneumoniae*.

Table 1. The clinical data of 8 *K. pneumoniae* strains.

Strain	Ward code	Patient code	Specimens type	Collection time
K8-01	S1	W	Sputum	July 17, 2010
K8-02	S1	W	Abdominal cavity drainage liquid	July 17, 2010
K8-03	S1	W	Blood	July 29, 2010
K8-04	S1	W	Blood	July 18, 2010
K8-05	S1	L	Blood	July 20, 2010
K8-06	S1	H	Blood	July 21, 2010
K8-07	S1	Y	Blood	July 29, 2010
K8-08	S1	J	Catheter	July 29, 2010

to 5 days to obtain results, depending on the organism and the methods utilized (Ligozzi et al., 2010). DiversiLab system is a typing method based the principle of rep-PCR, with the use of standardized operating, the high-resolution microfluidic chip and fluorescence detection system and an unified data processing software, has good repeatability, and can be used in the laboratory long-term epidemiological typing methods. It has been applied in *Acinetobacter baumannii* (Fontana et al., 2008), *Staphylococcus aureus* (Ross et al., 2005), *Pseudomonas aeruginosa* (Doléans-Jordheimetal., 2009), *Mycobacterium* (Cangelosi et al., 2004), and other molecular epidemiological studies. At present, reports on the DiversiLab system are rarely seen in China. In this study, we use the DiversiLab system to analyse genotypes of 8 *K. pneumoniae* strains, and compare the consistency between the DiversiLab system and Pulse Field Gel Electrophoresis (PFGE).

MATERIALS AND METHODS

Bacterial

The eight *K. pneumoniae* strains were collected from one surgical ward of a hospital in July 2010. All the strains were identified by using VITEK-2 compact automatic system (bioMérieux, Marcy l'Etoile, France). The clinical datas are shown in Table 1.

Antibiotic susceptibility tests

Antibiotic susceptibility test profiles for 11 antimicrobial agents including amoxicillin/clavulanic acid (AMC), aztreonam (ATM), ciprofloxacin (CIP), ceftriaxone (CRO), cefazolin (CFZ), ampicillin (AMP), meropenem (MEM), cefoperazone/sulbactam (CPZ/SU), cotrimoxazole (SXT), piperacillin/tazobactam (PIP/TA), gentamicin (GEN) were determined by using a broth microdilution method. Standards for antimicrobial susceptibility testing and interpretation were based on the CLSI 2011 guidelines (CLSI, 2011). *Escherichia coli* ATCC 25922 and *Pseudomonas aeruginosa* ATCC 27853 were used as reference strains for susceptibility testing.

Pulse field gel electrophoresis method

Genomic DNA was prepared as described previously (Gori et al., 1996; Pang et al., 2002), but with some modifications. *K. pneumoniae* isolates were grown overnight on Mueller-Hinton plate. Takings a

few pure colonies with a sterile cotton swab and dissolving with 2.5 ml TE buffer, then making 4.0 McFarland units, and incubating at 37°C. 150 µl prepared specimens were removed, 20 µl proteinase K and 30 µl 10% SDS (final concentration is 1%) were added, and mixed to warm-up at 56°C. Taking 2% low melting point gel and the prepared bacteria to fill the mold with the proportion of 1:1, then it was solidified at room temperature for 30 min. Then adding the molding of the plastic block to 1 ml lysis buffer which contains 5 µl proteinase K, and incubating for 2 h at 54°C shaking-bed. Then washed 2 times with 50°C preheated sterile distilled water, 10 min for each time, and washed 3 times with 50°C preheated TE buffer, 10 min for each time. Remove sample plastic block and cut down about 2 mm gel, then immersed it in digestion system which contains 200 µl restriction endonuclease enzyme Xba I, then 37°C digested overnight. Attach the digested gel block to the end of the glue comb, then place the comb at horizontally inverted plastic mold. The melted 1% electrophoresis agarose which had balanced at 50°C water to the mold was added, and then solidified at room temperature for 30 min. Setting the electrophoresis conditions as follows: the electrophoresis buffer 0.5 × TBE, switching time of 4 ~ 30 s, the field strength is 6 V/cm, temperature at 14°C, the electric field angle of 120°, and the electrophoresis time of 18.5 h, then view the result with ethidium bromide staining under ultraviolet light observations, pulsed-field gel electrophoresis (Tenover et al., 1995).

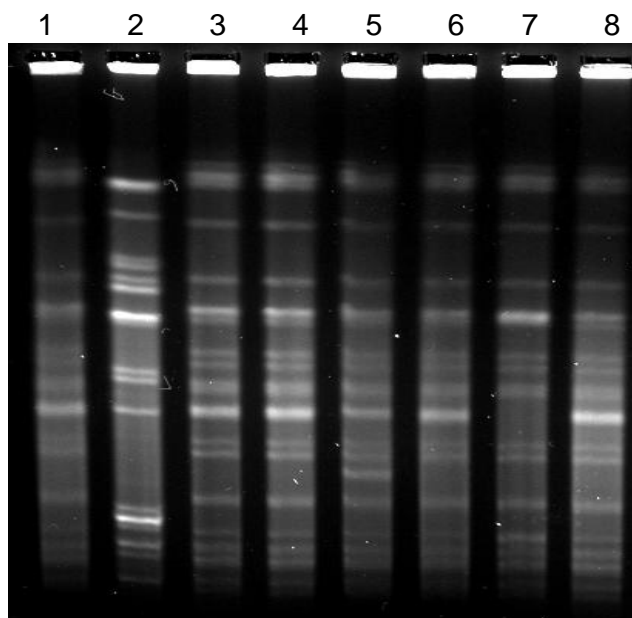
To determine the relationship between the band patterns as the following principles by visual inspection: 1) the same strains: the restriction map with same size and number is considered to be the one strain (main stream type); 2) closely related types: the electrophoretic bands have 3 or less different bands with the main phenotype isolates due to mutation, insertion, deletion or inversion are considered to be the subtype of the main type; 3) may related types: type with 4 to 6 bands different is considered to be of different type; 4) not relevant: has more than 7 different bands with the main type is considered no correlation in epidemiology.

Rep-PCR using DiversiLab system

Using MOBIO UltraClean (TM) microbial DNA extraction kit to extract bacterial DNA and adjusting the DNA concentration to 25 ~ 50 mg/L with visible spectrophotometer (NanoDrop ND-1000). Rep-PCR was performed by using the DiversiLab *K. pneumoniae* kit for DNA fingerprinting (bioMérieux, Marcy l'Etoile, France) according to the manufacturer's instructions. Preparing the 25 µl PCR amplification system: the 18 µl rep-PCRMM1, 2.5 µl GeneAmp10 × PCR buffer, 2.0 µl Primer Mix, 0.5 µl AmpliTaq DNA polymerase and 2 µl template. Thermal cycling parameters were as follows: initial denaturation of 94°C for 30 s, annealing at 50°C for 30 s, extension at 70°C for 90 s, as a cycle, a total of 35 cycles, and a final extension at 70°C for 3 min. Analysis of PCR products were implemented by using DiversiLab System (bioMérieux) in which the amplified fragment of various size and fluorescent intensities were separated

Table 2. MIC values ($\mu\text{g/ml}$) of 8 *K. pneumoniae* strains.

Strain	(AMC)	(ATM)	(CIP)	(CRO)	(CFZ)	(AMP)	(MEM)	(CPZ/SU)	(SXT)	(PIP/TA)	(GEN)
K8-01	8	≤ 1	1	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16
K8-02	≥ 32	≤ 1	≥ 4	≤ 1	8	≥ 32	≤ 4	≤ 16	≤ 20	≥ 128	≤ 1
K8-03	4	≤ 1	0.5	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16
K8-04	4	≤ 1	0.5	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16
K8-05	8	≤ 1	0.5	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16
K8-06	4	≤ 1	0.5	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16
K8-07	4	≤ 1	0.5	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16
K8-08	4	≤ 1	1	≥ 64	≥ 64	≥ 32	≤ 4	≤ 16	≥ 320	≤ 4	≥ 16

**Figure 1.** The PFGE typing results of eight *K. pneumoniae* strains. Note: 1 = K8-01, 2 = K8-02, 3 = K8-03, 4 = K8-04, 5 = K8-05, 6 = K8-06, 7 = K8-07, 8 = K8-08.

and detected using a microfluidics chip with the Aligent 2100 Bioanalyzer (Aligent Technologies, Santa Clara, CA, USA). Further analysis was performed with the web-based DiversiLab software version 3.3 with the Pearson correlation coefficient to determine distance matrices and the un-weighted-pair group method with arithmetic mean to create dendrograms.

The resulting DNA fingerprinting patterns were viewed as electropherograms, and the report included a dendrogram constructed from a similarity matrix and a virtual gel image of the fingerprint for each DNA sample. The criteria references of virtual gel image analysis are divided into three kinds: similar is greater than 97%, and no differences between bands; indistinguishable is greater than 95%, and has 1 to 2 different bands; different is less than 95%, and more than 2 different bands (Casolari et al., 2005).

RESULTS

Antibiotics susceptibility test

In 8 *K. pneumoniae* strains, the KB-02 was different with

other isolates. The KB-02 was sensitive to SXT, GEN, MEM, CPZ/SU, CRO and ATM, and were resistant to AMC, CIP, CFZ, AMP and PIP/TA. The other 7 isolates were multiple drug-resistant *K. pneumoniae*, which were sensitive to AMC, ATM, CIP, MEM, CPZ/SU and PIP/TA, and were resistant to CRO, CFZ, AMP, SXT and GEN. The results were in Table 2.

PFGE and rep-PCR using DiversiLab

The PFGE patterns after digestion with XbaI, and Rep-PCR using the DiversiLab system were performed for eight isolates. The eight isolates can be divided into two groups (A and B), seven strains were type A (K8-01, K8-03, K8-04, K8-05, K8-06, K8-07, K8-08), and one is type B (K8-02). Results are shown in Figure 1. The DiversiLab system showed the same results (Figures 2 and 3). The PFGE and the DiversiLab system interpretations were concordant for *K. pneumoniae* isolates. The DiversiLab system allowed a complete microbial typing analysis in approximately 4 h compared to 3 days for PFGE in our study.

DISCUSSION

There are many bacteria genotyping methods, such as DiversiLab system, MLST, PFGE and ERIC. Among them, PFGE is recognized as the gold standard of bacteria typing method. Witt (Te et al., 2009) applied the DiversiLab system, MLST and PFGE to study 93 MRSA strains, and the Simpson coefficient among three methods were 0.860, 0.877 and 0.905, respectively. Although, the resolution is different, the three kinds of methods are commonly used in genotyping of MRSA. Qu et al. (2010) used the DiversiLab system for genotyping of *A. baumannii*, which got good results. The DiversiLab system is fast and simple, besides its commercial kits, which is available for laboratory epidemiological typing methods. The eight *K. pneumoniae* strains were isolated from the same surgical ward of a hospital during July 2010, and the specimens from sputum, blood and abdominal drainage fluid, respectively. Antibiotics susceptibility test showed that among the eight strains, seven strains

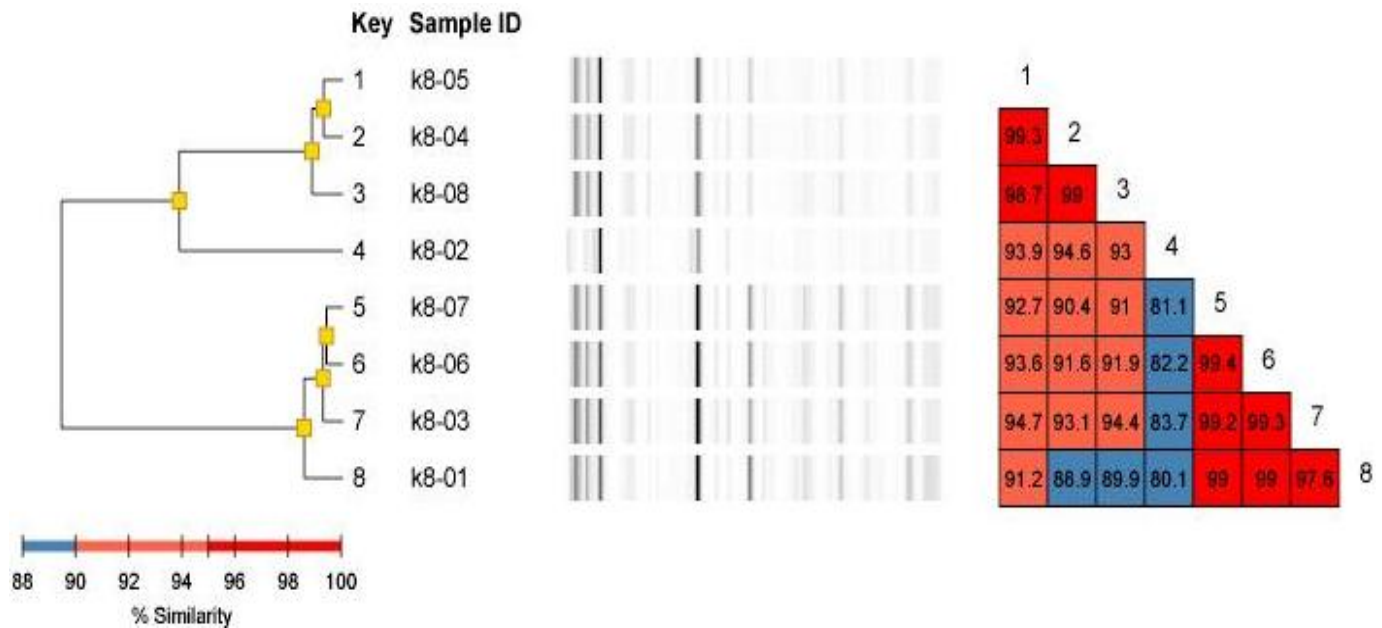


Figure 2. DiversiLab system typing results for *K. pneumoniae* strains; Note: The tree on the left side is the typing results of the DiversiLab system; figures in the right side of the matrix represent the similarity between the two strains, number on the right of the matrix represent strains, the Sample ID is the strain number.

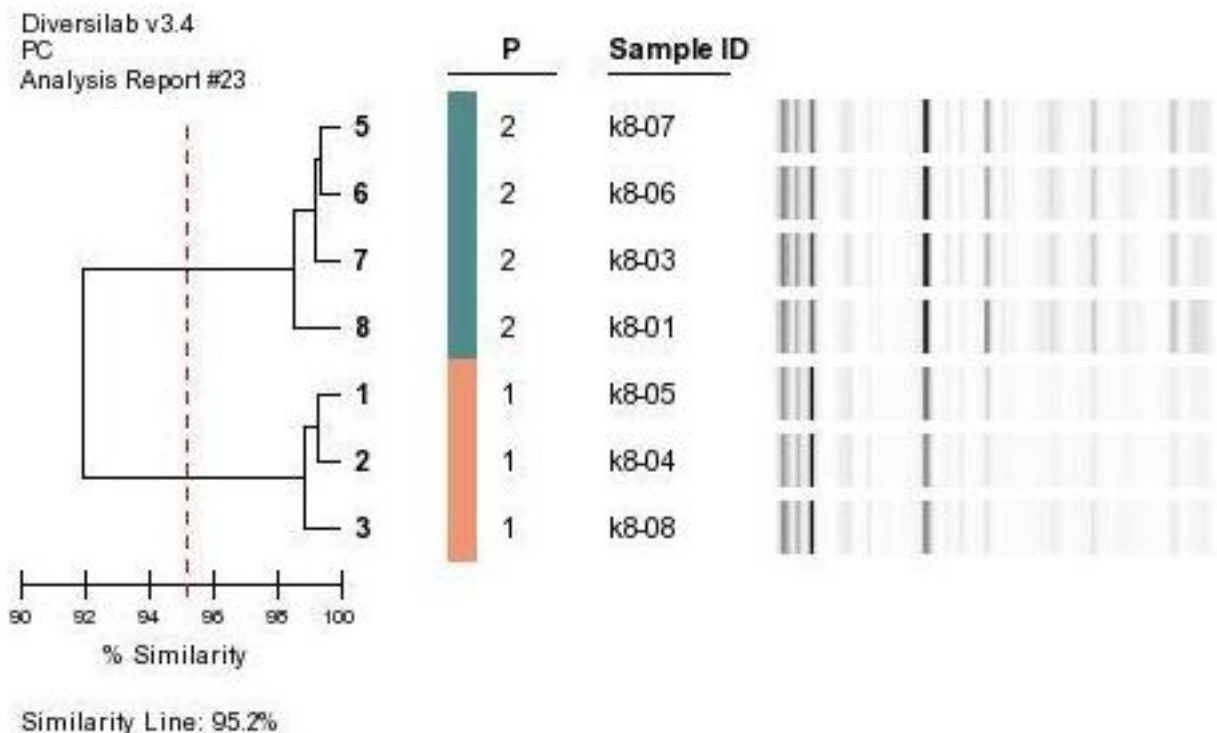


Figure 3. DiversiLab results of 7 *K. pneumoniae* strains except for K8-02 specimen.

have the same antimicrobial resistance pattern, except K8-2. The DiversiLab system and PFGE have proved these seven isolates belonging to the same type, so that

the infection is likely an outbreak of hospital. The susceptibility pattern of the isolate K8-02 from abdominal cavity drainage fluid was different from three other strains

that came from blood and sputum samples, which were all from the patients W. And PFGE patterns and the DiversiLab system showed no correlation among these four *K. pneumoniae* strains.

According to the time of the five patients infected, the DiversiLab system analysis found that W was the first infection patient, and subsequently infected with the L, H, Y and J. Because these four *K. pneumoniae* strains were the same type, in addition to, the PFGE map also showed the same result. Therefore, the patient W was most likely the source of this outbreak of nosocomial infection, and spreaded to the patients L, H, Y and J through exogenous way. At present, the most common typing method for *K. pneumoniae* is PFGE, which is the gold standard of bacterial genotyping methods and homology analysis (Wise et al., 2009). But the PFGE results usually can be read with naked eye, and different operators will influence the result. Furthermore, the technology is time-consuming, because it normally takes 3–5 days to get the final results. On the other hand, DiversiLab system that based REP-PCR is a standardized and automated systems, and the result collection and analysis process has no subjective restrictions. It can analyse 12 specimens in 40 min and the total process only need 6–8 h. The DiversiLab system has high resolution, good repeatability and simple operation, is a real-time quantitative detection. But the system also has shortcomings.

The analysis is race operation that needs only the 1 μ l each time, so that sometimes a bubble may need re-analysis of the amplified products and increases detection period and the cost. When using chip detection, even in the case of only several samples, the 12 sample holes and the standard hole must be added to the markers and gel fluorescent dye mixture, and result in reagent waste and cost. This study shows that the DiversiLab system is simple, fast, has high repeatability and high resolution, the results of quantitative processing, and various forms of output can be used as preferred genotyping method, especially on the short-term outbreak of a large number of specimens.

REFERENCES

- Cangelosi GA, Freeman RJ, Lewis KN, Livingston-Rosanoff D, Shah KS, Milan SJ, Goldberg SV (2004). Evaluation of a high-throughput repetitive-sequence-based PCR system for DNA fingerprinting of *Mycobacterium tuberculosis* and *Mycobacterium avium* complex strains. *J. Clin. Microbiol.* 42(6):2685-2693.
- Casolari C, Pecorari M, Fabio G, Cattani S, Venturelli C, Piccinini L, Tamassia MG, Gennari W, Sabbatini AM, Leporati G, Marchegiano P, Rumpianesi F, Ferrari F (2005). A simultaneous outbreak of *Serratia marcescens* and *Klebsiella pneumoniae* in a neonatal intensive care unit. *J. Hosp. Infect.* 61(4):312–320.
- Chen HB, Zhao CJ, Wang H, Cao B, Xu XL, Chu YZ, Hu ZD, Zhuo C, Hu BJ, Liu WE, Liao K, Zhang R, Zeng J, Wang Y, Luo YP, Wang ZW, Liu YM, Chen X, Tian B, Su DH, Zhou CM, Zou MX, Guo PH, Zhou HW, Jin Y (2013). An analysis of resistance of nosocomial infection pathogens isolated from 13 teaching hospitals in 2011. *Zhonghua Nei Ke Za Zhi.* 52(3):203-12.
- Clinical and Laboratory Standards Institute (CLSI) (2011). Performance standards for antimicrobial susceptibility testing: Twenty-first informational supplement, M100-S21. CLSI, Wayne, PA.
- Doléans-Jordheim A, Cournoyer B, Bergeron E, Croizé J, Salord H, André J, Mazoyer MA, Renaud FN, Freney J (2009). Reliability of *Pseudomonas aeruginosa* semi-automated rep-PCR genotyping in various epidemiological situations. *Eur. J. Clin. Microbiol. Infect. Dis.* 28(9):1105-1111.
- Fontana C, Favaro M, Minelli S, Bossa MC, Testore GP, Leonardis F, Natoli S, Favalli C (2008). *Acinetobacter baumannii* in intensive care unit: a novel system to study clonal relationship among the isolates. *BMC Infect. Dis.* 8(8):79.
- Gori A, Espinasse F, Deplano A, Nonhoff C, Nicolas MH, Struelens MJ (1996). Comparison of pulsed-field gel electrophoresis and randomly amplified DNA polymorphism analysis for typing extended-spectrum-beta-lactamase-producing *Klebsiella pneumoniae*. *J. Clin. Microbiol.* 34(10):2448-2453.
- Han H, Zhou H, Li H, Gao Y, Lu Z, Hu K, Xu B (2013). Optimization of Pulse-Field Gel Electrophoresis for Subtyping of *Klebsiella pneumoniae*. *Int. J. Environ. Res. Public Health* 10(7):2720-31.
- Ligozzi M, Fontana R, Aldegheri M, Scalet G, Lo Cascio G (2010). Comparative evaluation of an automated repetitive-sequence-based PCR instrument versus pulsed-field gel electrophoresis in the setting of a *Serratia marcescens* nosocomial infection outbreak. *J. Clin. Microbiol.* 48(5):1690–1695.
- Mazzariol A, Lo Cascio G, Ballarini P, Ligozzi M, Soldani F, Fontana R, Cornaglia G (2012). Rapid molecular technique analysis of a KPC-3-producing *Klebsiella pneumoniae* outbreak in an Italian surgery unit. *J. Chemother.* 24(2):93-6.
- Pang B, Jing H, Zheng H, Sun H, Zhao A, Xu J (2002). Molecular typing of Shiga-toxin producing *Escherichia coli* O157: H7 isolated in China with pulsed field gel electrophoresis. *Zhonghua Liu Xing Bing Xue Za Zhi.* 23 (2):123-126.
- QU Yan-yan, Wang Wen-fei, Zhou Hua, Fu Yi-qi, Yu Yun-song (2010). Investigation On the genotyping of *Acinetobacter baumannii* by DiversiLab system. *Clin. J. Lab. Med.* 33(5):425-429.
- Ross TL, Merz WG, Farkosh M, Carroll KC (2005). Comparison of an automated repetitive sequence-based PCR microbial typing system to pulsed-field gel electrophoresis for analysis of outbreaks of methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* 43(11):5642-5647.
- Te Witt R, Kanhai V, van Leeuwen WB (2009). Comparison of the DiversiLab system, Pulsed-field Gel Electrophoresis and Multi-Locus Sequence Typing for the characterization of epidemic reference MRSA strains. *J. Microbiol. Methods* 77(1):130-133.
- Tenover FC, Arbeit RD, Goering RV, Mickelsen PA, Murray BE, Persing DH, Swaminathan B (1995). Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J. Clin. Microbiol.* 33(9):2233-2239.
- Wise MG, Siragusa GR, Plumlee J, Healy M, Cray PJ, Seal BS (2009). Predicting *Salmonella enterica* serotypes by repetitive sequence-based PCR. *J. Microbiol. Methods* 76(1):18-24.

Full Length Research Paper

Towards efficient crude oil degradation by *Pseudomonas* sp. strain-O2: Application of Plackett-Burman design for evaluation of cultivation conditions

Mahmoud M. Berekaa

Environmental Sciences Department, Faculty of Science, Alexandria University, Maharam Bek, Alexandria, Egypt.

Accepted 2 September, 2013

Towards an efficient crude oil bioremediation, optimization study of crude oil degradation by *Pseudomonas* sp. strain-O2 was done. Preliminary experiments of crude oil degradation revealed that peptone was the optimal nitrogen source recording 73.3% removal of crude oil, in the presence of 0.5 g/L of yeast extract. The increase of phosphate ratio to 2.5 led to 80.1% removal of crude oil. To evaluate the significance of different culture conditions that affect crude oil biodegradation, Plackett-Burman factorial design was applied. Eleven variables were simultaneously examined. Among those variables, crude oil concentration was the highest positively significant variable that encourage crude oil degradation in *Pseudomonas* sp. strain-O2 affecting the degradation process, other factors namely, Na-succinate, $(\text{NH}_4)_2\text{SO}_4$, KH_2PO_4 and $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ showed moderate effect. While, yeast extract, inoculum concentration, agitation, K_2HPO_4 and NaCl were the lowest significant variables. Fractional factorial design experiments indicated that the pre-optimized medium showed approximately 1.5-folds increase in crude oil degradation by *Pseudomonas* sp. strain-O2.

Key words: Biodegradation, crude oil, *Pseudomonas* sp., experimental design, optimization.

INTRODUCTION

As a result of industrialization, many harmful substances including the spill of crude oil and various oil residues have been discharged into terrestrial and aquatic environments (Margesin, 2000). Dangerous accumulation of recalcitrant compounds in soil, sediments and groundwater are considered to be a potential health hazards (Korda et al., 1997). The clean-up of petroleum hydrocarbon-contaminated sites remains a priority task for restoration of the natural environment (ASTM, 1995). Considerable efforts are being spent to design cheap and feasible strategies for clean-up of contaminated sites.

Recently, bioremediation has proved to be a useful tool in removing oil (Boopathy, 2000; Ruiz et al., 2006). It attempts to accelerate the natural degradation rates by overcoming factors that limit microbial degradation (Atlas,

1991). Therefore, bioremediation technology causes the improvement of the natural capacity of microorganisms to degrade contaminants (Catallo and Portier, 1992; Atlas, 1991; Salanitro et al., 1997). Variety of bioremediation methods have been developed to support and increase the degradation activities of native microbial populations, allowing reduction in time and subsequent saving in costs. The two main approaches to bioremediation are the bio-stimulation and bioaugmentation (Korda et al., 1997).

On the other hand, hydrocarbon-degrading microorganisms are ubiquitous in most ecosystems where contaminants may serve as organic carbon sources (Atlas and Bartha, 1992; Margesin et al., 2000). Bacteria are the most active agents in petroleum biodegradation and there is evidence of their fundamental role as primary degraders

of spilled oil (Komukai-Nakamura et al., 1996; Ijah, 1998; Rahman et al., 2002; Head et al., 2006; da Cruz et al., 2011; Oliveira et al., 2012). Effect of various nutrients on the degradation of crude oil by different bacteria was investigated by several scientists (Gibbs, 1975; Dibble and Bartha, 1979; Wrenn et al., 1994; Oh et al., 2001; Berwick, 2004; Xu et al., 2005). Several factors, both physico-chemical and biological, affect the rate of microbial degradation of hydrocarbons in soil. Recently, growing interest in the use of several *Pseudomonades* during degradation of crude oil have been reported (Bosch et al., 1999; Evans et al., 2004; Wongsu et al., 2004; Meng et al., 2005; Emtiazzi et al., 2005; Toledo et al., 2006; Song et al., 2006; Ueno et al., 2006; Das and Mukherjee, 2007; Mittal and Singh, 2009). However, application of statistical experimental design for optimization of crude oil degradation with *Pseudomonas* sp. was rarely investigated. Recently, medium optimization for a novel crude-oil degrading lipase from *Pseudomonas aeruginosa* SL-72 using statistical approaches for bioremediation of crude-oil was reported (Nain et al., 2012).

On the other hand, experimental design techniques present a more balanced alternative to one-variable-at-a-time approach (OVAT) in which single factor is varied, while others are kept fixed. However, Plackett and Burman design comprise one type of two-level screening and can be constructed on the basis of fractional replication of a full factorial design (Plackett and Burman, 1947). This design is appropriate to face the large number of cultivation conditions under investigation and allow obtaining an unbiased estimates of linear effects of all factors with maximum accuracy for a given number of observations (Akhazarova and Kafarov, 1982).

The main aim of this work was to investigate the possible improvement of crude oil degradation by *Pseudomonas* sp. strain-O2 and to evaluate the influence of different cultivation condition on efficiency of crude oil degradation. Preliminary controlled experiments were conducted to address the most effective nitrogen source and its optimal concentration, yeast extract, $MgSO_4 \cdot 7H_2O$ and the level of phosphate salts that might affect crude oil degradation. To determine the significance among other physical and nutritional requirement, fractional factor design namely; Plackett-Burman experimental design was applied and the significant variables were determined.

MATERIALS AND METHODS

Microorganism and cultivation medium

Bacterial strain used in this study, *Pseudomonas* sp. strain-O2 was isolated and identified as previously mentioned (Mostafa et al., 2012). The bacterium was grown on minimal salts medium (MSM), it was the modified medium of Ijah (1998) with the following composition, (g/L): yeast extract, 0.5; NaCl, 0.5; $(NH_4)_2SO_4$, 2; $MgSO_4 \cdot 7H_2O$, 0.2; K_2HPO_4 , 5; KH_2PO_4 , 2 and trace elements (with the following composition, (g/L): $FeSO_4$, 5; H_3BO_4 , 0.025; $CuSO_4 \cdot 5H_2O$, 0.005; KI, 0.005; $CoSO_4$, 0.3; $MnSO_4 \cdot 4H_2O$, 3; $ZnSO_4 \cdot 7H_2O$, 5; $NaMoO_4$, 0.012, and distilled water up to 1 L), 0.1 mL.

Monitoring of crude oil biodegradation

Biodegradation of crude oil was investigated by cultivation of the bacterium in 250-mL Erlenmeyer flasks containing 100 mL sterile MSM amended with 0.92% (w/v) crude oil as sole carbon source. 1.5 mL of active inoculum, prepared from 24 h preculture of the bacterial strain grown on nutrient broth medium, was used for inoculation. All flasks were incubated at 30°C under shake conditions at 120 rpm. At the end of 4 days incubation period, flasks were analyzed for the residual crude oil as described below.

Extraction of crude oil

For extraction of the residual crude oil remaining at the end of cultivation period, each flask was acidified with 5 mL H_2SO_4 (1:1) pH 2; then extracted three times by 60 mL methylene chloride in a 250 mL separating funnel, the organic layer was drained through a funnel containing anhydrous sodium sulfate into a 50 mL boiling rounded flask; then moved to a rotary evaporator at about 40°C to reduce the volume of the extract to 1 mL. The residual oil was transferred to a pre-weighed 2 mL vial for gravimetric analysis, where the amount of crude oil remaining was determined and the percent of crude oil removal was calculated.

Influence of medium composition on crude oil degradation

A series of preliminary on-variable-at-a-time (OVAT) experiments were carried out to provide information for determination of settings of variables that might be used during experimental design. The effect of different N-sources was firstly investigated, nitrogen sources used (2 g/L) were; peptone, ammonium sulfate, ammonium chloride or sodium nitrate. Other tested conditions included; different levels of peptone (2 to 5 g/L), inoculum concentration (0.5 to 2% v/v), yeast extract (0 to 2.5 g/L), $MgSO_4 \cdot 7H_2O$ (0.1 to 0.5 g/L) and phosphate salts K_2HPO_4 and KH_2PO_4 (1- to 5-folds) the basal concentration (2:0.8 g/L).

Plackett-Burman design

Plackett-Burman experimental design most commonly used for screening purpose was applied to evaluate the significance of various medium components as well as environmental factors affecting crude oil degradation by *Pseudomonas* sp strain-O2. The different factors were prepared in two levels: -1 for low level and +1 for high level, based on Plackett-Burman statistical matrix design, which is a fraction of a two-level factorial design and allows the investigation of n-1 variables in at least n-experiments (Plackett and Burman, 1947). Eleven independent variables (Table 1) were screened in 14 combinations according to the design shown in Table 2. All trials were performed in triplicate and the average of observation was considered as the final result. The main effect of each variable was calculated simply as the difference between the average of measurements made at the high setting (+1) and the average of measurements observed at low setting (-1) of that factor. Plackett-Burman experimental design is based on the first order model (equation 1):

$$Y = \beta_0 + \sum \beta_i x_i \quad (1)$$

Where Y is the predicted response (% removal of crude oil), β_0 , β_i are constant coefficients, and x_i is the coded independent variables estimates or factors.

Analysis of data

The data of crude oil degradation was statistically analyzed.

Table 1. Variables and their levels employed in Plackett-Burman design for screening of culture conditions affecting crude oil degradation by *Pseudomonas* sp. strain-O2.

Code	Variable	Value	
		-1	+1
X ₁	Inoculum concentration	0.5	1.5
X ₂	Ammonium sulfate (g/L)	0.5	2
X ₃	Peptone (g/L)	2	5
X ₄	Yeast Extract (g/L)	0	5
X ₅	MgSO ₄ .7H ₂ O (g/L)	0.2	0.5
X ₆	K ₂ HPO ₄ (g/L)	2	5
X ₇	KH ₂ PO ₄ (g/L)	0.8	2
X ₈	NaCl (g/L)	5	2
X ₉	Agitation (rpm)	60	120
X ₁₀	Oil Concentration (g/L)	0.5	1.5
X ₁₁	Na-succinate (g/L)	0.5	2

Essential Experimental Design free software (Steppan et al., 2000) was used for data analysis and determination of coefficients. Factors having highest t-value and confidence level over 95% were considered to be highly significant on crude oil degradation.

RESULTS

Influence of medium composition by OVAT

Nitrogen, phosphate and sulfate levels

To provide information on the variable levels that might be used in experimental design study, a series of preliminary OVAT experiments were carried out. At the end of each experiment, cells were separated and the residual crude oil was estimated. Results in Figure 1 indicated that peptone was the optimal nitrogen source recording 73.3% removal of crude oil. Inorganic nitrogen sources such as; ammonium sulfate and sodium nitrate showed positive significance on the growth and crude oil degradation and recorded 71.1 and 69.6% removal of crude oil, respectively. Also, 2 g/L was the optimal peptone concentration, any further increase in concentration led to reduction in crude oil degradation efficiency (Table 3). On the other hand, to determine the most suitable level of yeast extract, MSM was amended with different concentrations of yeast extract ranging from 0 to 2.5 g/L. Results in Table 3 indicated that 0.5 g/L was the optimal concentration that led to approximately 83% removal of crude oil. Approximately, 60.4 and 52.9% removal of crude oil concentration was recorded when yeast extract was increased to 1 or 2.5 g/L, respectively.

Furthermore, one of the crucial chemical constituents in the medium that affect crude oil degradation is the phosphate salt. Results in Table 4 indicated that the increase of phosphate ratio to 2.5 led to 80.1% removal of amount of crude oil. Further increase or decrease in phosphate

salt ratio led to decrease in oil degradation efficiency and hence in percent removal of crude oil. Indeed, the optimal concentration of MgSO₄.7H₂O was 0.5 g/L recording approximately 60% removal of the crude oil (data not shown).

It is known that the concentration of the bacterial inoculum plays an important role in determining the efficiency of crude oil degradation. Results revealed that 1.5% (v/v) was the optimal inoculum concentration that led to approximately 80% removal of crude oil (data not shown).

Evaluation of cultivation condition by FFD

Fractional factorial design (FFD) is a kind of experimental design that enables researchers to evaluate the most significant factors affecting the process with the least number of trials. Plackett-Burman design is a FFD, which succeeds in ranking factors from different categories to enable better understanding of the medium effects, eleven factors were studied through the application of Plackett-Burman design. Evaluation of process variables was carried out according to the experimental matrix presented in Table 2, where the residual crude oil estimated and the percent of crude oil removal was the measured response. Variation in crude oil degradation expressed as % of crude oil removal (73 - 96.6%) is shown in Table 2. Results collectively showed that the highest crude oil removal of 96.6 was obtained in the combination number 9, while the lowest crude oil removal was obtained in combination numbers 6. Statistical analysis of these data revealed that the value of determination coefficient R², that measures the goodness of the model fitting, is >0.99. This indicates that less than 1% of the total variations are not explained by the model, which ensures the good adjustment of the model.

Moreover, the main effects of the examined variables on degradation of crude oil were calculated and illustrated graphically in Figure 2a.

On analysis of regression coefficients and *t*-value (Table 5), it was clear that mainly crude oil concentration, together with MgSO₄.7H₂O, (NH₄)₂SO₄, KH₂PO₄ and Na-succinate were found to be the most significant variables that encourage crude oil degradation in *Pseudomonas* sp. strain-O2. Whereas, bacterial inoculum, yeast extract concentration, agitation and NaCl were the lowest significant variables that discourage crude oil degradation.

The determination coefficient represents the quality of fitting the polynomial model, which can be represented as follows:

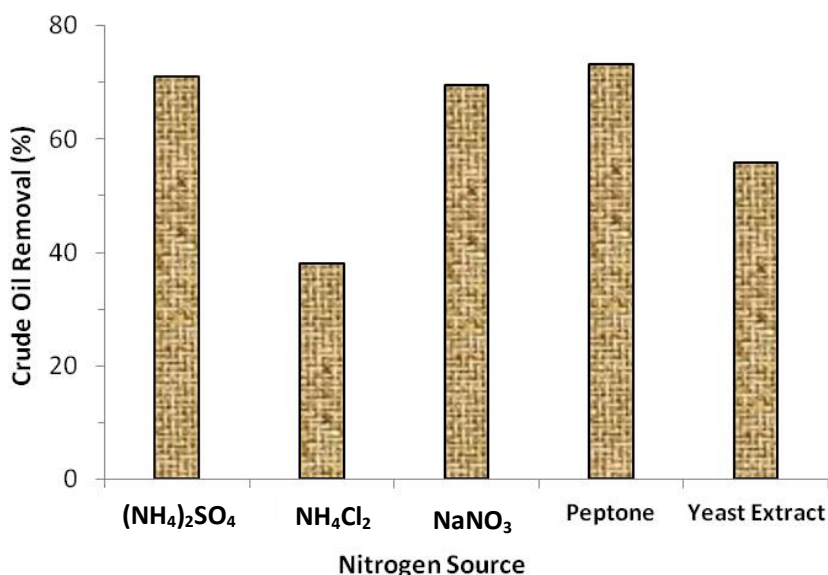
$$Y_{\% \text{ removal}} = 11.70 - 2.88X_1 + 0.7X_2 + 0.07X_3 - 4.16X_4 + 1.71X_5 - 0.46X_6 + 0.85X_7 - 0.29X_8 - 0.54X_9 + 6.11X_{10} + 0.52X_{11}$$

One of the advantages of the Plackett-Burman design is to rank the effect of different variables on the measured response independent on its nature (either nutritional or

Table 2. Plackett-Burman experimental design for evaluation of factors affecting crude oil degradation by *Pseudomonas* sp. strain-O2.

Experiment	X1	X2	X3	X4	X5	X6	X7	X8	X9	X10	X11	Oil removal (%)
1	1	-1	-1	-1	1	1	1	-1	1	1	-1	80.7
2	1	1	1	-1	1	1	-1	1	-1	-1	-1	92.3
3	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	-1	90.0
4	0	0	0	0	0	0	0	0	0	0	0	88.0
5	1	-1	1	1	-1	1	-1	-1	-1	1	1	91.5
6	-1	1	1	-1	1	-1	-1	-1	1	1	1	73.0
7	1	1	-1	1	1	-1	1	-1	-1	-1	1	96.4
8	1	1	-1	1	-1	-1	-1	1	1	1	-1	92.4
9	-1	-1	-1	1	1	1	-1	1	1	-1	1	96.6
10	1	-1	1	-1	-1	-1	1	1	1	-1	1	94.7
11	-1	-1	1	1	1	-1	1	1	-1	1	-1	81.4
12	0	0	0	0	0	0	0	0	0	0	0	88.0
13	-1	1	-1	-1	-1	1	1	1	-1	1	1	75.0
14	-1	1	1	1	-1	1	1	-1	1	-1	1	96.3

Variables coded as follows: X₁, inoculum concentration; X₂, ammonium sulfate; X₃, peptone; X₄, yeast extract; X₅, MgSO₄·7H₂O; X₆, K₂HPO₄; X₇, KH₂PO₄; X₈, NaCl; X₉, Agitation; X₁₀, Oil concentration; X₁₁, Na-succinate.

**Figure 1.** Effect of different nitrogen sources on degradation of crude oil by *Pseudomonas* sp. strain-O2.

physical factor) or sign (whether contributes positively or negatively). Interestingly, Figure 2b shows the ranking of factor estimates in a Pareto chart. The Pareto chart displays the magnitude of each factor estimate and is a convenient way to view the results of Plackett-Burman design (Strobel and Sullivan, 1999).

DISCUSSION

Given the environmental importance of bacteria, especially *Pseudomonas*, in degradation of crude oil and

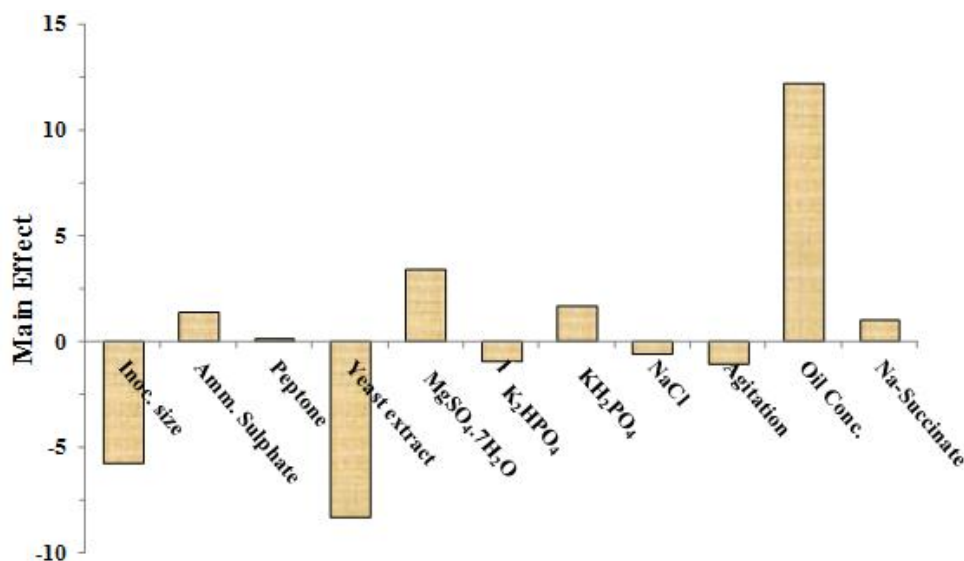
their application in bioremediation and namely, bioaugmentation and biostimulation, the influence of different growth parameters on degradation process was closely examined. In a series of controlled OVAT experiments, optimal concentrations of medium constituents were closely investigated. Preliminary investigations indicated that peptone is the optimal organic nitrogen source. Supplementation of the medium with yeast extract enhanced growth and % removal of crude oil from the culture medium reflecting its importance as nitrogen and vitamin source. The use of yeast extract as nitrogen as

Table 3. Effect of different peptone and yeast extract concentrations on crude oil degradation by *Pseudomonas* sp. strain-O2.

Peptone concentration (g/L)	Extracted crude oil (mg)	Crude oil removal (%)	Yeast extract concentration (g/L)	Extracted crude oil (mg)	Crude oil removal (%)
2	16.3	82.3	0	18.0	80.1
3	28.9	68.6	0.5	15.6	83.0
4	37.9	58.8	1.0	36.4	60.4
5	21.3	76.8	2.5	43.3	52.9

Table 4. Effect of different phosphate ratios on crude oil degradation by *Pseudomonas* sp. strain-O2.

K ₂ HPO ₄ : KH ₂ PO ₄ (g/L)	Fold	Extracted crude oil (mg)	Crude oil removal (%)
2 : 0.8	1	46.2	50.6
4 : 1.6	2	29.5	49.8
5 : 2	2.5	18.3	80.1
6 : 2.4	3	23.6	74.3
7 : 2.8	3.5	23.2	74.8
9 : 3.6	4.5	35.2	61.7
10 : 4	5	37.7	59.0

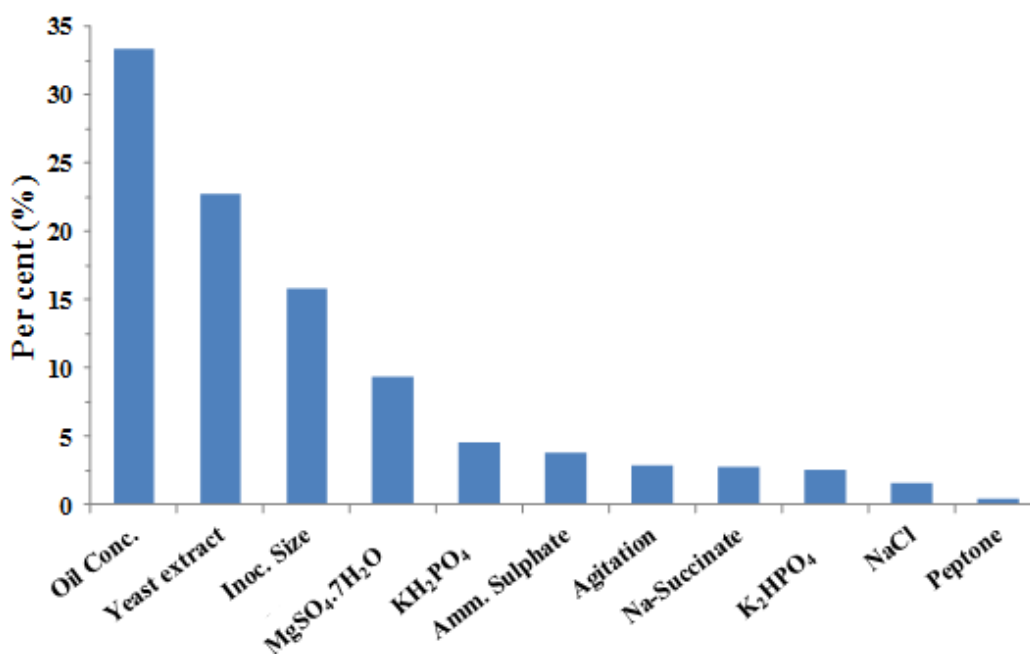
**Figure 2a.** Effect of different factors on crude oil degradation by *Pseudomonas* sp. strain-O2 as screened with Plackett-Burman design.

well as vitamin source during hydrocarbon and petroleum degradation by many organisms was reported (Lemos et al., 2002; Singh et al., 2005). Interestingly, increase in yeast extract concentration from 0.5 to 2.5 g/L led to approximately 20% reduction in crude oil removal. In concordance, results of Plackett-Burman experiments revealed that the increase in yeast extract concentration negatively affected crude oil degradation. On the other hand, one of the crucial chemical constituents in the

medium that affect crude oil degradation is the phosphate salt, especially due to its suggested importance in indirect control of pH and providing the organism with the required phosphate ions necessary for energy and ATP production. Indeed, buffering effect of phosphate salts during degradation of crude oil was reported by Emtiazi et al., (2005). This finding supported by the results of fractional factorial design which indicated that phosphate ion (KH₂PO₄), contributed positively to crude oil biode-

Table 5. Statistical analysis of data obtained from Plackett-Burman design showing coefficient values, t- and P-values of each variable.

Variable	Statistical analyse			
	Coefficient	t-Stat	P-value	Coefficient level (%)
X ₁	-2.88	9.98	0.009	98.4
X ₂	0.70	-2.43	0.135	04.0
X ₃	0.07	-0.24	0.828	99.8
X ₄	-4.16	14.3	0.004	99.4
X ₅	1.71	-5.91	0.027	93.5
X ₆	-0.46	1.59	0.251	96.0
X ₇	0.85	-2.95	0.098	89.0
X ₈	-0.28	0.99	0.424	99.2
X ₉	-0.53	2.00	0.183	95.6
X ₁₀	6.11	-21.1	0.002	96.1
X ₁₁	0.52	-1.54	0.261	48.4

**Figure 2b.** Pareto chart rationalizing the effect of each variable on crude oil degradation by *Pseudomonas* sp. strain-O2.

gradation and showed high significant effect reflected by the p-value (0.098). Furthermore, supplementation of fermentation medium with inorganic nitrogen source such as $(\text{NH}_4)_2\text{SO}_4$ contributed positively to crude oil degradation, consequently can be used instead of other organic nitrogen sources. In accordance, Nain (2012) reported that ammonium and phosphate ions contributed positively to production of crude oil degrading lipase from *P. aeruginosa* SL-72 during application of Plackett-Burman experimental design. The use of inorganic nitrogen sources, such as ammonium chloride and potassium nitrate, was preferably used during crude oil degradation (Wrenn

et al., 1994). Many scientists reported the importance of amendment of biodegradation medium with phosphate and inorganic nitrogen fertilizers (Westlake et al., 1978; Jobson et al., 1974; Piehler and Paerl, 1996; Emtiazi et al., 2005; Margesin et al., 2007). Furthermore, it was clear that the degradation efficiency was positively affected by the amount of crude oil due to the increase in the amount of C-source (p-value= 0.002). Similar finding was reported by Margesin et al. (2007) and Xu et al. (2005). Addition of Na-succinate as co-substrate to enhance crude oil degradation resulted in a slightly positive effect. Simple co-substrates were reported to

positively affect the rate of hydrocarbon degradation. Significant increase in degradation of crude oil as well as the saturated branched hydrocarbon (squalane) by addition of glycerol, rhamnolipid or Na-succinate was reported (Berekaa and Steinbuchel, 2000; Meng et al., 2005). Successful degradation of crude oil PAHs by co-metabolism was reported (Arun et al., 2011). Whereas, the other variables namely, agitation, NaCl and K_2HPO_4 were the lowest significant variables affecting crude oil degradation. Therefore, might be dropped in further optimization experiments.

The results of this study collectively revealed the possible optimization of crude oil degradation by *Pseudomonas* sp. strain-O2 through improvement of chemical and environmental parameters. Preliminary experiments gave an idea about the setting of some variables that might be used in experimental design. Plackett-Burman design showed that the most significant variables encourage crude oil degradation. Furthermore, the design succeeded to rank factors from different categories to enable better understanding of the medium effect. It is worthwhile to further optimize the significant variables determined in the present study to attain maximum crude oil degradation by applying other suitable statistical designs.

REFERENCES

- Akhazarova S, Kafarov V (1982). Experimental optimization. In: Chemistry and Chemical Engineering. Moscow: Mir Publishers.
- Arun K, Ashok M, Rajesh S (2011). Crude oil PAH constitution, degradation pathway and associated bioremediation microflora: an overview. *Int. J. Environ. Sci.* 7:1420-1439.
- ASTM (1995). Standard Guide for Risk-based Corrective Action Applied at Petroleum Release Sites, American Society for Testing and Materials, West Conshohocken, PA 19428, USA.
- Atlas RM, Bartha R (1992). Hydrocarbon biodegradation and oil spill bioremediation. *Advanc. Microbial Ecol.* 12: 287–338.
- Atlas RM (1991). Microbial hydrocarbon degradation-bioremediation of oil spills. *J. Chem. Technol. Biotechnol.* 52: 149-156.
- Berekaa MM, Steinbuchel A (2000). Microbial degradation of the multiply branched alkane 2,6,10,15,19,23-Hexamethyltetracosane (Squalane) by *Mycobacterium fortuitum* and *Mycobacterium ratisbonense*. *Appl. Environ. Microbiol.* 66: 4462-4467.
- Berwick GP (2004). Physical and chemical conditions for microbial oil degradation. *Biotech. Bioeng.* 26: 1294 – 1305.
- Boopathy R, (2000). Factors limiting bioremediation technologies. *Bioresource Technol.* 74: 63–67.
- Bosch R, Moore ERB, Garcia-valdes E, Pieper DH (1999). NahW, a novel, inducible salicylate hydroxylase involved in mineralization of Naphthalene by *Pseudomonas stutzeri* AN10. *J. Bacteriol.* 181: 2315-2322.
- Catallo WJ, Portier RJ (1992). Use of indigenous and adapted microbial assemblages in the removal of organic chemicals from soils and sediments. *Water Sci. Technol.* 25: 229–237.
- Da Cruz FG, de Vasconcellos PS, Angolini FFC, Dellagnezze MB, Garcia NSI, de Oliveira MV, Neto SDVE, Marsaioli JA (2011). Could petroleum biodegradation be a joint achievement of aerobic and anaerobic microorganisms in deep sea reservoirs?. *AMB Express* 1:47.
- Das K, Mukherjee AK (2007). Crude petroleum-oil biodegradation efficiency of *Bacillus subtilis* and *Pseudomonas aeruginosa* strains isolated from petroleum-oil contaminated soil from North-East India. *Bioresour. Technol.* 98: 1339-45.
- Dibble JT, Bartha R (1979). Effects of environmental parameters on biodegradation of oil sludge. *Appl. Environ. Microbiol.* 37: 729-739.
- Emtiaz G, Shakarami H, Nahvi I, Mirdamadian SH (2005). Utilization of petroleum hydrocarbons by *Pseudomonas* sp. and transformed *Escherichia coli*. *African J. Biotechnol.* 4: 172-176.
- Evans FF, Seldin L, Sebastin GV, Kjelleberg S, Holmstrom C, Rosado AS (2004). Influence of petroleum contamination and Biostimulation treatment on the biodiversity of *Pseudomonas* spp. In soil microcosms as evaluated by 16S rRNA based-PCR and DGGE. *Let. Appl. Microbiol.* 38: 93-8.
- Gibbs CF (1975). Quantitative studies on marine biodegradation of oil. I. Nutrient limitation at 14°C. *Proc. R. Soc. London, Ser. B.* 188(1090): 61–82.
- Head IM, Jones DM, Röling WFM (2006). Marine microorganisms make a meal of oil. *Nature Rev. Microbiol.* 4: 173-182.
- Ijah JJ (1998). Studies on relative capabilities of bacterial and yeast isolates from tropical soil in degrading crude oil. *Waste Manage.* 18: 293-299.
- Jobson AM, McLaughlin FD, Cook DW, Westlake (1974). Effect of amendments on the microbial utilization of oil applied to soil. *Appl. Microbiol.* 27: 166-71.
- Komukai-Nakamura S, Sugiura K, Yamauchi-Inomata Y, Toki H, Venkateswaran K, Yamamoto S, Tanaka H, Harayama S (1996). Construction of bacterial consortia that degrade Arabian Light crude oil. *J. Ferment. Bioeng.* 82: 570–574.
- Korda A, Santas P, Tenente A, Santas R (1997). Petroleum hydrocarbon bioremediation: sampling and analytical techniques, in situ treatments and commercial microorganisms currently used. *Appl. Microbiol. Biotechnol.* 48: 677–686.
- Lemos SJL, Rizzo AC, Millioli VS, Soriano AU, de Moura Sarquis MI, Santos R (2002). Petroleum degradation by filamentous fungi. The 9th Annual International Petroleum Environmental Conference, 22-25 Oct. 2002, Albuquerque, NM. Integrated Petroleum Environmental Consortium (IPEC), Univ. Of Tulsa, OK. p. 10.
- Margesin R (2000). Potential of cold-adapted microorganisms for bioremediation of oil-polluted Alpine soils. *Int. Biodet. Biodegr.* 46: 3–10.
- Margesin R, Hammerle M, Tscherko D (2007). Microbial activity and community composition during bioremediation of diesel-oil-contaminated soil: effects of hydrocarbon concentration, fertilizers, and incubation time. *Microb. Ecol.* 53: 259-69.
- Meng Q, Zhang G, Wu Y, Qian X (2005). Biodegradation of crude oil by *Pseudomonas aeruginosa* in the presence of rhamnolipids. *J. Zhejiang Univ. Sci.* 6B:725-730.
- Mittal A, Singh A (2009). Isolation of hydrocarbon degrading bacteria from soil contaminated with crude oil spills. *Indian J. Experiment. Biol.* 47:760-765.
- Mostafa AR, Blumenberg M, Berekaa MM, Michaelis W, Alaa AR (2012). Crude oil biodegradation efficiency of bacterial strains isolated from oil contaminated sediment from Western harbour of Alexandria, Egypt. Submitted for publication.
- Nain L, Verma S, Saxena J, Prasanna R, Sharma V (2012). Medium optimization for a novel crude-oil degrading lipase from *Pseudomonas aeruginosa* SL-72 using statistical approaches for bioremediation of crude-oil. *Biocatal. Agricul. Biotechnol.* 1: 321–329.
- Oh YS, Sim DS, Kim JJ (2001). Effects of nutrients on crude oil biodegradation in the upper intertidal zone. *Mar. Pollut. Bull.* 42: 1367-72.
- Oliveira LFP, Vasconcellos PS, Angolini FFC, da Cruz FG, Marsaioli JA, Neto VSE, Oliveira MV (2012). Taxonomic diversity and biodegradation potential of bacteria isolated from oil reservoirs of an offshore Southern Brazilian Basin. *J. Pet. Environ. Biotechnol.* 3(7) : 132.
- Piehler MF, Paerl HW (1996). Enhanced biodegradation of diesel fuel through the addition of particulate organic carbon and inorganic nutrients in coastal marine waters. *Biodegradation* 7: 239-47.
- Plackett RL, Burman JP (1947). The design of optimum multifactorial experiments. *Biometrika* 33: 305-325.
- Rahman KSM, Thahira-Rahman J, Lakshmanaperumalsamy P, Banat IM (2002). Towards efficient crude oil degradation by a mixed bacterial consortium. *Bioresour. Technol.* 85: 257-261.
- Ruiz M, Pasadakis N, Kalograkis N (2006). Bioremediation and toxicity

- determination of natural seawater polluted with weathered crude oil by salt-tolerant consortia. *Marine Pollution* 52: 1490–1493.
- Salanitro JP, Dorn PB, Hueseman MH, Moore KO, Rhodes IA, Jackson LMR, Vipond TE, Western MM, Wisniewski HL (1997). Crude oil hydrocarbon bioremediation and soil ecotoxicity assessment. *Environ. Sci. Technol.* 31: 1769–1776.
- Singh A, Ward OP, Kuhad RC (2005). Feasibility studies for microbial remediation hydrocarbon contaminated soil. In: *Manual of soil analysis, monitoring and assessing soil bioremediation*. Margesin P. and F. Schinner, (eds), 5: 131-153, Springer Berlin, Heidelberg.
- Song R, Hua Z, Li H, Chen J (2006). Biodegradation of petroleum hydrocarbons by two *Pseudomonas aeruginosa* strains with different uptake modes. *J. Environ. Sci. Health Tox. Hazard Subst. Environ. Eg.* 41: 733-48.
- Steppan D, Werner J, Yeater B (2000). Essential Regression and Experimental Design in MS Excel-free, user-friendly software package for doing multiple linear regression, step-wise regression, polynomial regression, model adequacy checking and experimental design in MS Excel. <http://www.geocities.com/SiliconValley/Network/1032/>.
- Strobel RJ, Sullivan GR (1999). Experimental design for improvement of fermentations. In: *Manual of industrial Microbiology and Biotechnology* (Demain, A. L., Davies, J. E., eds): 80-93. Washington: ASM Press.
- Toledo FL, Calvo C, Rodelas B, Gonzdzlezlopez J (2006). Selection and identification of bacteria isolated from waste crude oil with polycyclic aromatic hydrocarbons removal capabilities. *Syst. Appl. Microbiol.* 29: 244-52.
- Ueno A, Ito Y, Yamamoto Y, Yumoto I, Okuyama H (2006). Bacterial community changes in diesel-oil-contaminated soil microcosms stimulation with Luria-Bertani medium or bioaugmentation with petroleum-degrading bacterium *Pseudomonas aeruginosa* strain WatG. *J. basic Microbiol.* 46: 310-7.
- Westlake DW, Jobson AM, Cook FD (1978). In situ degradation of oil in a soil of the boreal region of the Northwest Territories. *Can. J. Microbiol.* 24: 254-60.
- Wongsa P, Tanaka M, Ueno A, Hasanuzzaman M, Yumoto I, Okuyama H (2004). Isolation and characterization of novel strains of *Pseudomonas aeruginosa* and *Serratia marcescens* possessing high efficiency to degrade gasoline, kerosene, diesel oil and lubricating oil. *Curr. Microbiol.* 49: 415-22.
- Wrenn AB, Haines RJ, Venosa DA, Kadkhodayan M, Suidan TM (1994). Effects of nitrogen source on crude oil biodegradation. *J. Ind. Microbiol. Biotechnol.* 13: 279-286.
- Xu R, Long LC, Lim YG, Obbard JP (2005). Use of slow-release fertilizer and biopolymers for stimulating hydrocarbon biodegradation in oil-contaminated beach sediments. *Mar. Pollut. Bull.* 51: 8-12.

Full Length Research Paper

Real time polymerase chain reaction for identification of *Candida* species isolated from blood stream infections in non-neutropenic cancer patients

Rasha A. Alm El-Din*, Hanan El-Bassat, Mohamed El-Bedewy and Mohamed A. Alm El-Din

Medical Microbiology and Immunology, Tropical Medicine, Internal Medicine, Clinical Oncology Departments, Faculty of Medicine, Tanta University, Egypt.

Accepted 26 August, 2013

This work aimed at identifying *Candida* species isolated from blood stream infection in non-neutropenic cancer patients by conventional and molecular methods. Out of 200 blood samples from cancer patients with blood stream infection, 89 were positive for *Candida* which were identified using chromogenic medium (CHROMagar *Candida*), and real-time polymerase chain reaction (RT-PCR) assays targeting the fungal ITS2 region using a LightCycler instrument. The present study revealed that non-*albicans Candida* species caused most of the cases of candidaemia in the cancer patients. A total of 89 *Candida* isolates were obtained, *Candida albicans* represent 15/89 patients (16.85%) and non-*albicans Candida* species accounts for 74/89 patients (83.1%) of which *Candida tropicalis* represent 32.6% followed by *Candida parapsiliosis* 23.6%, and then *Candida rugosa* (21.4%) and the lowest percentage was in *Candida glabrata* (5.6%). The results of the study revealed that real time-PCR could detect 81 out of 89 cases of candidaemia with sensitivity of 91.01%, specificity of 95.6%, PPV of 94.18% and NPV of 92.30%. The high prevalence of non-*albicans* candidaemia should be considered prior to antimicrobial therapy in non-neutropenic cancer patients with candidaemia. Real-time PCR represents a rapid and accurate method for diagnosis of candidaemia.

Key words: Candidaemia, *Candida albicans*, real-time polymerase chain reaction (RT-PCR).

INTRODUCTION

Candida species are currently the fourth most common cause of bloodstream infections worldwide, and the third most common cause of bloodstream infections in patients with malignancy (Chung et al., 2005). Candidiasis is not only associated with mortality of about 30 to 40% in cancer patients, but also extends the duration of hospital stay and increases the cost of medical care (Naglik et al., 2004). The clinical presentations of patients with blood stream infection caused by *albicans* and non-*albicans Candida* species are indistinguishable. However, non-

albicans candida species are often less susceptible to fluconazole than *C. albicans* and may require a greater dosage of antifungals to cure clinically (Chakrabarti et al., 2008). Early diagnosis of invasive fungal infections is important in the management of invasive fungal infection and in improving the outcome and preventing relapse of the underlying malignancy (Verma et al., 2003).

Previous studies have revealed blood stream infection rates ranging from 4 to 22% due to *Candida* species in cancer patients. The rise in incidence of non-*albicans*

Table 1. Sequence and concentration of the primers, probe and probe specific T_m in real time PCR assays (Hebart et al., 2000).

Probre or primer	Sequence (concn)	Probe specific T_m (°C)
Candida-specific primers	CCTGTTTGAGCGTCRTTT (0.15 μ M)	
	TCCTCCGCTTATTGATAT (0.5 μ M)	
Candida-specific probe		
C.alb-S	Cy5-CATTGCTTGC GGCGGTA-biotin (0.2 μ M)	66 or 55
C.glab-S	Cy5GTTTTACCAACTCGGTGTTGAT-biotin (0.2 μ M)	65
C.trop-S	Cy5-GGCCACCACAATTTATTTCA-biotin (0.2 μ M)	63
C.rugosa-S	Cy5-CGAGCGAACTAGACTTT-biotin (0.2 μ M)	60
C.para-S	Cy5-GAAAGCGGAGTATAAAC-biotin (0.2 μ M)	58

candidaemia and the emergence of antifungal resistance have further fuelled the need to carry out such a study (Halliday et al., 2005).

Azoles, antifungal agents have therapeutic activity against different *Candida* species. Among the azoles drugs, fluconazole shows that good tolerance has appeared and antifungal drug resistance is quickly becoming a major problem especially in immuno-compro-mised patients (Xu et al., 2000). This resistance also favors the emergence of *C. krusei* and *C. glabrata* (Cirak et al., 2003).

Conventional fungal identification methods may require 48 to 72 h or longer in order to yield definitive identifications, leading to significant morbidity and mortality that is associated with invasive *Candida* infections (Boyanton et al., 2008). Real time-polymerase chain reaction (PCR) technology can directly detect the presence of fungi in hours with high level of sensitivity and specificity (Chen et al., 2000; Loeffler et al., 2000).

The identification of species via melting curve analysis with species-specific hybridization probes further increases specificity. The fast turnaround time of less than 2 h is another advantage of the real-time PCR technology (Verweij and Meis, 2000). The aim of this study was to detect the incidence of *Candida* spp. in non-neutropenic cancer patients and evaluate the role of real time PCR in diagnosis of these cases.

MATERIALS AND METHODS

An approval was gotten from the ethical committee in Tanta Faculty of Medicine and a written consent from all participants in this study was performed over 12 months from March 2012 to March 2013 on cancer patients complaining of blood stream infection caused by *Candida* spp. (candidemia). An episode of candidaemia was defined as the isolation of *Candida* species from a single positive blood culture. 6 ml blood was taken from each patient; 3 ml were used for microbiological study using standard cultivation methods and 3 ml with citrate anticoagulant stored at -20°C until it was used for DNA extraction and real time-PCR.

Blood culture and organism identification

For diagnosis of candidaemia, 3 ml blood was collected in BacT/ALERT FA aerobic blood culture bottles (bioMérieux). Blood

culture bottles were subcultured onto Sabouraud's dextrose agar. *Candida* cultures were considered positive if a 10 CFU appeared on the plate. Identification of these isolates was carried out by growth on the chromogenic medium CHROMagar by colony morphology and pigmentation according to the manufacturer's instructions and biochemical reactions (Forbes et al., 2002).

Real time-PCR analysis

From blood samples, DNA was extracted with a High Pure PCR template preparation kit by following the instructions of the manufacturer. DNA was eluted with 100 μ l elution buffer and stored at -20°C until use (Hebart et al., 2000).

The 20 μ l real-time PCR mixtures were prepared with 2 μ l of LightCycler-FastStart DNA master SYBR green I (Roche Molecular Biochemicals, Mannheim, Germany), 4 mM MgCl_2 , primers and probes as shown in Table 1, and 3 μ l of DNA extract made up to 20 μ l with water. PCR was performed in a LightCycler instrument with preliminary denaturation for 10 min at 95°C , followed by 60 amplification cycles (with a temperature transition rate of $20^{\circ}\text{C}/\text{s}$) of denaturation at 95°C for 8 s, annealing at 55°C for 10 s, and primer extension at 72°C for 10 s, with a single fluorescence acquisition step at the end of the extension. This was followed by a melting analysis of the probe-PCR product consisting of 95°C for 30 s and then cooling to 35°C for 60 s before the temperature was raised to 98°C at a rate of $0.2^{\circ}\text{C}/\text{s}$ with continuous fluorescence acquisition. A final cooling step was performed at 40°C for 10 s. Samples were considered positive for *Candida* species upon the presence of a biprobe-specific melting peak. Specific T_m values of the biprobes are shown in Table 1.

RESULTS

During a 12 months period (from March 2012 to March 2013), out of 200 cancer patients with blood stream infection admitted to Oncology and Internal Medicine Departments in Tanta University Hospital, 89 were diagnosed with candidaemia were eligible for the study. 43 patients (48.3%) out of 89 were males and 46 (51.6%) were females, ranging in age from 15-79 (mean \pm SD, 49 ± 3.49) years. Gastrointestinal and breast cancer were the most frequent among the studied group accounting for 35.9 and 26.9%, respectively. The mean of the hospital stay was 3.58 (range; 1-9) days. Tables 2 and 3 present demographic and clinical characteristics of cancer patients with candidaemia.

Table 2. Demographic characteristics of cancer patients with candidiasis.

Patient characteristic	Number (%)
sex	
Male	43 (48.3)
Female	46 (51.6)
Age (years)	
Range	15-79
mean±SD	49 ±3.49
Days admitted in hospital (days)	
Range	1-9
Mean±SD	3 ±4.45
Cancer type	
GIT	32 (35.9)
breast	24 (26.9)
lung	10 (11.23)
Head and neck	12 (13.48)
Others	11 (12.35)
total	89 (100)

Table 3. Clinical criteria of patients with candidaemia.

Patients with candidaemia	<i>C. albicans</i>	Non <i>C. albicans</i>
No. 89	No.15 (16.85%)	No.74 (83.15%)
Age (years)	36.07±17.4	34±17.02
Sex male/female	7/8	35/29
Steroid therapy	1 (6.66%)	15 (20.27%)
Broad spectrum AB	15 (100.00%)	74 (100.00%)
Mortality	5 (33.33%)	30 (40.54%)

Table 4. Frequency of isolation of *Candida* spp. from 89 cancer patients with candidaemia.

<i>Candida</i> spp.	Number (%)
<i>Candida albicans</i>	15 (16.9)
<i>C. tropicalis</i>	29 (32.6)
<i>C. parapsilosis</i>	21 (23.6)
<i>C. rugosa</i>	19 (21.4)
<i>C. glabrata</i>	5 (5.6)
Total	89 (100.00)

Considering the etiology of candidaemia, *Candida albicans* was the cause in 15/89 patients (16.9%) in contrast to non-*albicans* *Candida* species in 74/89 patients (83.1%). Of the non-*albicans* species, *Candida tropicalis* was responsible for the maximum number of episodes (29/89, 32.6%), followed by *Candida parapsilosis* (21/89, 23.6%), *Candida rugosa* (19/89, 21.4%) and *Candida glabrata* (5/89, 5.6%) (Table 4). The

crude mortality in patients suffering from non-*albicans* and *albicans* candidaemia was 40.54 and 33.33%, respectively, the difference was not statistically significant. Distribution of *Candida* spp. in relation to cancer type is shown in Table 5.

As regard the results of real time-PCR, it was found that out of 89 blood samples which were positive for *Candida* spp. By conventional culture method 81 (91.01%) were positive for *Candida* spp. by real time-PCR and out of 111 negative blood samples by conventional culture methods 96 blood samples were negative for *candida* spp. by real time-PCR. Table 6 shows the sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) of real time-PCR when compared with conventional culture methods.

DISCUSSION

This study described bloodstream infections caused by *Candida* spp. in cancer patients using conventional methods

Table 5. Distribution of *Candida* spp. in relation to cancer type.

Diagnosis	<i>C. albicans</i>	<i>C. tropicalis</i>	<i>C. parapsilosis</i>	<i>C. rugosa</i>	<i>C. glabrata</i>
GIT	4 (4.49%)	7(7.86%)	5(5.61%)	3(3.37%)	1(1.12%)
breast	3 (3.37%)	7(7.86%)	2 (2.24%)	6(6.74%)	2(2.24%)
lung	1 (1.12%)	5 (5.61%)	3(3.37%)	4(4.49%)	-(00.00%)
Head and neck	6 (6.74%)	7(7.86%)	7(7.86%)	4(4.49%)	2(2.24%)
Others	1(1.12%)	3(3.37%)	4(4.49%)	2(2.24%)	-(00.00%)
Total 89 (100.00%)	15(16.9%)	29(32.6%)	21(23.6%)	19(21.4%)	55 (5.6%)

Table 6. Results of conventional culture and real time-PCR.

Sample	*BC results for <i>Candida</i> spp.	PCR +ve	PCR -ve	Total	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)
Blood (n=200)	Positive (89)	81	8	89	91.01	95.6	94.18	92.30
	Negative (111)	5	96	111				

*BC: Blood culture.

and real time PCR to provide information on the epidemiology of candidaemia in this patient population by conventional culture methods; the results of this study showed that *Candida* spp. could be isolated from 89 (44.5%) cases out of 200 cases of blood stream infection from oncology unit in Tanta university hospital in a period of 10 months (from March 2012 to March 2012). These results are different from the results of Borzotta and Beardsley (1999) that reported the rate of candidaemia in cancer patients to be 1.5%. This difference may be attributed to the difference of the regimen of chemotherapy or the rules of infection controls in each institution.

In this study, non-*albicans Candida* species accounted for the majority of the episodes of candidaemia in 74/89 patients (83.1%). Previous studies have shown high rates of carriage of these species on the hands of healthcare workers (Rangel-Frausto et al., 2006). Thus, the hands of healthcare workers could be one possible source of transmission; these results are in agreement with the two studies of Chakrabarti et al. (2009) and Xess et al. (2007), respectively that could isolate non-*Candidal* strains from 75 and 66% population of patients. In the present study, *C. tropicalis* was isolated from 29 (32.6%) cases, these results came in agreement with the results of Pfaller et al. (2010), who reported that worldwide, the frequency of bloodstream infections due to *C. tropicalis* has increased. In India, *C. tropicalis* ranks first among non-*albicans Candida* species in causing candidaemia (Chakrabarti et al., 2009; Xess et al., 2007).

In this study, *C. glabrata* could be isolated from 5 (5.6%) cases and *C. krusei* was not detected in the isolated strains, the results are in accordance with that of Vollmer et al (2008) that could isolate only one strain of *C. krusei* from different clinical specimens in immunocompromised patients.

In the present study, *C. tropicalis* and *C. parapsilosis* were the two most commonly isolated species from cases of candidaemia. Thus, the species distribution in this study closely matched that of previous studies from North India (Xess et al., 2007). However, points in which this study differed from others were the lower isolation rate of *C. glabrata* (5.6%) and the absence of any case of candidaemia due to *C. krusei*. These findings probably reflect the absence of selective pressure exerted by azole antifungal prophylaxis at our institutions, which would have led to isolation of species that are known to be inherently resistant to fluconazole such as *C. krusei*.

In the present study, *C. rugosa*, could be isolated from a large number of cases of fungaemia (19/89, 21.4%), the result which is in agreement with that of Minceş et al. (2009) who isolated *C. rugosa* from 25% of cases. Minceş et al. (2009) also was in agreement with our study in that blood stream infections with *Candida* spp. are especially common among patients in Intensive Care Units and associated with typical risk factors such as indwelling central vascular catheters, broad-spectrum antibiotics and steroid therapy. These results are in agreement with that of Colombo et al. (2006) who reported a large number of episodes of fungaemia due to *C. rugosa*. In addition, an outbreak of fungaemia due to *C. rugosa* was reported by Colombo et al. (2003).

Real-time PCR, is an approach that enables culture-independent screening of *Candida* infections within only a few hours. Rapid diagnosis would improve the survival rate in cancer patients. In this study, real-time PCR facilitated a rapid screening for simultaneous differentiation of the isolated 5 medically important *Candida* species causing blood stream infection in cancer patients in the same LightCycler run.

In the present study, real-time PCR showed mostly high

sensitivity. In some samples, PCR detected a greater number of different *Candida* species than culture. In these cases, patients had received antifungal therapy before sampling, which may be a reason for these discrepancies. Furthermore, whenever *C. albicans* was detected in addition to the cultured *Candida* (*C. glabrata* and *C. tropicalis*) species, *C. albicans* might have been overgrown by the other fungi and therefore missed by culture. On the other hand, culture detected a greater number of different species than real-time PCR in five samples, which have been already observed in previous studies (Willinger et al., 2003)

The present study revealed that blood culture is considered to be an important tool for the detection of systemic infection but it is time consuming and may lead to increase in the mortality of cancer patients with candidaemia. This is in contrast with Einsele et al. (1997) and Pfaller et al. (2010) who reported that the culture was positive in less than 50% of patients with disseminated candidiasis.

Conclusion

The high prevalence of non-*albicans* candidaemia should be considered prior to antimicrobial therapy in non-neutropenic cancer patients with candidaemia. Real-time PCR represents a rapid and accurate method for diagnosis of candidaemia.

ACKNOWLEDGEMENTS

The authors greatly acknowledge Medical Microbiology and Immunology, Tropical Medicine, Internal medicine and Clinical Oncology Departments, Faculty of Medicine, Tanta University for their great support and co-operation.

REFERENCES

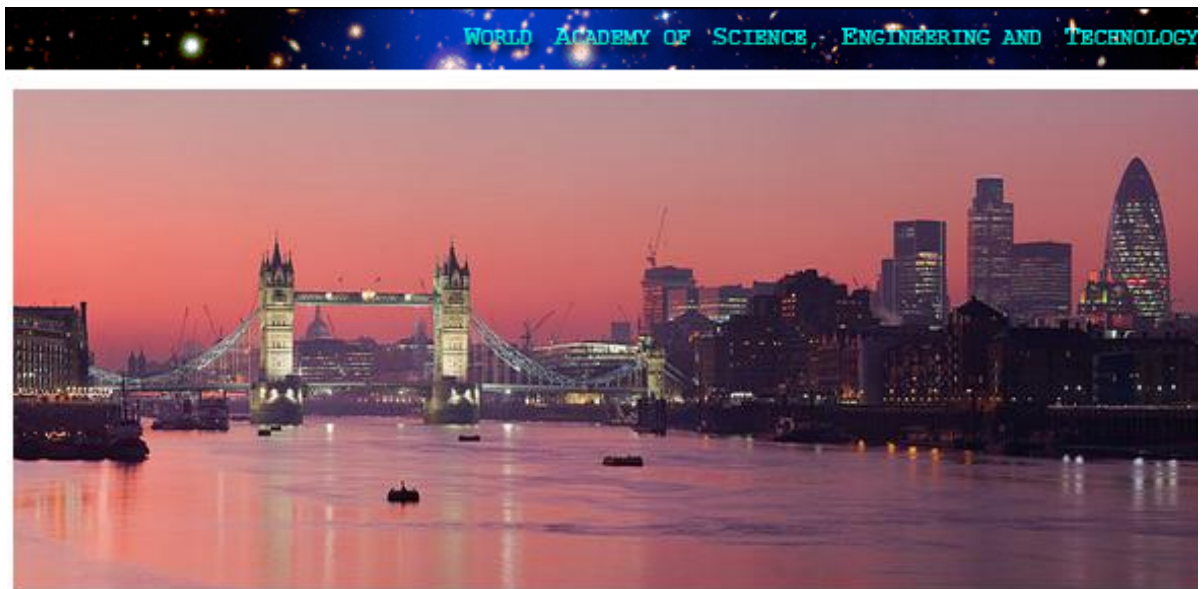
- Borzotta AP, Beardsley K (1999). *Candida* infections in critically ill trauma patients. *Arch. Surg.* 134:657–665
- Boyanton BL, Luna RA, Fasciano KG, Menne J (2008). DNA pyrosequencing-based identification of pathogenic *Candida* species by using the internal transcribed spacer 2 region. *Arch. Pathol. Lab. Med.* 132:667–74
- Chakrabarti A, Chatterjee SS, Rao KL, Zameer MM, Shivaprakash MR, Singhi S (2009). Recent experience with fungaemia: change in species distribution and azole resistance. *Scand. J. Infect. Dis.* 41: 275–284.
- Chakrabarti A, Chatterjee SS, Shivaprakash MR. (2008). Overview of opportunistic fungal infections in India. *Nippon Ishinkin Gakkai Zasshi.* 49:165–72.
- Chen YC, Eisner JD, Kattar MM, Rassouljian-Barrett SL, Lafe K, Yarfitz SL (2000). Identification of medically important yeasts using PCR-based detection of DNA sequence polymorphisms in the internal transcribed spacer 2 region of rRNA genes. *J. Clin. Microbiol.* 38:2302–10
- Chung YL, Emily CD, Heng FS, Rozita R, Kee PN, Pei PC (2005). Increased expression and hot spot mutations of the multidrug efflux transporter, CDR1 in azole-resistant *Candida albicans* isolates from vaginitis patients. *FEMS Microbiol. Lett.* 249:283–289
- Cirak YM, Kalkanci A, Kustimur S (2003). Use of molecular methods in identification of *Candida* species and evaluation of fluconazole resistance. *Mem Inst Oswaldo Cruz, Rio de Janeiro* 98:1027–32.
- Colombo AL, Melo AS, Crespo Rosas RF, Salomão R (2003). Outbreak of *Candida rugosa* candidemia: an emerging pathogen that may be refractory to amphotericin B therapy. *Diagn. Microbiol. Infect. Dis.* 46: 253–257.
- Colombo AL, Nucci M, Park BL, Nouér SA, Arthington-Skaggs B; for the Brazilian Network Candidemia Study (2006). Epidemiology of candidemia in Brazil: a nationwide sentinel surveillance of candidemia in eleven medical centers. *J. Clin. Microbiol.* 44:2816–2823.
- Einsele H, Hebart G, Roller J, Löffler I, Rothenhofer CA (1997). Detection and identification of fungal pathogens in blood by using molecular probes. *J. Clin. Microbiol.* 35:1353–1360.
- Forbes A, Danine F, Alice S Weissfeld. Bailey & Scott's (2002). *diagnostic Microbiology*. 12th edition. Elsevier Health Sciences.
- Halliday C, Hoile R, Sorrell T, James G, Yadav S, Shaw P (2005). Role of prospective screening of blood for invasive aspergillosis by polymerase chain reaction in febrile neutropenic recipients of haematopoietic stem cell transplants and patients with acute leukaemia. *Br. J. Haematol.* 132:478–86
- Hebart H, Löffler J, Reitze H, Engel A, Schumacher U, Klingebiel T (2000). Prospective screening by a panfungal polymerase chain reaction assay in patients at risk for fungal infections: Implications for the management of febrile neutropenia. *Br. J. Haematol.* 111:635–40
- Loeffler JN, Henke H, Hebart D, Schmidt L, Hagemeyer U, Einsele H (2000). Quantification of fungal DNA by using fluorescence resonance energy transfer and the LightCycler system. *J. Clin. Microbiol.* 38:586–590.
- Minces LR, Veldkamp PJ, Clancy CJ (2009). *Candida rugosa*: a distinctive emerging cause of candidaemia. A case report and review of the literature. *Scand. J. Infect. Dis.* 41:892–897
- Naglik J, Albrecht A, Bader O, Hube B (2004). *Candida albicans* proteinases and host/pathogen interactions. *Cell Microbiol.* 6:915–926
- Pfaller MA, Diekema DJ, Gibbs DL (2010). Results from the ARTEMIS DISK Global Antifungal Surveillance Study, 1997–2007: a 10.5-year analysis of susceptibilities of *Candida* species to fluconazole and voriconazole as determined by CLSI standardized disk diffusion. *J. Clin. Microbiol.* 48:1366–1377
- Rangel-Frausto, MS, Wiblin T, Blumberg TF (2006). National Epidemiology of Mycoses Survey (NEMIS): variations in rates of bloodstream infections due to *Candida* species in seven surgical intensive care units and six neonatal intensive care units. *Clin. Infect. Dis.* 29:253–258
- Verma AK, Prasad KN, Singh M, Dixit AK, Ayyagari A (2003). Candidemia in patients of a tertiary health care hospital from north India. *Ind. J. Med. Res.* 117:122–8
- Verweij PE, Meis JF (2000). Microbiological diagnosis of invasive fungal infections in transplant recipients. *Transplant Infect. Dis.* 2:80–87
- Vollmer T, Stormer M, Kleesiek K, Dreier J (2008). Evaluation of novel broad-range real-time PCR assay for rapid detection of human pathogenic fungi in various clinical specimens. *J. Clin. Microbiol.* 46: 1919–1926
- Willinger BA, Obradovic B, Selitsch J, Beck-Mannagetta W, Buzina H (2003). Detection and identification of fungi from fungus balls of the maxillary sinus by molecular techniques. *J. Clin. Microbiol.* 41:581–585.
- Xess I, Jain N, Has F (2007). Epidemiology of candidemia in a tertiary care centre of north India: 5-year study. *Infection* 35: 256–259.
- Xu J, Ramos AR, Vigalys R, Mitchell TG (2000). Clonal and spontaneous origins of fluconazole resistance in *Candida albicans*. *J. Clin. Microbiol.* 38:1214–20.

UPCOMING CONFERENCES

6th Trends in Medical Mycology Meeting, Copenhagen, Denmark, 11 Oct 2013



International Conference on Biochemistry and Pharmaceutical Sciences, London, UK, 19 Jan 2014



ICBPS 2014 : International Conference on Biochemistry and Pharmaceutical Sciences

**GB, London
January 20-21, 2014**

Conferences and Advert

October 2013

XXI Latin American Congress of Parasitology, Guayaquil, Ecuador, 6 Oct 2013

6th Trends in Medical Mycology Meeting, Copenhagen, Denmark, 11 Oct 2013

December 2013

International Conference on Molecular Virology and Microbiology, Bangkok, Thailand, 24 Dec 2013

International Conference on Cellular and Molecular Biology, Phuket, Thailand, 25 Dec 2014

January 2014

International Conference on Biochemistry and Pharmaceutical Sciences, London, UK, 19 Jan 2014

African Journal of Microbiology Research

Related Journals Published by Academic Journals

- *African Journal of Biotechnology*
- *African Journal of Biochemistry Research*
- *Journal of Bacteriology Research*
- *Journal of Evolutionary Biology Research*
- *Journal of Yeast and Fungal Research*
- *Journal of Brewing and Distilling*

academicJournals